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ORIGINAL RESEARCH

Transcription profiling of guanine nucleotide binding proteins during developmental regulation and pesticide response in *Solenopsis invicta* (Hymenoptera: Formicidae)

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Biological Control of Pests Research Unit, National Biological Control Laboratory, Mid South Area, Agricultural Research Service, United States Department of Agriculture, Stoneville, MS, USA **Abstract:** Guanine nucleotide binding proteins (GNBP), glycoproteins anchored on the cytoplasmic cell membrane, are mediators for many cellular processes. Complete cDNA of the GNBP gene β -subunit (*SiGNBP*) was cloned and sequenced from *Solenopsis invicta* Buren (Hymenoptera: Formicidae) worker ants. To understand whether *SiGNBP* is developmentally regulated in *S. invicta*, its expression levels in different developmental stages were examined using quantitative real-time polymerase chain reaction. *SiGNBP* was expressed in each developmental stage, and was especially highly expressed in the late larval and early pupal stages, as well among the dealate females (queens) ~10 days post nuptial flight. Quantitative real-time polymerase chain reaction also showed that mRNA transcription levels of *SiGNBP* in *S. invicta* workers were regulated during the duration of the study in response to heat shock, ultraviolet light, and boric acid treatment. These results suggest that the *SiGNBP* gene may play an important role in the development of *S. invicta*, and this has potential for use as a target in new insecticides for the control of fire ants.

Keywords: *Solenopsis invicta*, guanine nucleotide binding proteins, development, boric acid, mRNA transcription

Introduction

Transcriptional profiling is a powerful method for investigating the biological effects of chemicals and other stressors. It also offers a significant approach to identifying the relationship of gene expression to the development of the organisms. Information generated in such an approach is useful in identifying biomarkers for environmental and chemical stresses; this information can also be used in assessing new targets for drug development, such as in new insecticides for pest insect control. Guanine nucleotide binding proteins (GNBP), known as GTP-binding proteins and GTPases, are glycoproteins anchored on the cytoplasmic cell membrane, and are mediators for many cellular processes including signal transduction, protein transport, growth regulation, and polypeptide chain elongation. G- β subunits from heterotrimeric GNBPs modulate a wide array of signaling cascades by binding directly to diverse proteins, including effectors and regulators. GNBPs have been found in a variety of animals, plants, fungi, and insects.¹⁻⁶ The evolutionary aspects of GNBP-coupled receptors and their signaling pathways have revealed certain insect-specific features,⁷ indicating that the GNBP may be a good target for developing new specific insecticides.

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http://dx.doi.org/10.2147/OAIP.S35473

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The red imported fire ant is a widespread pest ant with significant medical and economic importance.^{8,9} As in all organisms, the development of ants requires multigene regulation, which is affected by environmental conditions such as temperature, radiation, and challenges from pesticides and parasites.^{10–22} However, the role of GNBPs in *Solenopsis invicta* Buren development has never been examined. In this study, the complete cDNA of the guanine nucleotide-binding protein gene β -subunit (*SiGNBP*) was cloned and sequenced from *S. invicta* workers, and its transcriptional expression related to the developmental stages and pesticide response was examined.

Materials and methods Fire ants

Colonies of S. invicta were collected in Washington County, Mississippi in 2010 and 2011. All developmental stages of S. invicta, (ie, eggs, larvae, pupae, and adults - including workers, male alates, female alates, and queens) were collected at numerous time points within each stage in the laboratory. Larval stages were morphologically identified by microscopy according to the criteria of Petralia and Vinson.²³ For the egg stage, 50 μ g of three samples were collected. For the larval stage, 50-100 larvae per sample were collected for RNA extraction. For pupae and adults, each sample contained 5-20 pupae and adults. Small workers and large workers were separated according to their sizes. An average of 20 large ants weighed 74.6 ± 1.5 mg, while an average of 20 small ants weighed 15.6 ± 2.19 mg. Three different colonies from Washington County, Mississippi in 2010 were used. Reproductive larvae were separated from the colony and were maintained with workers together in growth chambers in order to collect the time course samples for developmental study. Samples were frozen at -80°C in a freezer, and RNA extractions were processed when the collection stage was completed. Newly dealated queens were collected in Leland, Mississippi and were used for the time course studies of queens. Three queens were collected in 1, 5, 10, 16, 21, and 32 days, individually. The queens were provided with 10% sugar as food in a 50 mL glass test tube with cottons plugs. After about 8-10 days, the queens produced their first set of eggs.

