ORIGINAL RESEARCH miR-100 Inhibits Cell Growth and Proliferation by Targeting HOXAI in Nasopharyngeal Carcinoma

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Background: Increasing evidence indicates that the dysregulation of miRNAs plays a vital role in tumorigenesis and progression of nasopharyngeal carcinoma (NPC). Thus, it is necessary to further investigate the function and mechanism of miRNAs in NPC.

Methods: miR-100 expression was analyzed using publicly available databases and then tested using quantitative RT-PCR in NPC tissues and cell lines. MTT and colony formation assays and xenograft tumor model were used to test the NPC cell growth and proliferation abilities while modulating miR-100 expression. The target of miR-100 was predicted with TargetScan and validated with luciferase reporter assay, quantitative RT-PCR, and Western blot.

Results: The expression of miR-100 was significantly reduced in NPC tissues and cell lines. Overexpression of miR-100 obviously suppressed NPC cell growth and proliferation, whereas silencing miR-100 promoted NPC cell growth and proliferation in vitro. HOXA1 (homeobox A1) was validated as a direct target of miR-100, and restoring HOXA1 expression could reverse the inhibitive effect of miR-100 on NPC cell growth and proliferation. The mRNA and protein expression of HOXA1 was increased in NPC cell lines. Furthermore, ectopic expression of miR-100 inhibited xenograft tumor growth in vivo.

Conclusion: Taken together, our findings suggest that miR-100 could suppress NPC growth and proliferation through targeting HOXA1, providing a novel target for the miRNAmediated therapy for patients with NPC in the future.

Keywords: miR-100, nasopharyngeal carcinoma, proliferation, invasion, HOXA1

Introduction

Nasopharyngeal carcinoma (NPC) is a malignant epithelial tumor with the highest incidence in China, which accounts for 47.7% of the new cases worldwide.¹ As its unique anatomical location, radiotherapy is the primary treatment for NPC patients. During the past years, with the use of intensity-modulated radiation therapy and its combination with chemotherapy, the 5-year survival rate of NPC patients has been significantly improved. However, about 30% of NPC patients eventually die of relapse or metastasis.² Therefore, it is very urgent to carry out relevant studies to clarify the underlying mechanism of NPC tumorigenesis and development and provide new molecular targets for future treatment.

MiRNAs are small non-coding RNAs, and it can enhance mRNA degradation or inhibit its translation through base pairing with the 3'-untranslated region (3'-UTR) of mRNA.³ Many studies demonstrate that miRNA expression changes significantly in various tumors, making it as biomarkers for tumor diagnosis, subclassify, and prognosis prediction.⁴⁻⁶ More importantly, the changed miRNAs play very important roles in the process of tumorigenesis, development, and progression.⁷⁻⁹ It has

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been also found that miRNA expression is dysregulated in NPC,^{10–12} and the dysregulated miRNAs can promote NPC cell proliferation, invasion, and angiogenesis, including miR-125b, miR-506, miR-142-3p, miR-23a, and so on.^{13–16} Thus, previous studies indicate that miRNA plays vital roles in NPC carcinogenesis and development. It also should be noted that further exploration of the function and mechanism of miRNA involved in NPC development and progression would provide novel therapeutic targets.

Based on analyzing several publicly available microarray data, we found that miR-100 was obviously decreased in NPC. It has been reported that miR-100 is dysregulated in many types of cancers, and it functions as either a tumor suppressor or promoter in different tumor types.^{17,18} In NPC, miR-100 was found to be decreased and it can suppress NPC cell migration and invasion through targeting IGF1R.¹⁹ However, no study has elucidated the function and mechanisms of miR-100 in NPC growth and proliferation. Therefore, in our present study, we performed functional studies and found that overexpression of miR-100 inhibited NPC cell growth and proliferation in vitro, while silencing miR-100 had an opposite function. Mechanism studies indicated that HOXA1 was a direct target of miR-100, and the restoration of HOXA1 expression reversed the inhibitive effect of miR-100 overexpression on NPC cell growth and proliferation. Our findings suggest that miR-100 functions as a tumor suppressor in NPC, which might serve as a treatment target for NPC patients.

