Open Access Full Text Article

REVIEW

Chinese perspectives on clinical efficacy and safety of alectinib in patients with *ALK*-positive advanced non-small cell lung cancer

Hui Yu[1]
Si Sun[1]
Xingjiang Hu[2]
Jinjing Xia[3]
Jialei Wang[1]
Haiquan Chen[4]

[1]Department of Medical Oncology, Fudan University Shanghai Cancer Center, Shanghai, People's Republic of China; [2]Zhejiang Provincial Key Laboratory for Drug Evaluation and Clinical Research, First Affiliated Hospital, Zhejiang University, Hangzhou, Zhejiang, People's Republic of China; [3]Department of Medical Science Oncology, Shanghai Roche Pharmaceuticals Ltd., Shanghai, People's Republic of China; [4]Department of Thoracic Surgery, Fudan University Shanghai Cancer Center, Shanghai, People's Republic of China

Correspondence: Jialei Wang
Department of Medical Oncology, Fudan University Shanghai Cancer Center, 274 Dong'an Road, Xuhui District, Shanghai 200023, People's Republic of China
Tel +86 21 6417 5590
Fax +86 21 6417 0366
Email luwangjialei@126.com

Haiquan Chen
Department of Thoracic surgery, Fudan University Shanghai Cancer Center, 274 Dong'an Road, Xuhui District, Shanghai 200023, People's Republic of China
Tel +86 21 6417 5590
Fax +86 21 6417 0366
Email hqchen1@yahoo.com

Abstract: The incidence of lung cancer is increasing in China, in contrast to trends in Western countries, due to the increasing numbers of smokers and high levels of air pollution. Non-small-cell lung cancer (NSCLC) is the most common form of lung cancer, accounting for approximately 85% of lung cancers. Better understanding of the pathogenesis of NSCLC has led to the identification of multiple genetic mutations and chromosomal translocations such as those in the anaplastic lymphoma kinase (*ALK*) gene. To facilitate the identification of treatment targets, multiple guidelines (European Society for Medical Oncology, National Comprehensive Cancer Network, and American Society of Clinical Oncology) now recommend screening for genetic factors to help guide treatment decisions. In recent years, multiple ALK inhibitors have been developed to treat NSCLC, including the first-generation tyrosine kinase inhibitor (TKI) crizotinib; second-generation TKIs such as ceritinib, ensartinib, brigatinib, and alectinib; the third-generation TKI lorlatinib; and the fourth-generation TKI repotrectinib. These agents differ in structure, potency, and activity, both systemically and their effects on central nervous system (CNS) metastases. Recently, alectinib was approved in China to treat patients with locally advanced or metastatic NSCLC that were *ALK*+. Alectinib has demonstrated activity against NSCLC, including metastases within the CNS, with better tolerability than crizotinib. These ALK inhibitors represent significant advances in the treatment of NSCLC and yet patients will likely still exhibit disease progression. Alectinib offers greater potency with greater specificity as well as a better toxicity profile than many other TKIs that are currently available. Here, we review the role of *ALK* as a therapeutic target in NSCLC, the testing methods for identifying *ALK*-rearranged NSCLC, and the various TKIs currently being used or explored for treatment in this setting, with a focus on alectinib from a Chinese perspective.

Keywords: NSCLC, anaplastic lymphoma kinase, alectinib, tyrosine kinase inhibitor

Introduction

In China, the incidence of cancer is on the rise with lung cancer being the most common cancer type diagnosed and the most common cause of cancer-related deaths.[1–5] It is known that lung cancer morbidity and mortality trends reflect the prevalence of tobacco exposure 20–30 years earlier.[6,7] Therefore, due to the ongoing high levels of tobacco exposure and air pollution in China, both of which are significant risk factors in the development of lung cancer, the health burden of lung cancer is expected to increase in the future.[8–10]

Non-small-cell lung cancer (NSCLC) is the most common type of lung cancer, accounting for 85–90% of all lung cancer cases globally, and is associated with high rates of mortality.[1–5,11,12] According to the most recently released SEER Cancer Statistics Review, the 5-year survival rate (2009–2015) for NSCLC is 25.1%.[13] Improved understanding of the pathogenesis of NSCLC has resulted in the development of treatments that target various genetic mutations that drive the development and progression of NSCLC. This includes translocations of the anaplastic lymphoma kinase (*ALK*) gene[14] and the ROS1 proto-oncogene 1 receptor tyrosine kinase (*ROS1*) gene;[14] rearrangements in kinase-encoding genes such as epidermal growth factor receptor (*EGFR*),[14] Kirsten ras (*KRAS*),[14] v-raf murine sarcoma viral oncogene homolog B (*BRAF*),[14] neurotrophic tyrosine receptor kinase (*NTRK*),[14] and rearranged during transfection (*RET*);[14] fusions of neuregulin 1 (*NRG1*)[15] and fibroblast growth factor receptor 1/3 (*FGFR1/3*);[16] human epidermal growth factor receptor 2 (*HER2*) insertion;[16] and AKT serine/threonine kinase 1 (*AKT1*) mutation.[17] Some reports show that these mutations are mutually exclusive, while other reports show that some tumors can have concomitant mutations.[18–21] These mutations are capable of influencing responses to targeted therapy.[14] It has, therefore, become standard clinical practice to test for gene mutations and fusions in patients with NSCLC and to tailor treatment strategies accordingly.[22]

In the present review, we briefly discuss the role of *ALK* mutations and translocations in NSCLC, the testing methods for identifying *ALK*-positive (*ALK*+) NSCLC, and present the evidence for approved *ALK*-targeted therapies. This review focuses on alectinib in Chinese patients due to its recent and rapid approval (August 2018) as a monotherapy for locally advanced or metastatic *ALK*+ NSCLC in China.

Mechanism of action of ALK

The *ALK* gene is a member of the insulin receptor superfamily. It is located on the short arm of chromosome 2 (2p23) and encodes a receptor tyrosine kinase.[23,24] Similar to other receptor tyrosine kinases, ALK contains an extracellular domain, a transmembrane segment, and a cytoplasmic receptor kinase segment.[24] *ALK* mutations have been implicated in tumorigenesis, with involvement in the initiation and progression of several cancer types, including lymphomas, neuroblastoma, and NSCLC.[25,26] *ALK* translocations typically cause an increase in tyrosine kinase

activity, resulting in increased cell proliferation and survival *via* their effects on signaling pathways that include phospholipase Cγ, phosphatidylinositol 3-kinase (PI3K)–protein kinase B (AKT), mammalian target of rapamycin (mTOR), and mitogen-activated protein kinase (MAPK) signaling cascades, among others.[26] Translocation events at the *ALK* locus generate a variety of ALK fusion proteins, such as NPM1–ALK, that are found in multiple types of cancers.[26] *ALK* activation in cancer may also arise through overexpression and point mutation of full-length *ALK*.[27–29]

Anaplastic lymphoma kinase was first identified as the receptor tyrosine kinase in a novel fusion gene arising from chromosomal translocation t(2;5) in anaplastic large-cell lymphoma.[30] This rearrangement results in a nucleophosmin (NPM1)–ALK fusion protein[30,31] and subsequent constitutive activation of the ALK kinase, which is normally regulated by the extracellular ligand-binding domain of the full-length receptor.[25,30] Although ALK is highly expressed during embryogenesis and appears to be involved in brain and neural development,[32] its precise physiological role in mammals remains unclear. ALK, however, is not critical for viability as $Alk^{-/-}$ mice are viable.[33]

NSCLC and ALK

In 2007, the fusion oncogene echinoderm microtubule-associated protein-like 4 (*EML4*)–*ALK* was shown to be present in 3% to 6% of patients with NSCLC.[34] This fusion oncogene is the result of a chromosome inversion (inv[2][p21;p23]) in which *EML4* fuses with the juxta membranous portion of *ALK* and replaces the extracellular and intramembranous portions.[34] *EML4-ALK* fusions form ligand-independent dimers via the coiled-coil of EML4;[35] this ligand-independent dimerization results in constitutive downstream signaling of canonical ALK pathways.[34] The *EML4*–*ALK* oncogene was shown to induce tumor formation in nude mice.[26,34] Figure 1 shows an overview of the EML4–ALK pathways involved in tumorigenesis.

ALK gene rearrangements appear to arise more frequently in specific NSCLC patient populations. Demographic characteristics associated with more frequent rearrangements include young, female, never- or light-smoker status, or the presence of tumors with an adenocarcinoma histology.[23,36–39] The reported prevalence of *ALK* rearrangements in Chinese patients ranges between 3.3% and 11.6%, compared with 30% of patients with *EGFR* mutations;[40–42] however, the prevalence of *ALK* rearrangements in the Chinese population is similar to that reported in other Asian populations (for
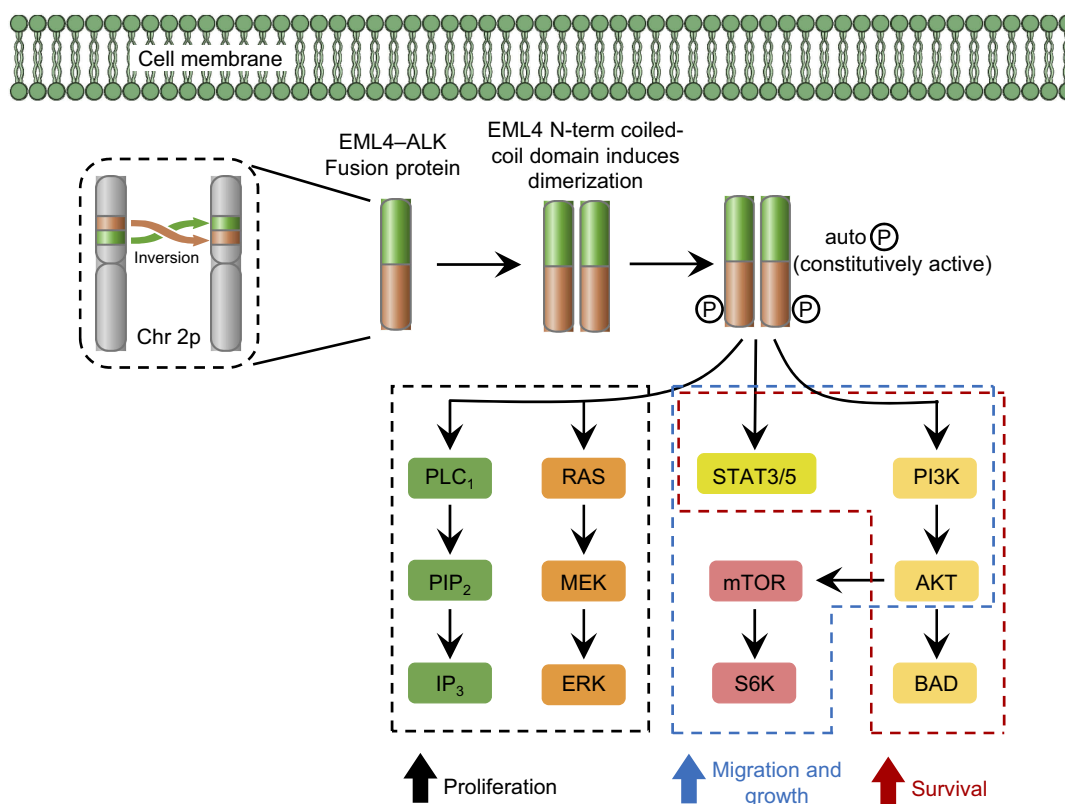
Figure 1 Overview of EML4–*ALK* pathways.
Abbreviations: AKT, protein kinase B; ALK, anaplastic lymphoma kinase; BAD, Bcl-2-associated death promoter; Chr 2p, chromosome 2p; EML4, echinoderm microtubule associated protein like 4; ERK, extracellular signal-regulated kinase; IP_3, inositol 1,4,5-triphosphate; MEK, mitogen-activated protein kinase kinase; mTOR, mammalian target of rapamycin; PI3K, phosphatidylinositol 3-kinase; PIP_2, phosphatidylinositol (3,4)-bisphosphate; PLC_1, phospholipase C-1; RAS, rat sarcoma; S6K, S6 kinase; STAT3/5, signal transducer and activator of transcription 3/5.