RNA extraction

Total RNAs were extracted using TRIzol reagent according to the manufacturer's instructions (Invitrogen, Life Technologies Corporation, Carlsbad, CA). Poly (A)⁺ RNA was isolated by applying Oligotex-dT suspension (QIAGEN, Valencia, CA). mRNA samples were quantified by Smart-Spec[™] Plus Spectrophotometry (Bio-Rad Laboratories, Inc, Hercules, CA).

GeneRacer cloning

The GeneRacer[™] kit was used to amplify the full-length gene of 5' and 3' cDNA ends by slightly modifying the manufacturer's instruction (Invitrogen). Polymerase chain reaction (PCR) products were cloned using the TOPO TA Cloning[®] kit for sequencing (Invitrogen). Transformed plasmids were inserted into One Shot[®] TOP10 Competent Cells (Invitrogen) and grown overnight on Luria-Bertani plates containing ampicillin and X-Gal (5-bromo-4chloro-3-indolyl- beta-D-galactopyranoside). Clones were isolated and grown overnight in LB-ampicillin broth at 37°C, and shaken at 230 RPM in the Thermo Scientific Analog MaxQ 4000 Shaker (Thermo Fisher Scientific Inc, Waltham, MA).

Gene sequencing of GeneRacer library

Clones of the GeneRacer library were purified with a QIAprep Miniprep (QIAGEN). The plasmid DNAs ($0.5 \mu g$) were then digested using the EcoRI enzyme (2.5 U) for 1.5 hours, and were run on a 1% agarose gel to confirm the DNA insert. Selected clones were then sent to the DNA Sequencing Core at the Interdisciplinary Center for Biotechnology Research at the University of Florida (Gainesville, FL) to be sequenced and analyzed using the National Center for Biotechnology Information BLASTN program to identify sequence homologies. The sequences were submitted into the National Center for Biotechnology Information GenBank; the accession number was HM130685.

cDNA synthesis for qPCR

To ensure that genomic DNA did not contaminate the sample, the Oligotex mRNA mini Kit (QIAGEN) was used to purify total RNA. A 250 ng aliquot of purified mRNA was reverse transcribed in a 20 μ L reaction volume using the Clone AMV first-Strand Synthesis Kit, while also using the Oligo (dT)20 primer for cDNA synthesis according to the manufacturer's instructions (Invitrogen). The reaction was terminated by heat inactivation at 95°C for 5 min. The cDNA samples used for the quantitative real-time PCR (qPCR) analysis were taken from ants in their developmental stages undergoing boric acid treatment; controls were diluted by adding 80 μ L ddH₂O (~450 ± 50 ng/ μ L) and stored at -20°C.

To design gene-specific primers, detailed analyses of the nucleotide sequence of genes found in the library (Genbank accession number: HM130685) were performed using the PRIMER3-Design Primer Pairs and Probes program from Biology Workbench (Department of Bioengineering, University of California, San Diego, CA). The primers for the *S. invicta* actin gene (Genbank accession number: HM130684) were also designed for internal control and comparison purposes.

qPCR amplification

The qPCR assay for the SiGNBP gene in S. invicta was performed using Platinum® SYBR® Green qPCR Super-Mix-UDG with ROX (Invitrogen) in a volume of 15 µL on an Applied Biosystems 7300 Fast Real-Time PCR System (Life Technologies Corporation, Foster City, CA). The PCR mixture consisted of 1 µL of diluted cDNA (~450 \pm 50 ng/µL), 0.5 µM primers, and 1X master mix. In every qPCR run, actin was used as an internal control to normalize for variation in the amount of the cDNA template. The PCR primers for the actin gene were SiActin-783-F 5'-CCTCTTCCAACCTTCCTTCC-3', SiActin-948R 5'-CTTTTGCATACGATCAGCGA-3', SiGNBP-13F 5'-TTACAGCTGAGAGGGACGCT-3', and SiGNBP-267R 5'-AAGACGCAATGTTTTGTCCC-3'. The PCR thermal cycling parameters were the same as described previously.^{22,24} Relative expression levels were calculated as follows. First, SiGNBP transcript levels relative to a standard (SiActin) were calculated using the following formula:

$$\Delta CT = CT (SiGNBP) - CT (SiActin)$$
(1)