Methods

Cell Culture

The NPC cell lines, including CNE1, CNE2, HNE1, HONE1, SUNE1, and C666-1, were maintained in RPMI-1640 medium (Gibco, Grand Island, NY, USA) supplemented with 10% fetal bovine serum (Gibco); and the human immortalized nasopharyngeal epithelial cell line NP69 was maintained in KSFM medium (Gibco) supplemented with bovine pituitary extract (BD Biosciences, San Diego, CA, USA) in a humidified atmosphere at 37°C with 5% CO₂. All NPC cell lines and NP69 were generously provided by Professor Zhiqiang Xiao (Central South University), and the use of these cell lines was approved by the Institutional Ethical Review Board of the Brain Hospital of Human Province.

RNA Isolation and Quantitative RT-PCR

Total RNA was extracted using the TRIzol Reagent (Invitrogen, Carlsbad, CA, USA), and then the cDNA was synthesized using a reverse transcriptase (Promega, Madison, WI, USA). Quantitative PCR was conducted using the SYBR Green reagents (Invitrogen). The primers for miR-100 were purchased from the RiboBio (Guangzhou), and for HOXA1 was synthesized: GGCTCGCCTCAATACATTCAC (forward) and CGTACTCTCCAACTTTCCCTG (reverse). All of the experiments were conducted in triplicate and normalized to the expression of U6 or GAPDH. The relative expression was calculated with the $2^{-\Delta\Delta CT}$ equation.

Transient Transfection and Stably Transfected Cell Line Construction

The miR-100 mimics or inhibitor and its controls were purchased from the GenePharma (Suzhou). The HOXA1 plasmid and its empty vector were purchased from the FulenGen (Guangzhou). Cells were transfected using the Lipofectamine 2000 reagent (Invitrogen) and harvested for assays after 48 hrs. The sequence of pri-miR-100 was amplified and cloned into the pSin-EF2-puromycin lentiviral plasmid (Addgene, Cambridge, MA, USA). The lenti-miR-Ctrl or lenti-miR-100 was used to transfect SUNE1 cells and then selected using puromycin.

MTT and Colony Formation Assay

Cells were seeded into 96-well plates with a density of 1000 NPC cells per well, and then incubated for 0–4 days. On each day, the cells were stained with MTT dye (Sigma, St. Louis, MO, USA), and the absorbance was tested at 490 nm using a spectrophotometer. In addition, cells were seeded into 6-well plates with a density of 400 cells and cultured for 10 days. Colonies were fixed and stained, and then counted with an inverted microscope.

Luciferase Reporter Assay

The wild type (WT) and mutant (MT) 3'-UTR sequences of HOXA1 were amplified and inserted into the psiCHECKTM vector (Promega). Then, the reporter plasmids, together with p-TK Renilla plasmid and miR-100 mimics or control were used to transfect NPC cells using Lipofectamine 2000 (Invitrogen). Cells were harvested 48 h later, and the luciferase signals were tested using the Dual-Luciferase Reporter Assay System (Promega).

Western Blot

Total protein was extracted from NPC cells with RIPA buffer, separated with 9% gels, and then transfected to PVDF membranes (Millipore, Billerica, MA, USA). The membranes were firstly incubated with anti-HOXA1 antibody (Abcam, Cambridge, MA, USA), and then with secondary antibody (Sigma). The α -tubulin was used as a loading control, and the bands were obtained with enhanced chemiluminescence.

Xenograft Tumor Growth

Female BALB/c nude mice aged 3~4 weeks were purchased from the Charles River Laboratories (Beijing). SUNE1 cells stably expressing miR-100 or miR-Ctrl were injected into the dorsal flank of mice. The tumor size was measured every 3 days for the calculation of the tumor volume. On day 30, the mice were sacrificed, and the tumors were weighted. All animal study protocols were performed following the guidelines of the Experimental Animal Care and Use Ethics Committee of the Brain Hospital of Hunan Province.