example, 7% in Japanese patients).[23] In general, the prevalence of *ALK* rearrangements in Asian populations is slightly greater than that in non-Asian populations.[43–46] It has been reported that the clinicopathologic features of *ALK*-rearranged lung adenocarcinoma in Western patients may differ from those in other populations.[47] However, there are suggestions that this discrepancy may result from differing pathologic interpretations rather than actual ethnicity-related genetic differences in patients with *ALK* fusion oncogene-positive lung cancer.[26]
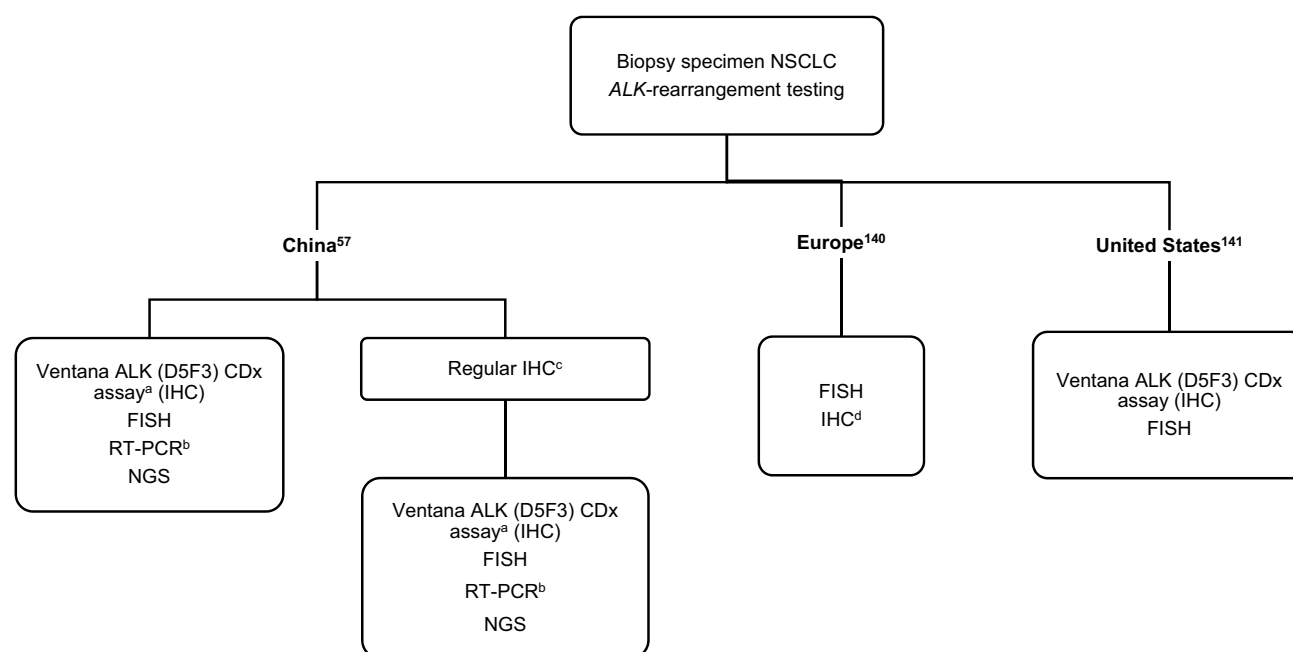
Testing methods for *ALK*-rearranged NSCLC

The general consensus of the International Association for the Study of Lung Cancer Atlas[48] is that all patients presenting with advanced NSCLC should be screened for the presence of *ALK* gene rearrangements.[14] *ALK–EML4* gene fusions and *ALK* rearrangements in NSCLC can be identified in clinical specimens using several techniques: immunohistochemistry (IHC), fluorescence in situ hybridization

(FISH), reverse transcriptase polymerase chain reaction (RT-PCR), and next generation sequencing (NGS).[49] Figure 2 shows the currently recommended testing methods for *ALK*-rearranged NSCLC.

FISH was clinically validated in studies with the first-generation tyrosine kinase inhibitor (TKI), crizotinib, prior to its approval by the US Food and Drug Administration (FDA) for *ALK*-rearranged NSCLC.[50] The *ALK* FISH approach is the gold standard and uses ALK break-apart probes that label the 5′ and 3′ ends of the *ALK* gene with fluorescent probes. Rearrangements in *ALK* result in a split appearance of the signal or loss of the 5′ signal in at least 15% of the cells counted.[51] FISH can be performed on formalin-fixed paraffin-embedded tissue or snap-frozen samples, but its use is limited by high costs, suboptimal reliability, and technical complexity.[49]

IHC is a widely available, cost-effective, and rapid technique for *ALK* screening that can be performed before FISH analysis.[52,53] In 2013, China was the first country to approve the VENTANA ALK (D5F3) CDx Assay, which uses IHC to identify the presence of *ALK*+ gene rearrangements. This

Biopsy specimen NSCLC
ALK-rearrangement testing

China[57]

Europe[140]

United States[141]

Ventana ALK (D5F3) CDx
assay[a] (IHC)
FISH
RT-PCR[b]
NGS

Regular IHC[c]

FISH
IHC[d]

Ventana ALK (D5F3) CDx
assay (IHC)
FISH

Ventana ALK (D5F3) CDx
assay[a] (IHC)
FISH
RT-PCR[b]
NGS

Figure 2 Testing methods for *ALK*-rearranged NSCLC as used in China,[57] Europe[140] and the United States.[141]
Notes: [a]Preferred method.[57] [b]Conducted in a laboratory meeting the qualifications of the National Center for Clinical Laboratories.[57] [c]Only performed when the Ventana ALK (D5F3) CDx assay is not reasonably available, primary screening only.[57] [d]Any IHC assay that has been validated against FISH.[140]
Abbreviations: NSCLC, non-small cell lung cancer; ALK, anaplastic lymphoma kinase; FISH, fluorescent in situ hybridization; RT-PCR, reverse transcription polymerase chain reaction; IHC, immunohistochemistry.

assay is a key step in qualitative identification, which is used to assess the presence of ALK protein in samples from patients with *ALK*+ NSCLC, and is often subsequently confirmed by FISH analysis. At present, Ventana IHC (D5F3) remains the most widely used testing method for *ALK*+ NSCLC.[54,55] Furthermore, it was recommended by Chinese guidelines as the first choice for *ALK*+ NSCLC diagnosis in 2018.[56–58]

Quantitative RT-PCR represents a highly sensitive method capable of detecting *ALK* mRNA that has both fusions and mutations. However, it has some limitations including (1) the requirement for frozen or fresh tissues or cells for RNA extraction,[49] and (2) the existence of many variants of *EML4-ALK*, which raises the possibility of additional variant fusions, making multiplexed RT-PCR assays difficult to optimize for clinical use.[59]

Emerging technologies for testing include NGS and liquid biopsy.[60,61] NGS is based on highly multiplexed PCR amplicon-based targeted sequencing for oncogenic fusions and represents a viable alternative to FISH because it can be performed on formalin-fixed paraffin-embedded samples, with minimal RNA input needed.[61] Although NGS requires a supporting e-infrastructure to be performed,[62] it offers the advantage of being able to test for a broader range

of mutations simultaneously as well as for both somatic and germline mutations.[60,61] Liquid biopsy involves examining cell-free DNA or circulating tumor DNA, a process that is advantageous when testing for metastatic cancers, but this method offers less sensitivity than tissue-testing for early stage cancers.[60]

The aforementioned techniques differ in their relative sensitivities and specificities, their usefulness for detecting a given type of mutation (eg, whether it involves a fusion or not), and their ease of conduct and affordability. One study assessed patients whose samples were tested by both Ventana IHC and RT-PCR; in addition, some of the samples were also tested by NGS.[56] In this study, it was determined that Ventana IHC was both a reliable and rapid method to identify suitable candidates for *ALK* gene-targeted therapy. An economic analysis from a Chinese healthcare system perspective investigated the cost-effectiveness of three *ALK* rearrangement testing methods: Ventana IHC, qRT-PCR, and IHC following FISH confirmation.[63] Here, it was shown that gene-guided therapy was a cost-effective option with or without Chinese patient-assisted programs. The results of this economic analysis were supported in a more recent follow-up study, which compared the cost-effectiveness and quality-

adjusted life-years when using NGS or multiplex PCR testing to guide therapy.[64] Furthermore, *ALK* testing under real-world settings across 31 hospitals around China is currently under investigation.