Second, an average ΔCT value for each sample was calculated. Third, relative expression levels were calculated using the equation:

$$100 \times 2$$
 [-average ΔCT] (2)

Boric acid experiments

Boric acid was delivered in a 10% sugar solution using feeding stations at room temperature (27°C). Two concentrations were used: 0.85 mg/mL (low dose, LD) and 8.5 mg/mL (high dose, HD). To trace the RNA expression levels of differentially transcribed genes for each dose, workers (200 mg) were sampled from each of the three different colonies at 4, 8, 12, and 14 days. Control workers were simultaneously collected from colonies that were supplied with boric acid-free sugar water. This time course study was replicated three times. mRNA was extracted and a cDNA library was created as previously described. qPCR was performed using primers designed for the genes *SiGNBP* and *SiActin*.

Sequence data processing

Multiple sequence alignments of GNBP and orthologues from other insects were performed with the Molecular Evolutionary Genetics Analysis 5.05 program (MEGA; http://www.megasoftware.net). A phylogenetic tree was constructed using the neighbor-joining method with the MEGA 5.05 program.²⁵ The neighbor-joining method is based on the minimum-evolution criterion, and is also a bottom-up clustering method for the creation of phenetic trees.²⁶

Statistical analysis

Comparisons of means were analyzed using the paired *t*-test, and *t*-values and *P*-values were reported when normality and equal variance tests were passed. Significant differences between the data were determined using SigmaPlot software (SigmaPlot[®]11.2; Systat Software, Inc, San Jose, CA).

Results

Identification of Solenopsis invicta guanine nucleotide-binding protein subunit beta-like protein gene

The complete cDNA sequence of the *SiGNBP* gene of *S. invicta* was deposited in the GenBank (accession number: HM130685). *SiGNBP* is 951 base pairs in length, and encodes a protein of 317 amino acids with a molecular mass of 35.7 kDa. A phylogenetic tree for *SiGNBP* nucleic acid sequences of guanine nucleotide binding proteins from other insect orthologues was constructed using the neighborjoining method with the MEGA 5.05 program (Figure 1). The phylogenetic analysis showed that *SiGNBP* was closely related to guanine nucleotide binding proteins from *Apis mellifera* L. and *Bombus terrestris* L.

SiGNBP RNA profile in different developmental stages of Solenopsis invicta

To understand how *SiGNBP* is regulated during the development of *S. invicta*, qPCR was performed to examine the relative transcription levels of *SiGNBP* in eggs, larvae, pupae, workers, male alates, female alates, and queens (Table 1A and B). In the egg stage, the relative RNA transcription level of *SiGNBP* ranged from 13.217 ± 1.665 to 58.919 ± 0.935 .



Figure I Construct/test neighbor-joining GNBP tree.

Notes: A phylogenetic tree was constructed using the neighbor-joining method for SiGNBP nucleic acid sequences of GNBPs from other insect orthologues using the MEGA 5.05 program. The scale bar indicates the number of changes inferred as having occurred along each branch. The accession numbers of reference nucleotide sequences used in this analysis are: HM130685, XM_003394751, XM_392962, XM_001600102, DQ85470.1, EF070464.1, JF417987.1, AF368031.1, AF368031.1, AY737531.1, HM449904.1, DQ073455.1, AY588074.1, XM_001988797.1, XM_968486.2, XM_001948034.2, HQ638203.1, XM_002065121.1, EU259815.1, BX063203, and XM_002003720.1. Abbreviations: GNBP, guanine nucleotide binding proteins; SiGNBP, guanine nucleotide binding proteins gene β -subunit.

During the early worker larval stages, *SiGNBP* RNA expression decreased from first instar (37.288 ± 1.300) to third instar (4.264 ± 0.037), but these levels significantly increased during the fourth instar (111.71 ± 1.417) as compared to the third instar (Table 1A, and supplementary Table S1).