Statistical Analyses

All experiments were conducted at least three times, and all results were shown as mean \pm SD. SPSS 22.0 (SPSS

Inc., Chicago, IL, USA) was used for data analysis by comparing with Student's *t*-test, and p < 0.05 was considered as a significant difference.

Results

miR-100 Is Reduced in NPC Clinical Samples and Cell Lines

We firstly analyzed the biggest publicly available NPC microarray data (GSE32960) and found that miR-100 was significantly reduced in NPC tissues (n=312) compared with normal nasopharynx tissues (n=18) (Figure 1A, p<0.001). Then, miR-100 was verified to be reduced in NPC tissues based on the other two publicly available microarray data with the accession number GSE36682 (Figure 1B, p<0.001) and GSE43039 (Figure 1C, p=0.049). In addition, we detected the miR-100 expression in the immortalized nasopharyngeal epithelial cell line NP69 and six NPC cell lines using quantitative RT-PCR, and found that miR-100 was obviously decreased in all NPC cell lines (Figure 1D, all p<0.05). These results indicate that miR-100 is reduced in NPC tissue samples and cell lines.





Overexpression of miR-100 Inhibits NPC Cell Growth and Proliferation

We transiently transfected SUNE1 and HONE1 cells with miR-100 mimics or its control (miR-Ctrl), and confirmed the transfection efficiency with quantitative RT-PCR (Figure 1E, all p<0.05). Then, we did functional experiments including MTT and colony formation assays. MTT assay showed that NPC cells transfected with miR-100 mimics grew more slowly than cells transfected with miR-Ctrl (Figure 2A and B, p<0.05). Colony formation assay found that NPC cells transfected with miR-100 mimics formed fewer and smaller colonies than cells transfected with miR-100 mimics formed fewer and smaller colonies than cells transfected with miR-100 mimics formed fewer and smaller colonies than cells transfected with miR-Ctrl (Figure 2C and D, p<0.05).

These results indicate that overexpression of miR-100 inhibits NPC cell growth and proliferation.

Silencing miR-100 Promotes NPC Cell Growth and Proliferation

We also transiently transfected SUNE1 and HONE1 cells with miR-100 inhibitor or its control (miR-Ctrl) and verified the transfection efficiency with quantitative RT-PCR (Figure 1F, p<0.05). Then, we also conducted the same functional experiments as mentioned above. We found that the reduction of miR-100 expression distinctly promoted SUNE1 and HONE1 cell growth and increased their colony formation rates (Figure 3A–D, p<0.05). These results



Figure 2 Overexpression of miR-100 inhibits NPC cell growth and proliferation. (A, B) Cell growth viabilities of SUNE1 and HONE1 cells transfected with miR-100 mimics or miR-Ctrl determined by the MTT assay. (C-D) Representative images and quantification of colonies of SUNE1 and HONE1 cells transfected with miR-100 mimics or miR-Ctrl determined by the colony formation assay. Data are presented as the mean \pm SD, and the p values were calculated using the Student's t-test; * p<0.05.



Figure 3 Silencing of miR-100 promotes NPC cell growth and proliferation. (A, B) Cell growth viabilities of SUNEI and HONEI cells transfected with miR-100 inhibitor or miR-Ctrl determined by the MTT assay. (C, D) Representative images and quantification of colonies of SUNEI and HONEI cells transfected with miR-100 inhibitor or miR-Ctrl determined by the colony formation assay. Data are presented as the mean \pm SD, and the p values were calculated using the Student's t-test; * p<0.05.

indicate that silencing miR-100 promotes NPC cell growth and proliferation.

HOXAI Is a Direct Target of miR-100 and Its Expression Is Increased in NPC

We predicted the potential target of miR-100 using the online database TargetScan and selected HOXA1 for further validation because of its crucial effect on cell growth. Figure 4A shows the putative binding site between miR-100 and HOXA1. Luciferase report assay found that over-expression of miR-100 could inhibit the luciferase activities of the HOXA1 3'-UTR WT reporter plasmid, but not the HOXA1 3'-UTR MT reporter plasmid (Figure 4B, p<0.05). Then, quantitative PCR and Western blot showed that

overexpression of miR-100 could suppress the mRNA and protein expression of HOXA1 (Figure 4C and D, p<0.05). Furthermore, HOXA1 mRNA and protein were upregulated in NPC cell lines (Figure 4E and F). These results indicate that HOXA1 is a direct target of miR-100 in NPC.