Chinese guidelines on the diagnosis and treatment of *ALK*+ NSCLC, set by the Chinese Society of Clinical Oncology Cancer Markers Expert Committee and the Chinese Expert Consensus Opinion of the diagnosis of *ALK*-positive NSCLC, provide guidance on choosing the most suitable technique for identifying *ALK* gene fusions and rearrangements.[57,65] As per their assessment, the highest level of evidence supports the use of Ventana IHC to screen for *ALK*+ NSCLC, which became widely accepted based on findings from later clinical studies, including the ASCEND-4 and ALEX studies of alectinib.[57,65–67] In contrast, the RT-PCR technique was recognized as being able to detect known fusion genes, but its performance requires an appropriately qualified laboratory.[57,65] Lastly, the committee determined that the blood sample testing was still considered experimental and should only be performed in certain circumstances.[57,65]

In China, a multicenter survey of 932 patients with advanced NSCLC showed that 71.4% of patients were tested for *EGFR* gene mutations, 44.7% were tested for *ALK* gene fusions, and 13.7% were tested for *ROS1* gene fusions.[68] The clinical assays most used in China to determine ALK involvement in NSCLC are IHC and RT-PCR, with IHC being the most widely used testing method for ALK.[69] The use of FISH is still rare due to the high cost and extensive operational requirements.[69] To date, the China National Medical Products Administration (NMPA) has approved three FISH kits, two IHC kits, six RT-PCR kits, and four NGS solutions, and current Chinese guidelines recommend NMPA-approved kits and methods in clinical *ALK*+ NSCLC diagnosis.[51] Although various options to test for *ALK* rearrangements are available, the 2018 Chinese guidelines state that their highest recommendation is for the Ventana IHC.[57,65] However, in acknowledging that this test may not be available at all sites, the guidelines committee recommends either forwarding samples to laboratories that could perform the Ventana IHC, or performing a primary screening locally using routine IHC and subsequently sending positive samples to other laboratories for confirmation by recommended methods, including FISH, Ventana IHC, or RT-PCR.[57,65]

ALK-targeted therapy in NSCLC
Overview of NSCLC treatment and the role of *ALK* inhibitors

During recent years, several ALK inhibitors that demonstrate significant benefits in the treatment of *ALK*+ NSCLC compared with conventional chemotherapy have become available in clinical practice around the world or are under clinical investigation.[37,39,70–72] Although the first generation TKI crizotinib is effective in patients with NSCLC compared with standard chemotherapy,[39] it is limited by the relatively shorter progression-free survival (PFS), its associated toxicities, and the disease progression of central nervous system (CNS) metastases due to its poor penetration of the blood–brain barrier.[49,73,74] Subsequent generations of TKIs were developed to achieve improved outcomes and fewer toxicities including second generation TKIs such as ceritinib, ensartinib, brigatinib, and alectinib.[23,75] The pharmacokinetic properties of the ALK inhibitors crizotinib, ceritinib, ensartinib, brigatinib, and alectinib are shown in Table 1, which outlines the absorption, distribution, metabolism, and excretion rates of each agent. Efficacy results for ALK inhibitors are presented in Table 2.

Limitations of *ALK* inhibitors

A limitation of the use of some members of this class of TKIs is the emergence of treatment resistance.[76,77] Several mechanisms of resistance to each targeted therapy have been identified, but these can be broadly categorized into two main classes: (1) alteration of the driver oncogene, or (2) activation of a critical parallel or downstream signaling pathway(s) that promotes pro-survival signaling.[14] Resistance to crizotinib involves mechanisms such as secondary mutations within the ALK tyrosine kinase domain and activation of alternative signaling pathways.[78] Ceritinib, more potent than crizotinib, is active against multiple *ALK* mutations that appear to result in resistance to crizotinib.[79,80] In the case of ceritinib, the increased potency appears to be associated with a higher rate of adverse events.[81] Alectinib is a second-generation ALK inhibitor that has both high selectivity[82] and efficient blood–brain barrier penetrance;[83,84] these characteristics are believed to contribute to the prevention of treatment resistance in naïve patients. In addition, alectinib can overcome crizotinib treatment resistance in patients who have relapsed.[85]

Each TKI can target different oncogenic drivers, which can affect the likelihood of treatment resistance occurring through secondary mutations or bypass mechanisms.

Table 1 Key pharmacokinetics of first- and second-line ALK inhibitors

Parameter	Crizotinib[115]	Alectinib[114]	Brigatinib[128]	Ceritinib[115]	Ensartinib[75]
T_{max} (h)	4–6	3–5	1–4	4–6	3.1–3.6
T_{ss} (d)	15	7	NR	15	NR
C_{ss} (ng/mL)	100–135	665	NR	800	NR
AUC_{inf} (ng mL/h)	2192–2946	7430	8165	NR	5330–5530
$T_{1/2}$ (h)	42	33	25	41	33.2–37.7
CL (L/h) single dose	100	N/A	NR	88.5	NR
CL (L/h) steady state	60	81.9	12.7	33	NR
Fb (%)	91	99	66	97	NR
V_d/F (L)	1772 (50 mg IV)	4016	153	4230 (750 mg orally)	NR
R	4.5	6	1.9–2.4	6.2	NR
Fecal (%)/urine (%) excretion	53/2.3	98/0.5	65/25	68/1.3	NR
Metabolism	CYP3A4/5	CYP3A4	CYP2C8/CYP3A4	CYP3A	NR

Abbreviations: AUC_{inf}, area under the curve from 0 to infinity; $T_{1/2}$, half-life; CL, clearance; C_{ss}, steady state concentration (ng/mL; μM); CYP3A, cytochrome P450 3A; CYP3A4, cytochrome P450 3A4; CYP3A4/5, cytochrome P450 3A4/5; F, bioavailability; Fb, fraction bound to plasma protein; IV, intravenous; N/A, not applicable; NR, not reported; R, accumulation ratio; T_{max}, time to maximum concentration; T_{ss}, time to steady state; V_d/F, volume of distribution.

Targets for each TKI are as follows: alectinib (inhibitor of ALK tyrosine kinase and RET kinase, including the *ALK* L1196M mutant); ensartinib (*ROS* proto-oncogene 1 receptor tyrosine kinase, *ROS1; MET* protocol-oncogene receptor tyrosine kinase, *MET; STE20* like kinase, *SLK; AXL* receptor tyrosine kinase, *AXL*; leukocyte receptor tyrosine kinase, *LTK; ABL* proto-oncogene 1 non-receptor tyrosine kinase, *ABL1*; and *EPH* receptor A2, *EPHA2*); brigatinib (inhibitor of *ALK* and *EGFR*, including *ALK* L1196M and *EGFR* T790M mutants); lorlatinib (ALK and ROS1); and repotrectinib (ALK, ROS1, and TRK).[14,23,75–87]

The treatment landscape in China

The first Chinese guidelines for the diagnosis and treatment of primary lung cancer were published in 2003, and then updated in 2011, 2015, and 2018.[57,88] First-line drug regimens in cases of advanced NSCLC include platinum-doublet chemotherapy and targeted molecular therapy drugs, such as gefitinib, erlotinib, or icotinib if EGFR mutations are detected; or crizotinib if ALK fusion genes are detected. Treatment of NSCLC in China is characterized by many factors, which include the accessibility and availability of diagnostic assays and treatments, insurance reimbursement rates,[89] and the accuracy of decision making in the Chinese healthcare system.[90]

More recently, Chinese guidelines for the treatment and diagnosis of *ALK*+ and *ROS1*+ NSCLC were published.[57] The first TKI to be approved by the Chinese FDA for advanced *ALK*+ NSCLC was crizotinib in 2013.[91] Global studies have shown that treatment resistance to crizotinib is

inevitable;[70,73] however, a retrospective study observed that continuation of crizotinib therapy in Chinese patients beyond the initial disease progression may provide further benefits.[92] The first second-generation TKI to be developed was ceritinib, which initially showed efficacy in the ASCEND-1[93] trial, and was approved by the US FDA shortly thereafter, in 2015.[81] However, a subsequent phase III study[94] showed inferior efficacy of ceritinib compared with the results of a phase II study.[95] Furthermore, with the publication of the ALEX[66] and J-ALEX[96] trials, alectinib achieved a median PFS of 34.8 months (95% CI: 17.7–not estimable) compared with 10.9 months (95% CI: 9.1–12.9) with crizotinib, making it the drug of choice for advanced *ALK*+ NSCLC (HR 0.43 [95% CI: 0.32–0.58]).[97]

Following priority review, alectinib was approved for use in China in August 2018 as a monotherapy to treat patients with locally advanced or metastatic *ALK*+ NSCLC.[98] This approval allows the use of alectinib for both ALK-inhibitor naïve or treated patients. Other second-generation TKIs, ensartinib and brigatinib; the third-generation TKI lorlatinib; and the fourth-generation TKI repotrectinib are not currently available in China.

Alectinib

Alectinib (CH5424802/RO5424802) is a potent and highly selective second-generation inhibitor of ALK tyrosine kinase that acts only on *ALK*+ NSCLC.[99,100] It also inhibits RET kinase activity and, thus, may prove efficacious against RET fusion+ tumors.[101] Furthermore, alectinib exhibits activity against multiple gate-keeper mutations that impart resistance to crizotinib, and can

Table 2 Clinical outcomes of phase I–III studies of ALK inhibitors in crizotinib-naïve or crizotinib resistant patients, with or without chemotherapy

Drug	Study phase Study name	Previous crizotinib		Crizotinib-naive	
		ORR	mPFS (months)	ORR	mPFS (months)
Alectinib	Phase I/II AF-001JP//JapicCTI-101264[129] *Naïve to ALK inhibitors*	–	–	94% (43/46)	NA
	Phase I/II dose escalation AF-002JG//NCT01588028[111] *Crizotinib pretreated*	55% (24/44)	NA	–	–
	Phase II NP28761//NCT01871805[127] *Crizotinib pretreated*	52% (35/67)	8.0	–	–
	Phase II NP28673//NCT01801111[121] *Crizotinib pretreated*	45% (61/96)	8.9	–	–
	Phase III global ALEX//NCT02075840[124] *Untreated*	–	–	83% (126/152)	Not reached
	Phase III J-ALEX//JapicCTI-132316[96] *Naïve to ALK inhibitors, Asian Patients*	–	–	92% (76/83)	Not reached
	Phase III ALUR, MO29750//NCT02604342[85] *Crizotinib pretreated*	54%[a] (13/24)	9.6	–	–
	Phase III ALESIA//NCT02838420[120] *Naïve to ALK inhibitors, Asian Patients*	91% (114/125)	Not reached	77% (48/62)	11.1
	Systematic meta-analysis of 8 studies[118] *Mixed patient status*	52% [46–58]	9.36[b] [7.38–11.34]	87% [81–92]	Not reported
Crizotinib	Phase I PROFILE 1001//NCT00585195[130]	–	–	61% (87/143)	9.7
	Phase II PROFILE 1005//NCT00932451[131]	–	–	54% (491/908)	8.4
	Phase III 2nd line PROFILE 1007//NCT0093283[39]	–	–	65% (113/173)	7.7
	Phase III 1st line PROFILE 1014//NCT01154140[70]	–	–	74% (128/172)	10.9
	Phase III 1st line PROFILE 1029//NCT01639001[132]	–	–	88% (91/104)	11.1
Ceritinib	Phase I ASCEND-1//NCT01283516[93]	56% (92/163)	6.9	72% (60/83)	18.4
	Phase II ASCEND-2//NCT01685060[95]	39% (54/140)	5.7	–	–
	Phase II ASCEND-3//NCT01685138[133]	–	–	64% (79/124)	11.1
	Phase III ASCEND-4//NCT01828099[134]	–	–	73% (137/189)	16.6
	Phase III ASCEND-5//NCT01828112[94]	39% (45/115)	5.4	–	–
	Phase I/II ASCEND-6//NCT02040870[135]	41% (42/103)	5.7	–	–

(Continued)

Table 2 (Continued).