For reproductive larvae, *SiGNBP* RNA expression increased significantly in the fourth instar as compared to all the early instars, reaching (35.525 ± 0.425) (Table 1B and supplementary Table S1). There was a high *SiGNBP* RNA expression in early reproductive pupae (144.053 ± 1.997); however, expression significantly decreased in the late reproductive pupae. The *SiGNBP* RNA expression level was similar in the younger small workers, larger workers, and female alates (Table 1A and B).

In the male alates and queens, the relative *SiGNBP* RNA levels were differentially expressed from teneral male alates (8.249 \pm 0.395) to the mature male alates (95.703 \pm 4.111) (Table 1B and supplementary Table S1). The relative *SiGNBP* RNA levels were differentially expressed in queens' postnuptial flight. About 10 days postnuptial flight, the relative *SiGNBP* RNA expression reached its highest expression level (991.56 \pm 15.46), which is significantly increased compared with day 1 (4.855 \pm 1.842) and day 5 (57.784 \pm 0.821) (Table 1B and supplementary Table S1). However, the relative *SiGNBP* RNA expression level was significantly reduced among the queens at 16 days postnuptial flight, as compared with the queens at day 10 postnuptial flight (Table 1B and supplementary Table S1).

Effects of boric acid on SiGNBP gene expression

To determine whether the transcription of SiGNBP in S. invicta was affected by boric acid treatment, workers were treated with different concentrations of boric acid in a 10% sugar solution. The qPCR time courses of SiGNBP expressed in workers were different between the two concentrations of the boric acid (HD and LD). In workers, relative SiGNBP expression levels increased two-fold for LD at 4 days of boric acid treatment, and then slightly increased for LD at 8-day and 12-day boric acid treatments compared with controls (Table 2A and B, Figure 2, and supplementary Table S2). After 14 days, SiGNBP expression was slightly decreased for LD boric acid when treatment compared with the control treatment of 10% sugar alone. However, boric acid HD-treated workers showed a decrease in SiGNBP RNA expression at 4, 8, 12, and 14-day exposures. Boric acid HD-treated workers showed a decrease in SiGNBP RNA expression (Figure 2, Table 2A and B, and supplementary Table S2).

Discussion SiGNBP gene expression during Solenopsis invicta development

We examined and analyzed changes in *SiGNBP* RNA expression in *S. invicta* eggs, larvae, pupae, and adults. The *SiGNBP* gene was expressed in varying quantities during the egg and early instar stages. Previous studies showed that different forms of GNBP-coupled sensory transduction may mediate developmental interactions during both early and late stages of *Drosophila* embryogenesis.⁶ Guanine nucleotide-binding proteins mediate signals between serotonin receptors and adenylate cyclase in *Schistosoma mansoni*, which indicates that the expression of this gene is developmentally mediated.¹⁰

There were significant differences in the expression of the *SiGNBP* gene between early (first to third instars) and late larval stages of worker ants (fourth instar). In addition, the expression of the *SiGNBP* gene in the early pupal stage was significantly higher than that in the late pupal stages of both worker and reproductive ants. These changes may indicate that the expression of the *SiGNBP* gene is important for late larval and pupal development in *S. invicta*. The fourth instar larvae play a critical role in the survival of the colony; they digest solid foods and provide protein and other nutrients to the other members in the colony.¹⁵ An ant colony may be sensitive toward any disruption to the fourth instar larvae, which may suggest that *SiGNBP* is important to fourth instar larvae