Restoration of HOXA1 Reverses the Inhibitive Effects of miR-100 in NPC

We then investigate whether HOXA1 was essential for the regulatory effect of miR-100 on NPC cell growth and proliferation. We transiently co-transfected SUNE1 and HONE1 cells with miR-100 mimics and HOXA1 plasmid or its empty vector, and then did MTT and colony formation assays. The functional results showed that the ectopic expression of

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Figure 4 HOXA1 is a direct target of miR-100 and its expression is decreased in NPC (A) Wild-type (WT) and mutant (MT) 3'-UTR of HOXA1 with putative bind site of miR-100. (B) Relative luciferase activities of SUNE1 and HONE1 cells determined by the luciferase reporter assay. (C, D) The mRNA and protein expression of HOXA1 in SUNE1 and HONE1 cells transfected with miR-100 mimics or miR-Ctrl. (E, F) The mRNA and protein expression of HOXA1 in NPC cell lines determined by quantitative RT-PCR and Western blot. The data are presented as the mean \pm SD; p values were calculated using Student's t-test; * p<0.05.

HOXA1 reversed the inhibitive effects of miR-100 on NPC cell growth and proliferation (Figure 5A–D, p<0.05). These results indicate that HOXA1 is a functional target of miR-100 in NPC.

Overexpression of miR-100 Inhibits NPC Xenograft Tumor Growth

We constructed xenograft tumor growth model through injecting SUNE1 cells that stably expressing miR-100 or miR-Ctrl into the dorsal flank of nude mice, and all of the mice formed xenograft tumors (Figure 6A). The tumors formed in the miR-100 overexpression group grew more slowly and smaller than the miR-Ctrl group (Figure 6B, p<0.05). In addition, the tumor size and weight were significantly different between the two groups (Figure 6C and D, p<0.05). The expression of miR-100 was increased in xenograft tumor tissues of miR-100 overexpression group than the miR-Ctrl group (Figure 6E, p<0.05). These results indicate that overexpression of miR-100 inhibits NPC xenograft tumor growth.

Discussion

Recurrence and metastasis are two major causes of treatment failure and death for patients with NPC, and it is urgent to better understanding the molecular mechanisms related to NPC tumorigenesis and progression, which would guide a more personalized therapy for NPC. In our present study, we reported that miR-100 was decreased in NPC and functioned as a tumor suppressor by inhibiting NPC cell growth and proliferation.

Recently, miRNAs have been reported to be dysregulated in NPC based on genome-wide profiling.^{10,12} The dysregulated miRNAs play an important role in NPC tumorigenesis and progression by promoting NPC cell proliferation, invasion, and angiogenesis.^{13–16} It has been found that miR-125b is upregulated in NPC, and it can promote NPC cell proliferation and inhibit cell apoptosis by targeting A20 and activating the NFkB signaling pathway.¹³ miR-506 can inhibit NPC tumor growth and metastasis through inactivating the Wnt/β-catenin signaling pathway by targeting LHX2.¹⁴ EZH2-DNMT1mediated epigenetic silencing of miR-142-3p promotes NPC cell invasion and metastasis by targeting ZEB2.15 miR-23a can promote NPC cell growth, migration, and angiogenesis by targeting TSGA10.¹⁶ Here, in our present study, we found that miR-100 was obviously decreased in NPC tissue samples by analyzing three publicly available microarray data. Functional experiments verified that miR-100 could suppress NPC cell growth and proliferation in vitro and inhibit xenograft



Figure 5 Restoration of HOXA1 reverses the inhibitive effect of miR-100 in NPC (A, B) Cell growth viabilities of SUNE1 and HONE1 cells transfected with miR-100 mimics and HOXA1 plasmids or empty vector determined by the MTT assay. (C, D) Representative images and quantification of colonies of SUNE1 and HONE1 cells transfected with miR-100 mimics and HOXA1 plasmids or empty vector determined by colony formation assay. Data are presented as the mean \pm SD, and the p values were calculated using the Student's *t*-test; * p<0.05.