Drug	Study phase Study name	Previous crizotinib		Crizotinib-naive	
		ORR	mPFS (months)	ORR	mPFS (months)
Ensartinib	Phase I/II NCT01625234[75]	69% (20/29)	9.0	80% (12/15)	26.2
Brigatinib	Phase I/II, phase II portion NCT01449461[136]	71% (50/70)	13.4	100% (8/8)	Not reached
	Phase III ALTA-1L//NCT02737501[137]	–	–	71% (97/137)	Not reached
Lorlatinib	Phase I, dose escalation component NCT01970865[138]	53% (17/32)	NA	100% (2/2)	NA
	Phase II NCT01970865[139]	70% (41/59)	Not reached	90% (27/30)	Not reached

Notes: [a]Represents CNS ORR in patients with measurable baseline CNS disease. [b]Represents all patients in analysis set: analysis not performed by previous crizotinib exposure.
Abbreviations: ALK, anaplastic lymphoma kinase; CNS, central nervous system; ORR, objective response rate; mPFS, median progression-free survival; NA, not available.
Note, parentheses enclose fractions of patients and square brackets enclose 95% confidence intervals.

cross the blood–brain barrier.[83,102–106] Notably, alectinib shows activity both in patients who are crizotinib treatment-naïve and in patients who have demonstrated resistance to crizotinib (Table 2).[99,107] In contrast to ceritinib, alectinib has a lower incidence of adverse events.[81] The clinical outcomes of phase I–III studies of alectinib are presented in Table 2.

Prior to its indication in China last year, alectinib was approved in the EU for *ALK+* advanced NSCLC patients either as a first-line treatment[108] or for those previously treated with crizotinib,[109] and in the US for *ALK+* metastatic NSCLC as detected by an US FDA-approved test.[110]

Mode of action

The unique chemical structure of alectinib means it 1) targets both *ALK* rearrangements and RET fusion+ tumors, but not MET or ROS1 kinase activity, and 2) overcomes acquired resistance to crizotinib through its ability to target the mutations that develop with crizotinib treatment.[83,102–106] Its features include a scaffold-like structure with lipophilic properties and an ability to cross the blood–brain barrier, possibly because it is not a substrate of the key efflux transporter that delays blood–brain barrier penetration. Additionally, an increased potency over crizotinib is evident in the three-fold increase in in vitro ALK inhibition (53 nM alectinib versus 150.8 nM crizotinib).[99] Together, these characteristics may contribute to its ability to overcome resistance

to other ALK inhibitors caused by mutations[83,84,100,102,111] and its increased potency over crizotinib for treating CNS metastases.[83,84] Successful treatment of CNS metastases is demonstrated based on a pooled analysis of data from two phase II studies (NCT01871805 and NCT01801111) that reported a 64.0% CNS objective response rate (95% confidence interval [CI]: 49.2–77.1) and a median CNS duration of response of 10.8 months (95% CI: 7.6–14.1) in crizotinib-refractory *ALK+* NSCLC patients with measurable CNS disease (n=50) at baseline.[112]

Metabolism and pharmacokinetics

Alectinib is primarily metabolized by the cytochrome P450 (CYP) 3A4 enzyme, producing its major active metabolite M4 (Table 1).[113] Most of the drug is excreted in the feces.[113] The pharmacokinetics of alectinib relative to other ALK inhibitors are presented in Table 1. Alectinib exhibits a time to maximum concentration of 3–5 hrs, and a half-life of 33 hrs.[114] The maximum steady-state concentration for alectinib is higher than that of crizotinib, by approximately 4-fold, while alectinib achieves this concentration earlier than crizotinib (Table 1).[114,115] It is important to note that the pharmacokinetics of alectinib does not appear to differ by race. Analyses of pharmacokinetic data from the phase III ALEX trial of alectinib, which was prospectively stratified by Asian and non-Asian patients, did not exhibit any notable differences.[116]

Clinical efficacy of alectinib

The global phase III ALEX study demonstrated prolonged PFS in newly diagnosed patients receiving alectinib versus those receiving crizotinib.[97] The study reported a median PFS of 34.8 months (95% CI: 17.7–not estimable), with only 12% of patients developing brain metastases, as compared with 10.9 months (95% CI: 9.1–12.9) and 45%, respectively, in patients treated with crizotinib. In another phase III trial (ALUR) across 13 countries in Europe and Asia, alectinib versus platinum-based chemotherapy in crizotinib-pretreated patients exhibited prolonged median PFS, both investigator assessed and independent review committee assessed: 9.6 months (95% CI: 6.9–12.2) with alectinib and 1.4 months (95% CI: 1.3–1.6) with chemotherapy (investigator assessed) with a hazard ratio (HR) of 0.15 (95% CI: 0.08–0.29; $p<0.001$).[85] Therefore, the long-term benefits of alectinib in ALK inhibitor-naïve patients have been demonstrated in a number of different populations.[117]

A systematic meta-analysis of eight alectinib trials was published recently and presented the overall pooled efficacy and safety results, and results stratified by prior treatment status.[118] The pooled overall response rate (ORR) was 70% (95% CI: 57–82), with ALK-inhibitor-naïve patients having an ORR of 87% (95% CI: 81–92) versus crizotinib-resistant patients, who had an ORR of 52% (95% CI: 46–58).[118] Table 2 summarizes clinical efficacy results from individual trials of alectinib.

A recent report of the pooled efficacy data after 15- and 18-month follow-up of two key phase II studies (NCT01871805 and NCT01801111; n=255) showed that alectinib has a durable response rate (median =14.9 months; 95% CI: 11.1–20.4) with a 78.8% disease control rate (95% CI: 72.3–84.4) and median PFS of 8.3 months (95% CI: 7.0–11.3).[119] There were also no specific ethnic differences in the efficacy of alectinib observed.

Clinical evidence in Chinese patients

The superior efficacy of alectinib compared with crizotinib has been reported in an Asian versus non-Asian patient subgroup analysis in the global phase III ALEX study[66,116] and in the ALESIA phase III randomized clinical trial, which comprised patients from China, South Korea, and Thailand.[120] The ALESIA study reported a significant improvement in both investigator-assessed (HR: 0.22; 95% CI: 0.13–0.38; $p<0.0001$; median PFS not estimable [alectinib] vs 11.1 months [crizotinib]) and independent

review committee-assessed (HR: 0.37; 95% CI: 0.22–0.61; $p<0.0001$) PFS in the alectinib vs crizotinib groups.[120] The percent of patients experiencing disease progression or death was higher with crizotinib versus alectinib treatment (60% vs 21%, respectively).[120] Although patients receiving alectinib were treated for a longer period than those receiving crizotinib (14.7 vs 12.6 months, respectively), fewer grade 3–5 adverse events (alectinib, 36 [29%] of 125; crizotinib, 30 [48%] of 62) and serious adverse events (alectinib, 19 [15%] of 125; crizotinib, 16 [26%] of 62) were reported in the alectinib vs crizotinib groups.[120] Pharmacokinetic results from a subset of 20 frequently sampled Chinese patients (600 mg alectinib, twice daily) demonstrated nearly identical pharmacokinetic profiles to white patients (historical data from a phase II global study).[120,121] These clinical data are consistent with data reported for the ALEX study,[66] suggesting that alectinib could be suitable to treat both crizotinib-naïve and crizotinib-refractory patients with *ALK*+ NSCLC, irrespective of ethnicities.

The ALEX trial also included 43 Chinese patients, of which 25 received alectinib and 18 received crizotinib, and results were reported at ESMO-Asia 2017.[116] Median PFS (independent review committee-assessed) was longer in alectinib-treated patients (25.7 vs 14.8 months; HR: 0.57, 95% CI: 0.24–1.38; $p=0.16$), suggesting the efficacy of alectinib in Chinese patients is consistent with that in the global population.

Activity in the CNS

Brain metastases are a common complication during the advanced stages of NSCLC, particularly in patients who have *ALK* gene rearrangements.[122] Therefore, systemic therapies with intracranial efficacy are the preferred long-term treatment option for patients with *ALK*+ NSCLC. Alectinib has a unique physical structure that contributes to its potency, including an improved ability to penetrate the blood–brain barrier compared to other TKIs.[83,84] Alectinib has shown potential efficacy against brain metastases as reported in the pooled analysis of data from two phase II studies (NCT01871805 and NCT01801111) involving crizotinib-refractory *ALK*+ NSCLC patients with measurable CNS disease (n=50) at baseline, which reported a 64.0% CNS ORR (95% CI: 49.2–77.1) and a median CNS duration of response of 10.8 months (95% CI: 7.6–14.1).[110] In a retrospective study of patients with advanced *ALK*+ NSCLC treated with alectinib, the ORR

was 73.3% and the disease control rate was 100.0%, with a median CNS duration of response of 19.3 months.[123]

Patients receiving alectinib in the phase III study, ALEX, exhibited longer times to CNS progression than those patients receiving crizotinib, and the two groups were balanced in CNS disease levels at baseline.[111] A recently published update of the phase III ALEX study supports the superior CNS efficacy of alectinib over crizotinib.[124] Here, it was reported that alectinib had a significantly longer time to CNS progression, regardless of the presence of CNS metastases at baseline. Notably, when evaluating the cumulative incidence rate (CIR) of CNS progression in patients without baseline CNS metastases, it was shown that the 12-month CIR was only 4.6% in the alectinib group compared with 31.5% in the crizotinib group.[125] In addition, the CIR of CNS progression in patients with brain metastases at baseline was 16.0% and 58.3%, respectively. These data demonstrate a superior efficacy and capability of delaying CNS progression for patients treated with alectinib, even in patients without brain metastases at baseline.