Sample	Sample	Mean Ct \pm SD		Relative S	GIGNBP expres	sion level	
stage	name	SiActin	SiGNBP	∆Ct-I	∆Ct-2	∆Ct-3	$100 \times 2^{-\Delta Ct} \pm SD$
Egg	Egg I	21.325 ± 0.157	24.245 ± 0.024	2.738	3.101	2.920	13.217 ± 1.665
	Egg 2	$\textbf{16.488} \pm 0.004$	17.251 ± 0.026	0.786	0.740	0.763	$\textbf{58.919} \pm \textbf{0.935}$
	Egg 3	18.191 ± 0.086	18.978 ± 0.040	0.833	0.741	0.787	57.951 ± 1.848
	Egg 4	19.839 ± 0.308	$\textbf{21.549} \pm \textbf{0.058}$	1.651	1.767	1.709	$\textbf{30.582} \pm \textbf{1.233}$
Larvae	LI	18.733 ± 0.010	$\textbf{20.157} \pm \textbf{0.041}$	1.474	1.373	1.423	$\textbf{37.288} \pm \textbf{1.300}$
	L2	18.518 ± 0.002	21.172 ± 0.015	2.666	2.641	2.654	15.893 ± 0.140
	L3	18.040 ± 0.029	$\textbf{22.592} \pm \textbf{0.016}$	4.563	4.540	4.552	$\textbf{4.264} \pm \textbf{0.037}$
	L4	21.171 ± 0.007	$\textbf{21.012} \pm \textbf{0.017}$	-0.178	-0.141	-0.160	.7 ± .4 7
Pupae	PI	$\textbf{19.687} \pm \textbf{0.045}$	20.860 ± 0.071	1.148	1.198	1.173	$\textbf{44.353} \pm \textbf{0.781}$
	P2	17.163 ± 0.035	$\textbf{22.447} \pm \textbf{0.015}$	5.333	5.235	5.284	$\textbf{2.567} \pm \textbf{0.088}$
Small	SW	18.215 ± 0.193	$\textbf{19.448} \pm \textbf{0.021}$	1.060	1.405	1.233	$\textbf{42.558} \pm \textbf{5.098}$
workers	SW	17.278 ± 0.088	18.999 ± 0.029	1.604	1.838	1.721	30.335 ± 2.462
	SW	17.391 ± 0.152	$\textbf{19.374} \pm \textbf{0.010}$	1.831	2.134	1.983	25.305 ± 2.656
	SW	$\textbf{18.688} \pm \textbf{0.008}$	$\textbf{22.266} \pm \textbf{0.034}$	3.621	3.536	3.579	$\textbf{8.370} \pm \textbf{0.246}$
	SW	$\textbf{18.397} \pm \textbf{0.025}$	$\textbf{22.064} \pm \textbf{0.061}$	3.631	3.704	3.667	7.871 ± 0.198
	SW	18.028 ± 0.003	21.143 ± 0.012	5.022	4.911	4.966	$\textbf{3.199} \pm \textbf{0.124}$
Larger	LW	16.076 ± 0.024	17.710 ± 0.010	1.640	1.613	1.627	32.386 ± 0.299
workers	LW	16.672 ± 0.061	$\textbf{18.878} \pm \textbf{0.024}$	2.243	2.170	2.206	21.673 ± 0.549
	LW	$\textbf{18.029} \pm \textbf{0.023}$	20.991 ± 0.002	2.965	2.982	2.974	12.730 ± 0.281
	LW	18.028 ± 0.013	21.143 ± 0.012	3.124	3.107	3.116	11.538 ± 0.070
	LW	17.862 ± 0.036	21.620 ± 0.025	3.748	3.770	3.759	$\textbf{7.388} \pm \textbf{0.056}$
	LW	18.122 ± 0.016	23.018 ± 0.076	4.803	4.896	4.988	$\textbf{3.360} \pm \textbf{0.216}$

Table IA Expression of SiGNBP in different developmental stages of Solenopsis invicta

 $\label{eq:bilder} \textbf{Abbreviations:} Sign BP, \ guanine \ nucleotide \ binding \ proteins \ gene \ \beta-subunit; \ Ct, \ cycle \ threshold; \ SD, \ standard \ deviation.$