tumor growth in vivo. It is worth noting that a recent study reports that miR-100 can inhibit NPC cell migration and invasion by targeting IGF1R.¹⁹ These findings indicate that miR-100 plays important roles in NPC development and progression.

miR-100, as a member of the miR-99 family, has been reported to be dysregulated in many types of cancers, and it can function as either a tumor suppressor or promoter, which depends on the tumor types and microenvironment.^{17,18} Downregulation of miR-100 has been found in esophageal squamous cell carcinoma, non-small cell lung cancer, breast cancer, and so on.^{20–22} On the other hand, upregulation of

miR-100 is observed in small cell lung cancer, renal cell carcinoma, and pancreatic adenocarcinoma, and so on.^{23–25} It has been also found that miR-100 play vital roles in various biological processes, such as cell proliferation, apoptosis, cell cycle, migration, differentiation, and angiogenesis.^{17,18} For example, miR-100 inhibits breast cancer proliferation and survival by targeting IGF2.²² miR-100 inhibits gastric cancer tumor growth and metastasis and growth by targeting ZBTB7A.²⁶ miR-100 promotes cell differentiation and survival in acute myeloid leukemia by targeting RBSP3.²⁷ miR-100 promotes hepatocellular carcinoma cell metastasis by enhancing ICMT-Rac1 signaling.²⁸ In this study, we found



Figure 6 Overexpression of miR-100 inhibits NPC xenograft tumor growth (A) Xenograft tumor growth models in nude mice were constructed by injecting with SUNEI cells stably expressing miR-100 or miR-Ctrl, and the xenograft tumors were formed. (B) The growth curves of the tumor volumes. (C) Representative images of the excised tumors. (D) The weights of the excised tumors. (E) Relative expression of miR-100 in xenograft tumors of the miR-100 overexpression group and miR-Ctrl group. Data are presented as the mean \pm SD, and the p values were calculated using the Student's t-test; * p<0.05.

that miR-100 was decreased in NPC and functioned as a tumor-suppressive miRNA, enriching the understanding of the function and mechanism of miR-100 in tumors.

As we have known, miRNAs exert their function by base-pairing with the 3'-UTR of their target genes.³ Many target genes of miR-100 have been identified and verified, including mTOR, IGFR, PLK1, AKT1, RAP1B, FGFR3, and so on.^{29–34} As each miRNA can regulate multiple different target genes, our study predicted HOXA1 as a potential direct target of miR-100 with the public available database TargetScan. We then verified HOXA1 as the target of miR-100 in NPC using luciferase report assay, quantitative RT-PCR and Western blot. Similarly, HOXA1 was also found to be a direct target of miR-100 in lung cancer and breast cancer.^{23,35} It should be noted that HOXA1 can be regulated by several other miRNAs, such as miR-30b, miR-99a, and miR-577, and so on.^{36–38} More importantly, HOXA1 has been reported as an oncogene in various types of cancers.^{39–41} In our study, we found that HOXA1 was increased in NPC and it was regulated by miR-100, thereby enriching the understanding of miR-100/HOXA1 signaling pathway involvement in NPC tumorigenesis and progression.

In conclusion, our study revealed that miR-100 expression was reduced, and functioned as a tumor suppressor in NPC. Ectopic expression of miR-100 could suppress NPC cell growth and proliferation in vitro, and inhibit xenograft tumor growth in vivo by targeting HOXA1. Our findings make a better understanding of the mechanisms involved in miR-100-mediated NPC tumorigenesis and progression, further providing an opportunity for the exploration of novel miRNA-based targeted treatments for NPC patients.

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Author Contributions

TZ and ST designed the study. WH, YH, CJ, YZ, WL, and WZ carried out all of the experiments, prepared figures and drafted the manuscript. All authors participated in the data analysis and the interpretation of all of the results, drafting or revising the article, gave final approval of the version to be published, and agree to be accountable for all aspects of the work.

Disclosure

The authors report no conflicts of interest in this work.

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