A systematic meta-analysis has also described the clinical outcomes among patients with *ALK*+ NSCLC and brain metastases.[89] Patients who received alectinib exhibited a pooled ORR of 52.0% (95% CI: 45.0–59.0), with an ORR of 59.0% (95% CI: 47.0–71.0) among ALK-inhibitor-naïve patients versus 48.0% (95% CI: 38.0–57.0) among crizotinib-resistant patients.[89] In the phase III ALUR study of European and Asian patients previously treated with crizotinib, it was shown that there was a greater CNS ORR in patients receiving alectinib versus those receiving chemotherapy (54.2% vs 0%, *p*<0.001).[85] Similarly, Asian patients receiving alectinib in the ALESIA study exhibited larger CNS ORR compared with patients receiving crizotinib: 73% vs 22%, respectively, in patients with and without measurable CNS lesions at baseline.[120]

The observed efficacy of alectinib in *ALK*+ NSCLC patients without brain metastases suggests that alectinib may also prevent NSCLC progression to the brain. Therefore, alectinib is a CNS active ALK TKI that is not only active against baseline brain metastases but may also provide long-term benefits to the patient through its ability to potentially prevent the development of further brain metastases.

Safety profile of alectinib

Alectinib is generally well tolerated and has been reported to have a good toxicity profile.[119,126,127] The incidence of adverse events in clinical trials is relatively low and this

has been reported in both patients with and without baseline brain metastases.[85,112] A systematic meta-analysis of eight clinical trials reported that the most common adverse events were constipation (29%), anemia (25%), myalgia (18%), peripheral edema (18%), dysgeusia (18%), and blood creatine phosphokinase increased (18%).[118] This meta-analysis also reported a pooled discontinuation rate of 7% (95% CI: 4–10) and a pooled dose reduction or interruption rate of 33% (95% CI: 24–42) among alectinib-treated patients.[118]

The Japanese J-ALEX clinical trial compared alectinib (300 mg b.i.d) to crizotinib (250 mg b.i.d) and showed that alectinib had a better safety profile than crizotinib.[96] In comparison to crizotinib, the incidence of grade 3/4 adverse events (26% versus 52%) and proportion of patients who discontinued treatment (9% versus 20%) were lower in alectinib-treated patients. Furthermore, the phase III ALEX trial also reported fewer grade 3–5 adverse events in treatment-naïve patients receiving alectinib relative to those receiving crizotinib.[66] The phase III ALEX trial was a global study and showed that at a dose of 600 mg (b.i.d), grade 3–5 adverse events were reported in 41% of patients, with 11% of patients discontinuing treatment. In both the J-ALEX and ALEX trials, the incidence of adverse events was consistently lower compared to crizotinib except for myalgia and anemia, which were more frequent after alectinib treatment. Similar results were also observed in the phase III ALUR trial of crizotinib-pretreated patients and in the phase III ALESIA trial in Asian patients: fewer grade ≥3 adverse events and fewer adverse events leading to discontinuation occurred in patients receiving alectinib versus those receiving the other treatment.[85,120]

Conclusions

NSCLC has a very poor prognosis;[13] however, the development of treatments that target pathways involved in tumorigenesis can improve patient survival. Targeting the NSCLC oncogenic-driver gene *ALK* has resulted in the development of various *ALK* inhibitors with notable effects in prolonging patient survival. Currently recommended testing methods for identifying *ALK* gene rearrangements in NSCLC patients include the use of Ventana IHC, which is recommended by the Chinese guidelines.[57,65] Once the *ALK* gene mutation status has been assessed, current TKI therapies confer efficacy, albeit with the risk of toxicities or eventual regression in some cases. Several of these agents, however, have multiple targets that may contribute to the development of resistance (eg, crizotinib) or that may result

in greater toxicity (eg, ceritinib). Alectinib, a TKI that potently and selectively targets the ALK and MET pathways, results in prolonged PFS, even in patients with CNS involvement or patients with or without previous exposure to crizotinib, with lower rates of adverse events and dose reductions or interruptions due to adverse events. Recently approved in China, alectinib may prove to be the best first-line agent to help improve the outcomes of *ALK+* locally advanced or metastatic NSCLC patients.

Acknowledgments

Author contributions

All authors contributed to data analysis, drafting and revising the article, gave final approval of the version to be published, and agree to be accountable for all aspects of the work.

Disclosure

Jinjing Xia is an employee of Shanghai Roche Pharmaceuticals Ltd. All other authors report no conflicts of interest in this work.

References

1. Zhou C. Lung cancer molecular epidemiology in China: recent trends. *Transl Lung Cancer Res*. 2014;3:270–279.doi:10.3978/j.issn.2218-6751.2014.09.01
2. Gridelli C, Peters S, Sgambato A, et al. ALK inhibitors in the treatment of advanced NSCLC. *Cancer Treat Rev*. 2014;40:300–306. doi:10.1016/j.ctrv.2013.07.002
3. Torre LA, Bray F, Siegel RL, et al. Global cancer statistics, 2012. *CA Cancer J Clin*. 2015;65:87–108. doi:10.3322/caac.21294
4. Chen W, Zheng R, Baade PD, et al. Cancer statistics in China, 2015. *CA Cancer J Clin*. 2016;66:115–132. doi:10.3322/caac.21338
5. Chen W, Zheng R, Zhang S, et al. Annual report on status of cancer in China, 2010. *Chin J Cancer Res*. 2014;26:48–58.
6. Alberg AJ, Brock MV, Samet JM. Epidemiology of lung cancer: looking to the future. *J Clin Oncol*. 2005;23:3175–3185. doi:10.1200/JCO.2005.10.462
7. Peace LR. A time correlation between cigarette smoking and lung cancer. *Statistician*. 1985;34:371–381. doi:10.2307/2987825
8. Liu BQ, Peto R, Chen ZM, et al. Emerging tobacco hazards in China: 1. Retrospective proportional mortality study of one million deaths. *BMJ*. 1998;317:1411–1422. doi:10.1136/bmj.317.7159.682
9. Yang L, Parkin DM, Li L, et al. Time trends in cancer mortality in China: 1987–1999. *Int J Cancer*. 2003;106:771–783. doi:10.1002/ijc.11300
10. Chen W, Zheng R, Zeng H, Zhang S. Epidemiology of lung cancer in China. *Thorac Cancer*. 2015;6:209–215. doi:10.1111/tca.2015.6.issue-2
11. Barzi A, Pennell NA. Targeting angiogenesis in non-small cell lung cancer. *Eur J Clin Med Oncol*. 2010;2:31–42.
12. Siegel R, Ma J, Zou Z, Jemal A. Cancer statistics, 2014. *CA Cancer J Clin*. 2014;64:9–29. doi:10.3322/caac.21208
13. Howlader N, Noone AM, Krapcho M, et al. *SEER Cancer Statistics Review, 1975-2016*. National Cancer Institute; Posted April 2019. Available from: https://seer.cancer.gov/csr/1975_2016/. Accessed June 8, 2019.
14. Addeo A, Tabbò F, Robinson T, et al. Precision medicine in ALK rearranged NSCLC: A rapidly evolving scenario. *Crit Rev Oncol/Hematol*. 2018;122:150–156. doi:10.1016/j.critrevonc.2017.12.015
15. Fernandez-Cuesta L, Plenker D, Osada H, et al. CD74–NRG1 fusions in lung adenocarcinoma. *Cancer Discov*. 2014;4:415–422. doi:10.1158/2159-8290.CD-13-0646
16. Zheng D, Wang R, Ye T, et al. MET exon 14 skipping defines a unique molecular class of non-small cell lung cancer. *Oncotarget*. 2016;5:41691–41702.
17. Do H, Solomon B, Mitchell PL, Fox SB, Dobrovic A. Detection of the transforming AKT1 mutation E17K in non-small cell lung cancer by high resolution melting. *BMC Res Notes*. 2008;1:14. doi:10.1186/1756-0500-1-23
18. Gainor JF, Varghese AM, Ou S-HI, et al. ALK rearrangements are mutually exclusive with mutations in EGFR or KRAS: an analysis of 1,683 patients with non-small cell lung cancer. *Clin Cancer Res*. 2013;19:4273–4281. doi:10.1158/1078-0432.CCR-13-0318
19. Lee T, Lee B, Choi YL, et al. Non-small cell lung cancer with concomitant EGFR, KRAS, and ALK mutation: clinicopathologic features of 12 cases. *J Pathol Transl Med*. 2016;50:197. doi:10.4132/jptm.2016.03.09
20. Sweiss RF, Thomas S, Bank B, et al. Concurrent EGFR mutation and ALK translocation in non-small cell lung cancer. *Cureus*. 2016;8:e513.
21. Lo Russo G, Imbimbo M, Corrao G, et al. Concomitant EML4-ALK rearrangement and EGFR mutation in non-small cell lung cancer patients: a literature review of 100 cases. *Oncotarget*. 2017;8:59889–59900. doi:10.18632/oncotarget.17431
22. Stella GM, Scabini R, Inghilleri S, et al. EGFR and KRAS mutational profiling in fresh non-small cell lung cancer (NSCLC) cells. *J Cancer Res Clin Oncol*. 2013;139:1327–1335. doi:10.1007/s00432-013-1444-y
23. Wu J, Savooji J, Liu D, et al. Second- and third-generation ALK inhibitors for non-small cell lung cancer. *J Hematol Oncol*. 2016;9:19. doi:10.1186/s13045-016-0251-8
24. Duyster J, Bai R-Y, Morris SW. Translocations involving anaplastic lymphoma kinase (ALK). *Oncogene*. 2001;20:5623. doi:10.1038/sj.onc.1204594
25. Iwahara T, Fujimoto J, Wen D, et al. Molecular characterization of ALK, a receptor tyrosine kinase expressed specifically in the nervous system. *Oncogene*. 1997;14:439. doi:10.1038/sj.onc.1200849
26. Hallberg B, Palmer RH. Mechanistic insight into ALK receptor tyrosine kinase in human cancer biology. *Nat Rev Cancer*. 2013;13:685.
27. Kruczynski A, Delsol G, Laurent C, et al. Anaplastic lymphoma kinase as a therapeutic target. *Expert Opin Ther Targets*. 2012;16:1127–1138. doi:10.1517/14728222.2012.719498
28. Mossé YP, Wood A, Maris JM. Inhibition of ALK signaling for cancer therapy. *Clin Cancer Res*. 2009;15:5609–5614. doi:10.1158/1078-0432.CCR-09-0547
29. Palmer RH, Vernersson E, Grabbe C, et al. Anaplastic lymphoma kinase: signalling in development and disease. *Biochem J*. 2009;420:345–361. doi:10.1042/BJ20082397
30. Morris S, Kirstein M, Valentine M, et al. Fusion of a kinase gene, ALK, to a nucleolar protein gene, NPM, in non-Hodgkin's lymphoma. *Science*. 1994;263:1281–1284. doi:10.1126/science.8122112

31. Shiota M, Nakamura S, Ichinohasama R, et al. Anaplastic large cell lymphomas expressing the novel chimeric protein p80NPM/ALK: a distinct clinicopathologic entity. *Blood*. 1995;86:1954–1960.