Sample	Sample	Mean $Ct \pm SD$		Relative S	GiGNBP expre	ssion level	
stage	name	SiActin	SiGNBP	∆Ct-I	∆Ct-2	∆Ct-3	$100 \times 2^{-\Delta Ct} \pm SD$
Reproductive	RLI	$\textbf{16.248} \pm \textbf{0.007}$	19.364 ± 0.019	3.129	3.116	3.103	11.537 ± 0.102
larvae	RL2	$\textbf{16.235} \pm \textbf{0.004}$	$\textbf{20.139} \pm \textbf{0.007}$	3.915	3.894	3.904	$\textbf{6.678} \pm \textbf{0.050}$
	RL3	15.125 ± 0.011	18.148 ± 0.036	2.998	3.048	3.023	12.304 ± 0.263
	RL4	16.083 ± 0.022	17.560 ± 0.039	1.459	1.493	1.476	35.525 ± 0.425
Reproductive	RPI	19.147 ± 0.007	18.621 ± 0.027	-0.507	-0.547	-0.527	144.05 ± 1.997
pupae	RP2	19.623 ± 0.046	21.610 ± 0.075	1.959	2.015	1.987	25.225 ± 0.495
Female alates	FA	17.150 ± 0.022	$\textbf{18.354} \pm \textbf{0.028}$	1.199	1.210	1.205	$\textbf{43.386} \pm \textbf{0.162}$
	FA	19.219 ± 0.003	$\textbf{21.012} \pm \textbf{0.039}$	1.738	I.847	1.793	28.860 ± 1.091
	FA	18.015 ± 0.003	$\textbf{21.136} \pm \textbf{0.029}$	3.153	3.089	3.121	11.493 ± 0.253
Male alates	MAIª	18.714 ± 0.023	$\textbf{22.314} \pm \textbf{0.046}$	3.531	3.669	3.599	$\textbf{8.249} \pm \textbf{0.395}$
	MA4	16.753 ± 0.036	$\textbf{21.648} \pm \textbf{0.113}$	4.971	4.817	4.894	$\textbf{3.362} \pm \textbf{0.178}$
	MA8	19.733 ± 0.012	$\textbf{22.962} \pm \textbf{0.005}$	3.212	3.246	3.229	10.666 ± 0.124
	MA12	$\textbf{18.478} \pm \textbf{0.026}$	$\textbf{22.075} \pm \textbf{0.171}$	3.452	3.742	3.597	$\textbf{8.265} \pm \textbf{0.832}$
	MA14	$\textbf{24.174} \pm \textbf{0.056}$	$\textbf{24.237} \pm \textbf{0.118}$	0.001	0.125	0.063	95.703 ± 4.111
	MA18	21.121 ± 0.492	$\textbf{23.547} \pm \textbf{0.121}$	2.205	2.426	2.630	18.817 ± 2.772
Queens	QI⁵	18.193 ± 0.079	22.307 ± 0.071	4.105	4.123	4.114	5.775 ± 0.035
	Q5	$\textbf{20.659} \pm \textbf{0.005}$	21.451 ± 0.003	0.789	0.793	0.791	$\textbf{57.784} \pm \textbf{0.821}$
	Q10	$\textbf{26.346} \pm \textbf{0.058}$	$\textbf{23.037} \pm \textbf{0.036}$	-3.332	-3.287	-3.310	991.56 ± 15.46
	Q16	$\textbf{19.137} \pm \textbf{0.040}$	$\textbf{22.762} \pm \textbf{0.065}$	3.649	3.601	3.625	$\textbf{8.106} \pm \textbf{0.136}$
	Q21	$\textbf{21.078} \pm \textbf{0.030}$	$\textbf{22.177} \pm \textbf{0.011}$	1.080	1.118	1.099	$\textbf{46.679} \pm \textbf{0.607}$
	Q32	19.205 ± 0.038	$\textbf{20.969} \pm \textbf{0.142}$	1.868	1.660	1.764	29.445 ± 2.123

Table IB Expression of SiGNBP in different developmental stages of Solenopsis invicta

Notes: ^aMale alates day post emerging from reproductive pupae; ^bdealate female (queens) day post nuptial flight.

 $\label{eq:abbreviations: SiGNBP, guanine nucleotide binding proteins gene β-subunit; Ct, cycle threshold; SD, standard deviation.$

 Table 2A Expression of SiGNBP under boric acid feeding conditions in Solenopsis invicta

Time point	Ct ± SD		Relative SiGNBP expression level			
	SiActin	SiGNBP	∆ Ct-I	∆Ct-2	∆Ct-3	
Cont-4d'ª	19.023 ± 0.121	21.842 ± 0.041	2.656	2.981	2.818	
Cont-8dª	18.029 ± 0.023	20.991 ± 0.002	2.965	2.982	2.974	
Cont-12dª	$\textbf{18.028} \pm \textbf{0.013}$	21.143 ± 0.012	3.124	3.107	3.116	
Cont-14ª	17.770 ± 0.083	20.727 ± 0.052	2.988	2.927	2.957	
BA-4d⁵	19.230 ± 0.025	22.114 ± 0.011	2.848	2.884	2.920	
BF-8d⁵	18.156 ± 0.044	$\textbf{21.282} \pm \textbf{0.037}$	3.133	3.120	3.126	
BF-12d ^b	18.760 ± 0.116	$\textbf{22.282} \pm \textbf{0.037}$	3.968	3.841	3.905	
BF-14d⁵	19.266 ± 0.011	23.766 ± 0.123	4.365	4.634	4.499	

Notes: ^aNonboric acid, only 10% sugar feeding in *Solenopsis invicta* workers; ^bboric acid (0.85 mg/mL) within 10% sugar feeding in *Solenopsis invicta* workers. **Abbreviations:** SiGNBP, guanine nucleotide binding proteins gene β-subunit; Ct, cycle threshold.

Abbreviations: Signor, guanine nucleolide binding proteins gene p-subunit; Ct, cycle urresno

and thus is an excellent target for developing intervening methods, such as a new insecticide.

Numerous physiological changes occur during the development of ant adults. A previous study showed that arginine kinase, a primary enzyme in cell metabolism and adenosine 5'-triphosphate-consuming processes, was differentially expressed among larvae and adults, queens and workers, and among female alates and queens.¹² The results showed that the *SiGNBP* gene is not only differentially expressed among eggs, larvae, adults, workers, female alates, and queens, but it is also differentially expressed within the larval (ie, first instar, second instar, third instar, and fourth instar), pupal (early and late pupae), as well as the worker (early and late workers) stages. The relatively low levels of *SiGNBP* expression in older workers and female alates suggest that the *SiGNBP* gene may play a role in the attenuation of gene expression.

SiGNBP gene responses to pesticide

Many insecticides or repellents, including bifenthrin, chlorfenapyr, fipronil, and thiamethoxam, have been evaluated for activity against the red imported fire ant, *S. invicta.*²⁸⁻³² Molecular studies of insecticide resistance, including identification of the genes involved in target site and metabolic resistance mechanisms, have advanced rapidly over the past decade.³²⁻³⁴ A previous study showed that boric acid significantly decreased the survivorship of workers in the target organs.³⁵

SiGNBP expression levels of worker *S. invicta* were upregulated at 4, 8, and 12 days, and were downregulated at 14 days after a low dose of boric acid treatment. However, *SiGNBP* expression levels were downregulated at 4, 8, 12, and 14 days after a high dose of boric acid treatment.

In conclusion, expression of the *SiGNBP* gene in the life cycle of *S. invicta* was highly regulated developmentally and environmentally. *SiGNBP* RNA expression has, for the first time, been examined in detail for all developmental stages of *S. invicta*. The current study suggests that *SiGNBP* plays an important role in the development of *S. invicta*, and it may provide useful information for designing novel strategies for fire ant control. Using RNA interference technology to knock down the *SiGNBP* gene may provide additional targets that can be developed as new pesticides.

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Time point	Ct ± SD		Relative SiGNBP expression level			
	SiActin	SiGNBP	∆Ct-I	∆Ct-2	∆Ct-3	
Cont-4d'a	19.023 ± 0.121	21.842 ± 0.041	2.656	2.981	2.818	
Cont-8d ^a	18.029 ± 0.023	20.991 ± 0.002	2.965	2.982	2.974	
Cont-12d ^a	$\textbf{18.028} \pm \textbf{0.013}$	21.143 ± 0.012	3.124	3.107	3.116	
Cont-14ª	17.770 ± 0.083	$\textbf{20.727} \pm \textbf{0.052}$	2.988	2.927	2.957	
BA-4d ^b	19.230 ± 0.025	22.114 ± 0.011	2.848	2.884	2.920	
BF-8d⁵	18.156 ± 0.044	$\textbf{21.282} \pm \textbf{0.037}$	3.133	3.120	3.126	
BF-12d ^b	18.760 ± 0.116	22.282 ± 0.037	3.968	3.841	3.905	
BF-14d ^b	19.266 ± 0.011	$\textbf{23.766} \pm \textbf{0.123}$	4.365	4.634	4.499	

Notes: ^aNonboric acid, only 10% sugar feeding in *Solenopsis invicta* workers; ^bboric acid (0.85 mg/mL) within 10% sugar feeding in *Solenopsis invicta* workers. **Abbreviations:** *SiGNBP*, guanine nucleotide binding proteins gene β-subunit; Ct, cycle threshold.