32. Webb TR, Slavish J, George RE, et al. Anaplastic lymphoma kinase: role in cancer pathogenesis and small-molecule inhibitor development for therapy. *Expert Rev Anticancer Ther*. 2009;9:331–356. doi:10.1586/14737140.9.3.331

33. Weiss JB, Xue C, Benice T, Xue L, Morris SW, Raber J. Anaplastic lymphoma kinase and leukocyte tyrosine kinase: functions and genetic interactions in learning, memory and adult neurogenesis. *Pharmacol Biochem Behav*. 2012;100(3):566–574. doi:10.1016/j.pbb.2011.10.024

34. Soda M, Choi YL, Enomoto M, et al. Identification of the transforming EML4–ALK fusion gene in non-small-cell lung cancer. *Nature*. 2007;448:561. doi:10.1038/nature05984

35. Soda M, Takada S, Takeuchi K, et al. A mouse model for EML4-ALK-positive lung cancer. *Proc Natl Acad Sci U S A*. 2008;105:19893–19897. doi:10.1073/pnas.0805381105

36. Iragavarapu C, Mustafa M, Akinleye A, et al. Novel ALK inhibitors in clinical use and development. *J Hematol Oncol*. 2015;8:17. doi:10.1186/s13045-015-0122-8

37. Kwak EL, Bang YJ, Camidge DR, et al. Anaplastic lymphoma kinase inhibition in non-small-cell lung cancer. *N Engl J Med*. 2010;363:1693–1703. doi:10.1056/NEJMoa1011205

38. Shaw AT, Costa DB, Iafrate AJ, et al. Clinical activity of the oral ALK and MET inhibitor PF-02341066 in non-small lung cancer (NSCLC) with EML4-ALK translocations. *J Thorac Oncol*. 2009;4:S305–S306. doi:10.1097/JTO.0b013e3181b6be12

39. Shaw AT, Kim DW, Nakagawa K, et al. Crizotinib versus chemotherapy in advanced ALK-positive lung cancer. *N Engl J Med*. 2013;368:2385–2394. doi:10.1056/NEJMoa1214886

40. An SJ, Chen ZH, Su J, et al. Identification of enriched driver gene alterations in subgroups of non-small cell lung cancer patients based on histology and smoking status. *PLoS One*. 2012;7:e40109. doi:10.1371/journal.pone.0040109

41. Li Y, Li Y, Yang T, et al. Clinical significance of EML4-ALK fusion gene and association with EGFR and KRAS gene mutations in 208 Chinese patients with non-small cell lung cancer. *PLoS One*. 2013;8:e52093. doi:10.1371/journal.pone.0052093

42. Zhang X, Zhang S, Yang X, et al. Fusion of EML4 and ALK is associated with development of lung adenocarcinomas lacking EGFR and KRAS mutations and is correlated with ALK expression. *Mol Cancer*. 2010;9:188. doi:10.1186/1476-4598-9-254

43. Barlesi F, Blons H, Beau-Faller M, et al. Biomarkers (BM) France: results of routine EGFR, HER2, KRAS, BRAF, PI3KCA mutations detection and EML4-ALK gene fusion assessment on the first 10,000 non-small cell lung cancer (NSCLC) patients (pts). *J Clin Oncol*. 2013;31:8000. doi:10.1200/JCO.2013.49.0219

44. Koivunen JP, Mermel C, Zejnullahu K, et al. EML4-ALK fusion gene and efficacy of an ALK kinase inhibitor in lung cancer. *Clin Cancer Res*. 2008;14:4275–4283. doi:10.1158/1078-0432.CCR-08-0168

45. Takeuchi K, Choi YL, Soda M, et al. Multiplex reverse transcription-PCR screening for EML4-ALK fusion transcripts. *Clin Cancer Res*. 2008;14:6618–6624. doi:10.1158/1078-0432.CCR-08-1018

46. Wu YC, Chang IC, Wang CL, et al. Comparison of IHC, FISH and RT-PCR methods for detection of ALK rearrangements in 312 non-small cell lung cancer patients in Taiwan. *PLoS One*. 2013;8:e70839. doi:10.1371/journal.pone.0070839

47. Rodig SJ, Mino-Kenudson M, Dacic S, et al. Unique clinicopathologic features characterize ALK-rearranged lung adenocarcinoma in the western population. *Clin Cancer Res*. 2009;15:5216–5223. doi:10.1158/1078-0432.CCR-09-0547

48. International Association for the Study of Lung Cancer. *IASLC Atlas of ALK and ROS1 Testing in Lung Cancer*. 2nd ed. (FL): Editorial Rx Press; FT. Myers, FL, 2016.

49. Li G, Dai WR, Shao FC. Effect of ALK-inhibitors in the treatment of non-small cell lung cancer: a systematic review and meta-analysis. *Eur Rev Med Pharmacol Sci*. 2017;21:3496–3503.

50. US Food and Drug Administration. FDA approves xalkori with companion diagnostic for a type of late-stage lung cancer [press release]. 2011. Available from: http://web.archive.org/web/20110901151213/http://www.fda.gov/NewsEvents/Newsroom/PressAnnouncements/ucm269856.htm . Accessed July 30, 2019.

51. Shaw AT, Solomon B, Kenudson MM. Crizotinib and testing for ALK. *J Natl Compr Canc Netw*. 2011;9:1335–1341. doi:10.6004/jnccn.2011.0115

52. Cutz JC, Craddock KJ, Torlakovic E, et al. Canadian anaplastic lymphoma kinase study: a model for multicenter standardization and optimization of ALK testing in lung cancer. *J Thorac Oncol*. 2014;9:1255–1263. doi:10.1097/JTO.0000000000000011

53. Zhou J, Zhao J, Sun K, et al. Accurate and economical detection of ALK positive lung adenocarcinoma with semiquantitative immunohistochemical screening. *PLoS One*. 2014;9:e92828. doi:10.1371/journal.pone.0092828

54. Ying J, Guo L, Qiu T, et al. Diagnostic value of a novel fully automated immunochemistry assay for detection of ALK rearrangement in primary lung adenocarcinoma. *Ann Oncol*. 2013;24:2589–2593. doi:10.1093/annonc/mdt295

55. Liu L, Zhan P, Zhou X, et al. Detection of EML4-ALK in lung adenocarcinoma using pleural effusion with FISH, IHC, and RT-PCR methods. *PLoS One*. 2015;10:e0117032. doi:10.1371/journal.pone.0117032

56. Xu CW, Wang WX, Chen YP, et al. Simultaneous VENTANA IHC and RT-PCR testing of ALK status in Chinese non-small cell lung cancer patients and response to crizotinib. *J Transl Med*. 2018;16:93. doi:10.1186/s12967-018-1468-9

57. Zhang XC, Lu S, Zhang L. Chinese guidelines for the treatment and diagnosis of anaplastic lymphoma kinase-positive and ROS1-positive non-small-cell lung cancer [Chinese]. *Chin J Pathol*. 2018;47:241–247.

58. Zhang YG, Jin ML, Li L, et al. Evaluation of ALK rearrangement in chinese non-small cell lung cancer using FISH, immunohistochemistry, and real-time quantitative RT- PCR on paraffin-embedded tissues. *PLoS One*. 2013;8:e64821. doi:10.1371/journal.pone.0064821

59. Hout DR, Schweitzer BL, Lawrence K, et al. Performance of a RT-PCR assay in comparison to FISH and immunohistochemistry for the detection of ALK in non-small cell lung cancer. *Cancers*. 2017;9:99. doi:10.3390/cancers9080099

60. Yohe S, Thyagarajan B. Review of clinical next-generation sequencing. *Arch Pathol Lab Med*. 2017;141:1544–1557. doi:10.5858/arpa.2016-0501-RA

61. Vendrell JA, Taviaux S, Beganton B, et al. Detection of known and novel ALK fusion transcripts in lung cancer patients using next-generation sequencing approaches. *Sci Rep*. 2017;7:12510. doi:10.1038/s41598-017-12679-8

62. Spjuth O, Bongcam-Rudloff E, Dahlberg J, et al. Recommendations on e-infrastructures for next-generation sequencing. *Gigascience*. 2016;5:26. doi:10.1186/s13742-016-0132-7

63. Lu S, Zhang J, Ye M, Wang B, Wu B. Economic analysis of ALK testing and crizotinib therapy for advanced non-small-cell lung cancer. *Pharmacogenomics*. 2016;17:985–994. doi:10.2217/pgs-2016-0017

64. Lu S, Yu Y, Fu S, Ren H, Gangopadhyay N. Cost-effectiveness of ALK testing and first-line crizotinib therapy for non-small-cell lung cancer in China. *PLoS One*. 2018;13:e0205827. doi:10.1371/journal.pone.0205827

65. Zhang X, Lu S, Zhang L, et al. The Chinese expert consensus opinion of the diagnosis of ALK-positive NSCLC (2013 version) [Chinese]. *Chin J Pathol*. 2013;6:402–406.

66. Peters S, Camidge DR, Shaw AT, et al. Alectinib versus Crizotinib in untreated ALK-positive non-small-cell lung cancer. *N Engl J Med*. 2017;377:829–838. doi:10.1056/NEJMoa1704795

67. Hida T, Seto T, Horinouchi H, et al. Phase II study of ceritinib in alectinib-pretreated patients with anaplastic lymphoma kinase-rearranged metastatic non-small-cell lung cancer in Japan: ASCEND-9. *Cancer Sci*. 2018;109:2863–2872. doi:10.1111/cas.13721

68. Zhou Q, Song Y, Zhang X, et al. A multicenter survey of first-line treatment patterns and gene aberration test status of patients with unresectable Stage IIIB/IV nonsquamous non-small cell lung cancer in China (CTONG 1506). *BMC Cancer*. 2017;17:462. doi:10.1186/s12885-017-3451-x

69. Niu FY, Wu Y-L. Personalized treatment strategies for non-small-cell lung cancer in Chinese patients: the role of crizotinib. *Onco Targets Ther*. 2015;8:999–1007.

70. Solomon BJ, Mok T, Kim DW, et al. First-line crizotinib versus chemotherapy in ALK-positive lung cancer. *N Engl J Med*. 2014;371:2167–2177. doi:10.1056/NEJMoa1410490

71. Su S, Wu YL. Clinical trials of tyrosin kinase inhibitor for lung cancer in China: a review. *J Hematol Oncol*. 2017;10:147. doi:10.1186/s13045-017-0514-z

72. Khan M, Lin J, Liao G, et al. ALK inhibitors in the treatment of ALK positive NSCLC. *Front Oncol*. 2019;8:557. doi:10.3389/fonc.2018.00557

73. Costa DB, Shaw AT, Ou SH, et al. Clinical experience with crizotinib in patients with advanced *ALK*-rearranged non-small-cell lung cancer and brain metastases. *J Clin Oncol*. 2015;33:1881–1888. doi:10.1200/JCO.2014.59.0539

74. Betts K, Song J, Guo J, et al. Real-world treatment patterns and brain metastases development in ALK-positive non-small cell lung cancer. AMCP Managed Care & Specialty Pharmacy Annual Meeting; 2016; San Francisco, CA.

75. Horn L, Infante JR, Reckamp KL, et al. Ensartinib (X-396) in ALK-positive non-small cell lung cancer: results from a first-in-human phase I/II, multicenter study. *Clin Cancer Res*. 2018;24:2771–2779. doi:10.1158/1078-0432.CCR-17-2398

76. Isozaki H, Takigawa N, Kiura K. Mechanisms of acquired resistance to ALK inhibitors and the rationale for treating ALK-positive lung cancer. *Cancers*. 2015;7:763–783. doi:10.3390/cancers7020763

77. Doebele RC, Pilling AB, Aisner DL, et al. Mechanisms of resistance to crizotinib in patients with ALK gene rearranged non-small cell lung cancer. *Clin Cancer Res*. 2012;18:1472–1482. doi:10.1158/1078-0432.CCR-11-2906

78. Dagogo-Jack I, Shaw AT. Crizotinib resistance: implications for therapeutic strategies. *Ann Oncol*. 2016;27(Suppl3):iii42–iii50. doi:10.1093/annonc/mdw141

79. Friboulet L, Li N, Katayama R, et al. The ALK inhibitor ceritinib overcomes crizotinib resistance in non-small cell lung cancer. *Cancer Discov*. 2014;4:662–673. doi:10.1158/2159-8290.CD-13-0646

80. Marsilje TH, Pei W, Chen B, et al. Synthesis, structure-activity relationships, and in vivo efficacy of the novel potent and selective anaplastic lymphoma kinase (ALK) inhibitor 5-chloro-N2-(2-isopropoxy-5-methyl-4-(piperidin-4-yl)phenyl)-N4-(2-(isopropylsulf onyl)phenyl)pyrimidine-2,4-diamine (LDK378) currently in phase 1 and phase 2 clinical trials. *J Med Chem*. 2013;56:5675–5690.

81. Khozin S, Blumenthal GM, Zhang L, et al. FDA approval: ceritinib for the treatment of metastatic anaplastic lymphoma kinase-positive non-small cell lung cancer. *Clin Cancer Res*. 2015;21:2436–2439. doi:10.1158/1078-0432.CCR-14-3157

82. Santarpia M, Altavilla G, Rosell R. Alectinib: a selective, next-generation ALK inhibitor for treatment of ALK-rearranged non-small-cell lung cancer. *Expert Rev Respir Med*. 2015;9:255–268. doi:10.1586/17476348.2015.1009040

83. Kodama T, Hasegawa M, Takanashi K, et al. Antitumor activity of the selective ALK inhibitor alectinib in models of intracranial metastases. *Cancer Chemother Pharmacol*. 2014;74:1023–1028. doi:10.1007/s00280-014-2578-6

84. Masters GA, Temin S, Azzoli CG, et al. Systemic therapy for stage IV non-small-cell lung cancer: American Society of Clinical Oncology Clinical Practice Guideline Update. *J Clin Oncol*. 2015;33:3488–3515. doi:10.1200/JCO.2015.62.1342

85. Novello S, Mazieres J, Oh IJ, et al. Alectinib versus chemotherapy in crizotinib-pretreated anaplastic lymphoma kinase (ALK)-positive non-small-cell lung cancer: results from the phase III ALUR study. *Ann Oncol*. 2018;29:1409–1416. doi:10.1093/annonc/mdx807

86. Huang WS, Liu S, Zou D, et al. Discovery of Brigatinib (AP26113), a phosphine oxide-containing, potent, orally active inhibitor of anaplastic lymphoma kinase. *J Med Chem*. 2016;59:4948–4964. doi:10.1021/acs.jmedchem.6b00306

87. Drilon AE, Ou S-HI, Cho BC, et al. A phase 1 study of the next-generation ALK/ROS1/TRK inhibitor ropotrectinib (TPX-0005) in patients with advanced ALK/ROS1/NTRK+ cancers (TRIDENT-1). *J Clin Oncol*. 2018;36:2513. doi:10.1200/JCO.2018.36.15_suppl.2513

88. Zhi XY, Yu JM, Shi YK. Chinese guidelines on the diagnosis and treatment of primary lung cancer (2015 version). *Cancer*. 2015;122:162.

89. Li X, Zhou Q, Wang X, et al. The effect of low insurance reimbursement on quality of care for non-small cell lung cancer in China: a comprehensive study covering diagnosis, treatment, and outcomes. *BMC Cancer*. 2018;18:683. doi:10.1186/s12885-018-4242-8

90. Yang LL, Zhang XC, Yang XN, et al. Lung cancer treatment disparities in China: A question in need of an answer. *Oncologist*. 2014;19:1084–1090. doi:10.1634/theoncologist.2014-0007

91. Daverman R Prizer Inc. gains China approval of drug for kinase-specific lung cancer. Available from: https://www.biospace.com/article/releases/pfizer-inc-gains-china-approval-of-drug-for-kinase-specific-lung-cancer-/. Accessed June 8, 2019.

92. Hong X, Chen Q, Ding L, et al. Clinical benefit of continuing crizotinib therapy after initial disease progression in Chinese patients with advanced ALK-rearranged non-small-cell lung cancer. *Oncotarget*. 2017;8:41631–41640. doi:10.18632/oncotarget.15892

93. Kim DW, Mehra R, Tan DSW, et al. Activity and safety of ceritinib in patients with *ALK*-rearranged non-small-cell lung cancer (ASCEND-1): updated results from the multicentre, open-label, phase 1 trial. *Lancet Oncol*. 2016;17:452–463. doi:10.1016/S1470-2045(15)00614-2

94. Shaw AT, Kim TM, Crino L, et al. Ceritinib versus chemotherapy in patients with ALK-rearranged non-small-cell lung cancer previously given chemotherapy and crizotinib (ASCEND-5): a randomised, controlled, open-label, phase 3 trial. *Lancet Oncol*. 2017;18:874–886. doi:10.1016/S1470-2045(17)30072-4

95. Crino L, Ahn MJ, De Marinis F, et al. Multicenter phase II study of whole-body and intracranial activity with ceritinib in patients with *ALK*-rearranged non-small-cell lung cancer previously treated with chemotherapy and crizotinib: results from ASCEND-2. *J Clin Oncol*. 2016;34:2866–2873. doi:10.1200/JCO.2015.65.5936

96. Hida T, Nokihara H, Kondo M, et al. Alectinib versus crizotinib in patients with ALK-positive non-small-cell lung cancer (J-ALEX): an open-label, randomised phase 3 trial. *Lancet*. 2017;390:29–39. doi:10.1016/S0140-6736(17)31928-1

97. Camidge DR, Mok T, Gadgeel SM, et al. Updated efficacy and safety data from the global phase III ALEX study of alectinib (ALC) vs crizotinib (CZ) in untreated advanced ALK+ NSCLC. ASCO 2018 Annual Meeting; 2018; Chicago, USA.

98. China National Drug Administration grants rapid approval of Roche's Alecensa (alectinib) as a treatment for ALK-positive lung cancer [press release]. August 20, 2018. Available from: https://www.roche.com/media/releases/med-cor-2018-08-20.htm. Accessed June 8, 2019.

99. Sakamoto H, Tsukaguchi T, Hiroshima S, et al. CH5424802, a selective ALK inhibitor capable of blocking the resistant gate-keeper mutant. *Cancer Cell*. 2011;19:679–690. doi:10.1016/j.ccr.2011.04.004

100. Vavala T, Novello S. Alectinib in the treatment of ALK-positive non-small cell lung cancer: an update on its properties, efficacy, safety and place in therapy. *Ther Adv Med Oncol*. 2018;10:1758835918789364. doi:10.1177/1758835918789364

101. Kodama T, Tsukaguchi T, Satoh Y, et al. Alectinib shows potent antitumor activity against RET-rearranged non-small cell lung cancer. *Mol Cancer Ther*. 2014;13:2910–2918. doi:10.1158/1535-7163.MCT-14-0274

102. Kodama T, Tsukaguchi T, Yoshida M, Kondoh O, Sakamoto H. Selective ALK inhibitor alectinib with potent antitumor activity in models of crizotinib resistance. *Cancer Lett*. 2014;351:215–221. doi:10.1016/j.canlet.2014.05.020

103. Gainor JF, Dardaei L, Yoda S, et al. Molecular mechanisms of resistance to first- and second-generation ALK inhibitors in *ALK*-rearranged lung cancer. *Cancer Discov*. 2016;6:1118–1133. doi:10.1158/2159-8290.CD-16-0596

104. Katayama R, Lovly CM, Shaw AT. Therapeutic targeting of anaplastic lymphoma kinase in lung cancer: a paradigm for precision cancer medicine. *Clin Cancer Res*. 2015;21:2227–2235. doi:10.1158/1078-0432.CCR-14-2791

105. Awad MM, Shaw AT. ALK inhibitors in non-small cell lung cancer: crizotinib and beyond. *Clin Adv Hematol Oncol*. 2014;12:429–439.