Figure 2 Boric acid treatment.

Notes: The SiGNBP transcription level in response to boric acid treatments, quantified by qPCR. S. invicta workers were exposed to low dose (0.85 mg/mL), and high dose (HD, 8.5 mg/mL) at 4, 8, 12, and 14 days.

Abbreviations: Cont, control; BA, boric acid; SiGNBP, guanine nucleotide binding proteins gene β -subunit; qPCR, quantitative real-time polymerase chain reaction.

Acknowledgments

We thank Drs. Zibiao Guo (University of Mississippi Medical Center) and Cesar Solorzano Torres (USDA-ARS) for critical reviews of the manuscript. We also thank Richard Evans and Sharla Burke (USDA-ARS) for technical support.

Disclosure

The authors report no conflict of interest in this work.

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Supplemental material

Table SIA Paired t-test data for comparison of relative SiGNBP gene
transcription between developmental stages in Solenopsis invicto

Stage and ages	N	Df	t-value	P-value
LI and L2	3	2	93.606	<0.001*
L2 and L3	3	2	38.995	<0.001*
L3 and L4	3	2	-55.610	<0.001*
L4 and PI	3	2	49.385	<0.001*
PI and P2	3	2	85.455	<0.001*
SW(Y) and SW(O)	3	2	13.107	=0.006*
LW(Y) and LW(O)	3	2	591.843	<0.001*

 pricta
 gene transcription between LD and HD boric acid treatments in

 Solenopsis invicta workers
 Solenopsis invicta workers

Boric acid treatments	N	df	t-value	P-value
LD and Cont 4d	3	2	-38.448	<0.001*
LD and Cont 8d	3	2	-32.053	<0.001*
LD and Cont 12d	3	2	-5.023	=0.037*
LD and Cont 14d	3	2	104.372	<0.001*
HD and Cont 4d	3	2	49.802	<0.001*
HD and Cont 8d	3	2	157.478	<0.001*
HD and Cont 12d	3	2	73.056	<0.001*
HD and Cont 14d	3	2	14.788	=0.005*

Table S2 Paired t-test data for comparison of relative SiGNBP

Note: *Statistical significance (*P* < 0.05).

Abbreviations: SiGNBP, guanine nucleotide binding proteins gene $\beta\mbox{-subunit;}$ LD, low dose; HD, high dose.

Note: *Statistical significance (P < 0.05).

Abbreviations: SiGNBP, guanine nucleotide binding proteins gene β -subunit; SW, small worker; LW, large worker.

Table SIB Paired t-test data for comparison of relative SiGNBP gene transcription between developmental stages in Solenopsis invicta

Stage and ages	Ν	Df	t-value	P-value
RLI and RL2	3	2	162.15	=0.001*
RL2 and RL3	3	2	144.81	<0.001*
RL3 and RL4	3	2	-100.45	<0.001*
RL4 and RPI	3	2	-77.33	<0.001*
RPI and RP2	3	2	36.792	<0.001*
FAI and FA2	3	2	27.166	=0.001*
FA2 and FA3	3	2	22.373	=0.002*
MA1 and MA4	3	2	14.762	=0.005*
MA4 and MA8	3	2	-41.739	<0.001*
MA8 and MA12	3	2	5.803	=0.028*
MAI2 and MAI4	3	2	-46.210	<0.001*
MA14 and MA18	3	2	38.975	<0.001*
Q1 and Q5	3	2	-1893.457	<0.001*
Q5 and Q10	3	2	-105.15	<0.001*
Q10 and Q16	3	2	111.133	<0.001*
Q16 and Q21	3	2	-89.963	<0.001*
Q21 and Q32	3	2	12.028	<0.007*

Note: *Statistical significance (P < 0.05).

Abbreviation: SiGBNP, guanine nucleotide binding proteins gene β -subunit.

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