106. Mologni L, Ceccon M, Pirola A, et al. NPM/ALK mutants resistant to ASP3026 display variable sensitivity to alternative ALK inhibitors but succumb to the novel compound PF-06463922. *Oncotarget*. 2015;6:5720–5734. doi:10.18632/oncotarget.3122

107. Kinoshita K, Oikawa N, Tsukuda T. Chapter Nineteen - anaplastic lymphoma kinase inhibitors for the treatment of ALK-positive cancers. In: Desai MC, editor. *Annual Reports in Medicinal Chemistry*. Vol. 47. Cambridge (MA): Academic Press; 2012:281–293.

108. European Commission approves Roche's Alecensa (alectinib) as first-line treatment in ALK-positive lung cancer [press release]. December 21, 2017. Available from: https://www.roche.com/investors/updates/inv-update-2017-12-21.htm. Accessed June 8, 2019.

109. Roche receives EU approval of Alecensa (alectinib) for people with previously treated ALK-positive non-small cell lung cancer [press release]. February 21, 2017. Available from: https://www.roche.com/media/releases/med-cor-2017-02-21.htm. Accessed June 8, 2019.

110. FDA approves Genentech's Alecensa (alectinib) as first-line treatment for people with specific type of lung cancer [press release]. November 6, 2017. Available from: https://www.gene.com/media/press-releases/14689/2017-11-06/fda-approves-genentechs-alecensa-alectin. Accessed June 8, 2019.

111. Gadgeel SM, Gandhi L, Riely GJ, et al. Safety and activity of alectinib against systemic disease and brain metastases in patients with crizotinib-resistant *ALK*-rearranged non-small-cell lung cancer (AF-002JG): results from the dose-finding portion of a phase 1/2 study. *Lancet Oncol*. 2014;15:1119–1128. doi:10.1016/S1470-2045(13)70510-2

112. Gadgeel SM, Shaw AT, Govindan R, et al. Pooled analysis of CNS response to Alectinib in two studies of pretreated patients with ALK-positive non-small-cell lung cancer. *J Clin Oncol*. 2016;34:4079–4085. doi:10.1200/JCO.2016.68.4639

113. Chugai Pharmaceutical Co Ltd. Alecensa (alectinib) [Japanese prescribing information]. 2014. Available from: https://chugai-pharm.jp/hc/ss/pr/drug/alc_cap0150/guide/PDF/lg/alc_150_guide_lg.pdf. Accessed January 15, 2019.

114. Genentech USA Inc. Alecensa (Alectinib) [prescribing information]. Available from: https://www.gene.com/download/pdf/alecensa_prescribing.pdf. Accessed January 15, 2019.

115. Muller IB, De Langen AJ, Honeywell RJ, Giovannetti E, Peters GJ. Overcoming crizotinib resistance in *ALK*-rearranged NSCLC with the second-generation ALK-inhibitor ceritinib. *Expert Rev Anticancer Ther*. 2016;16:147–157. doi:10.1586/14737140.2016.1131612

116. Mok TSK, Peters S, Camidge DR, et al. Alectinib (ALC) vs crizotinib (CRZ) in treatment-naïve ALK+ non-small-cell lung cancer (NSCLC): Asian vs non-Asian subgroup analysis of the ALEX study. *Ann Oncol*. 2017;28:X186–X195. doi:10.1093/annonc/mdx075

117. Tamura T, Kiura K, Seto T, et al. Three-year follow-up of an Alectinib phase I/II study in ALK-positive non-small-cell lung cancer: AF-001JP. *J Clin Oncol*. 2017;35:1515–1521. doi:10.1200/JCO.2016.70.5749

118. Fan J, Xia Z, Zhang X, et al. The efficacy and safety of alectinib in the treatment of ALK+ NSCLC: a systematic review and meta-analysis. *Onco Targets Ther*. 2018;11:1105–1115. doi:10.2147/OTT.S156170

119. Yang JC, Ou SI, De Petris L, et al. Pooled systemic efficacy and safety data from the pivotal phase ii studies (NP28673 and NP28761) of Alectinib in ALK-positive non-small cell lung cancer. *J Thorac Oncol*. 2017;12:1552–1560. doi:10.1016/j.jtho.2016.09.002

120. Zhou C, Kim SW, Reungwetwattana T, et al. Alectinib versus crizotinib in untreated Asian patients with anaplastic lymphoma kinase-positive non-small-cell lung cancer (ALESIA): a randomised phase 3 study. *Lancet Respir Med*. 2019;7:437–446. doi:10.1016/S2213-2600(19)30053-0

121. Ou SH, Ahn JS, De Petris L, et al. Alectinib in crizotinib-refractory *ALK*-rearranged non-small-cell lung cancer: a phase II global study. *J Clin Oncol*. 2016;34:661–668. doi:10.1200/JCO.2015.63.9443

122. Toyokawa G, Seto T, Takenoyama M, Ichinose Y. Insights into brain metastasis in patients with ALK+ lung cancer: is the brain truly a sanctuary? *Cancer Metastasis Rev*. 2015;34(4):797–805. doi:10.1007/s10555-015-9592-y

123. Lin JJ, Jiang GY, Joshipura N, et al. Efficacy of alectinib in patients with ALK-positive NSCLC and symptomatic or large CNS metastasis. *J Thorac Oncol*. 2019;14:683–690. doi:10.1016/j.jtho.2018.12.002

124. Gadgeel S, Peters S, Mok T, et al. Alectinib versus crizotinib in treatment-naive anaplastic lymphoma kinase-positive (ALK+) non-small-cell lung cancer: CNS efficacy results from the ALEX study. *Ann Oncol*. 2018;29:2214–2222. doi:10.1093/annonc/mdx807

125. Gadgeel S, Shaw AT, Barlesi F, et al. Cumulative incidence rates for CNS and non-CNS progression in two phase II studies of alectinib in ALK-positive NSCLC. *Br J Cancer*. 2018;118:28–42. doi:10.1038/bjc.2017.395

126. Larkins E, Blumenthal GM, Chen H, et al. FDA approval: alectinib for the treatment of metastatic, ALK-positive non-small cell lung cancer following crizotinib. *Clin Cancer Res*. 2016;22:5171–5176. doi:10.1158/1078-0432.CCR-16-0190

127. Camidge DR, Gadgeel S, Oi SH, et al. MA07.02 Updated efficacy and safety data from the phase 2 NP28761 study of alectinib in ALK-positive non-small-cell lung cancer. *J Thorac Oncol*. 2017;12:S378. doi:10.1016/j.jtho.2016.09.002

128. ARIAD Pharmaceuticals Inc. ALUNBRIG™ (brigatinib) [US pre-scribing information]. Available from: https://www.accessdata.fda.gov/drugsatfda_docs/label/2017/208772lbl.pdf. Accessed February 14, 2018.

129. Seto T, Kiura K, Nishio M, et al. CH5424802 (RO5424802) for patients with *ALK*-rearranged advanced non-small-cell lung cancer (AF-001JP study): a single-arm, open-label, phase 1-2 study. *Lancet Oncol*. 2013;14:590–598. doi:10.1016/S1470-2045(13)70142-6

130. Camidge DR, Bang YJ, Kwak EL, et al. Activity and safety of crizotinib in patients with ALK-positive non-small-cell lung can-cer: updated results from a phase 1 study. *Lancet Oncol*. 2012;13:1011–1019. doi:10.1016/S1470-2045(12)70140-7

131. Blackhall F, Ross Camidge D, Shaw AT, et al. Final results of the large-scale multinational trial PROFILE 1005: efficacy and safety of crizotinib in previously treated patients with advanced/metastatic ALK-positive non-small-cell lung cancer. *ESMO Open*. 2017;2: e000219. doi:10.1136/esmoopen-2017-000219

132. Wu YL, Zhou J, Shi YK, et al. Results of PROFILE1029, a phase III comparison of first-line crizotinib versus chemotherapy in East Asian patients with ALK-positive advanced non-small cell lung cancer. *J Thorac Oncol*. 2018;13:1539–1548. doi:10.1016/j.jtho.2018.06.012

133. Felip E, Orlov S, Park K, et al. ASCEND-3: A single-arm, open-label, multicenter phase II study of ceritinib in ALKi-naïve adult patients (pts) with *ALK*-rearranged (ALK+) non-small cell lung cancer (NSCLC). *J Clin Oncol*. 2015;33:8060. doi:10.1200/jco.2015.33.15_suppl.8060

134. Soria JC, Chiari R, Paz-Ares L, et al. First-line ceritinib versus platinum-based chemotherapy in advanced ALK-rearranged non-small-cell lung cancer (ASCEND-4): a randomised, open-label, phase 3 study. *Lancet*. 2017;389:917–929. doi:10.1016/S0140-6736(17)30123-X

135. Zhang L, Shi Y, Tan DSW, et al. ASCEND 6: single-arm, open label, multicenter phase 1/2 study of ceritinib in Chinese pts with advanced ALK-rearranged (ALK+) non-small cell lung cancer (NSCLC) previously treated with crizotinib. *Ann Oncol*. 2016;27(Supplement_9):ix139–ix156. doi:10.1093/annonc/mdw141

136. Gettinger SN, Bazhenova LA, Langer CJ, et al. Activity and safety of brigatinib in *ALK*-rearranged non-small-cell lung cancer and other malignancies: a single-arm, open-label, phase 1/2 trial. *Lancet Oncol*. 2016;17:1683–1696. doi:10.1016/S1470-2045(16)30392-8

137. Camidge DR, Ahn MJ, Yang JC, et al. Brigatinib versus crizotinib in ALK-positive non–small-cell lung cancer. *N Engl J Med*. 2018;379:2027–2039. doi:10.1056/NEJMoa1810171

138. Shaw AT, Felip E, Bauer TM, et al. Lorlatinib in non-small-cell lung cancer with ALK or ROS1 rearrangement: an international, multicentre, open-label, single-arm first-in-man phase 1 trial. *Lancet Oncol*. 2017;18:1590–1599. doi:10.1016/S1470-2045(17)30072-4

139. Solomon BJ, Besse B, Bauer TM, et al. Lorlatinib in patients with ALK-positive non-small-cell lung cancer: results from a global phase 2 study. *Lancet Oncol*. 2018;19:1654–1667. doi:10.1016/S1470-2045(18)30144-X

140. Planchard D, Popat S, Kerr K, et al. Metastatic non-small cell lung cancer: EMSO clinical practice guidelines for diagnosis, treatment and follow-up. *Ann Oncol*. 2018;29(Supplement_4):iv192–iv237. doi:10.1093/annonc/mdx807

141. Ettinger DS, Alsner DL, Wood DE, et al. NCCN guidelines insights: non-small cell lung cancer, version 5.2018. *J Natl Compr Canc Netw*. 2018;16:807–821. doi:10.6004/jnccn.2018.0062