

Turbidimetric method for evaluation of photocatalytic activities of suspended fine particles

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Abstract: A spectrophotometer with special cuvette was developed for evaluating the photocatalytic activities of suspended fine particles. The spectrophotometer can continuously irradiate UV light using LED to the sample solution, and changes in the absorbance at 664 nm during photocatalytic degradation of methylene blue (MB) were monitored continuously. From the onset of MB degradation, the absorbance decreased and reached a steady value at the end of the reaction. This process was expressed by first order kinetics and the photocatalytic activities of various fine particles could be evaluated quantitatively based on the reaction rate constant (k). The effect of photocatalysis using various TiO₂ fine particles on the physiological activities of *Euglena gracilis* was related with k value.

Keywords: photocatalyst, fine nano sized particles, specialized spectrophotometer, *Euglena gracilis*, rate constant

Introduction

The use of nano sized fine particles is of increasing importance because they are suitable for manifold applications in the fields of electronics,¹ sensing,² catalysis,³ and medical,⁴ etc. Some types of particles (for example TiO₂) show photocatalytic activities. Many kinds of photocatalytic fine particles with various activities are commercially produced and widely used for various applications. They are used as photocatalysts in environmental decontamination of air, soil, and water,⁵⁻⁷ pigments,⁸ and as cosmetic additives.⁹ Large quantities of the fine particles are discharged to the environment intentionally or unintentionally in the course of their production, use, and disposal. These will inevitably lead to pollution of both the biotic and abiotic components of the environment. Reactive oxygen species (ROS) produced by photocatalysis cause inactivation of various organisms.¹⁰⁻¹³ Studies on the potential effects of these photocatalysts on human health and on the environment are therefore very important.^{13,14} In order to assess the potential risks of photocatalytic fine particles or to efficiently utilize them in various fields, it is very necessary to develop methods for quantitative evaluation of their photocatalytic activities. Many kinds of fine particles with various photocatalytic activities are commercially produced but there are few reports of their relative photocatalytic activities.¹⁵

In this study, an apparatus was developed for quantitative monitoring of photocatalytic reaction of nano fine particles based on spectrophotometry. With this apparatus, it was possible to evaluate the photocatalytic activities of various fine particles on the basis of the reaction rate constant (k) in first order kinetic. The effect of photocatalysis using TiO₂

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fine particles, which measured k values, on the physiological activities of *Euglena gracilis* was also investigated.

Materials and methods

Chemicals

Titanium oxide anatase form [ANA] (100% anatase crystalline form, average particle size = 50 nm, Wako Pure Chemical Industries, Japan), titanium oxide rutile form [RU] (100% rutile crystalline form, average particle size = 100 nm, Wako) titanium oxide amorphous [AM] (average diameter = 40 nm, Wako); AP25 (20% rutile 80% anatase crystalline form, average particle size = 40 nm, Japan Aerosil Co, Tokyo, Japan), STS-01 (photocatalytic sol, average particle size = 7 nm, Ishihara Sangyo Kaisha, Ltd, Osaka, Japan), and STS-02 (photocatalytic sol, average particle size = 7 nm, Ishihara Sangyo) were used in this study. Methylene blue (MB) and Daigo's artificial seawater SP (in mg per liter: $\text{MgCl}_2 \cdot 6\text{H}_2\text{O}$, 9, 474; $\text{CaCl}_2 \cdot 2\text{H}_2\text{O}$, 1, 326; Na_2SO_4 , 3, 505; KCl , 597; NaHCO_3 , 171; KBr , 85; $\text{Na}_2\text{B}_4\text{O}_7 \cdot 10\text{H}_2\text{O}$, 34; SrCl_2 , 12; NaF , 3; LiCl , 1; KI , 0.07; $\text{CoCl}_2 \cdot 6\text{H}_2\text{O}$, 0.0002; $\text{AlCl}_3 \cdot 6\text{H}_2\text{O}$, 0.008; $\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$, 0.005; $\text{Na}_2\text{WO}_4 \cdot 2\text{H}_2\text{O}$, 0.002; $(\text{NH}_4)_6\text{Mo}_7\text{O}_{24} \cdot 4\text{H}_2\text{O}$, 0.02; $\text{MnCl}_2 \cdot 4\text{H}_2\text{O}$, 0.0008; NaCl , 20, 747) were purchased from Wako.

Cultivation of *Euglena gracilis*

E. gracilis IAM E-6 (strain Z), obtained from the algal collection of the Institute of Applied Microbiology, University of Tokyo, Japan was used in this study as a model micro-algae. *E. gracilis* produces antioxidant such as alpha-tocopherol. Modified Hutner medium was used for the cultivation as described previously.¹⁶ The medium was sterilized by autoclaving at 121°C for 15 min. To prevent deactivation by heat, filter-sterilized vitamin B12 was added to the medium after sterilization. Cultivation was carried out by inoculating *E. gracilis* into a 500 mL Erlenmeyer flask containing 100 mL medium and the flask was then incubated under illumination by black light (National Electric, Tokyo) on rotary shaker (120 rpm) at 25°C. The distance between the flask and the black light was adjusted to give a light intensity of 1.0 mW/cm² at the surface of the flask. The ANA, AP25, or RU (100 mg/L) was added to the *E. gracilis* cell culture.

Analytical methods

The alpha-tocopherol was extracted from the cell according to the methods.¹⁷ *E. gracilis* cells harvested by centrifugation at $1,000 \times g$ for 5 min were washed with N_2 saturated distilled water before the addition of 0.1 mL of 50 mM L-ascorbic acid and extracted with 3.75 mL of chloroform-methanol

(1:2 v/v) by shaking vigorously for 20 min in N_2 . The mixture was centrifuged at $1,000 \times g$ for 5 min to remove cell debris. The extraction was repeated two times. The cell debris was extracted with 4.75 mL of chloroform-methanol-water (1:2:0.8 v/v). An equal volume of distilled water was added to combined extracts and mixed gently. The chloroform layer was separated from the water-methanol layer after cooling on ice and centrifuged at $1,000 \times g$ for 5 min, and evaporated to dryness at room temperature under a stream of N_2 gas to obtain the lipid fraction (the alpha-tocopherol content of the cells was determined from the fraction). Vitamin E homolog kit for biochemistry and analysis (Eisai, Tokyo, Japan) was used for the preparation of the calibration curve. The alpha-tocopherol was measured using high-performance liquid chromatography equipped with an Intelligent Spectrofluorometer 821-FP (Nihon Bunko, Tokyo, Japan) and packed silica gel column (150A-5, 4.6×250 mm, GL Science, Tokyo, Japan). The fluorescent lamp was used for the detection of alpha-tocopherol. The wavelengths were $\text{Ex} = 295$ nm and $\text{Em} = 340$ nm. A mixture of *n*-hexane, 1,4-dioxane, and ethanol (97.6:2.0:0.4 v/v) was used as the elute at a flow rate of 1.5 mL/min. The column temperature was 40°C. Dry cell weight was determined according to the method described previously.¹⁶ The total chlorophyll (a + b) concentration was determined according to the method¹⁸ using N, N'-dimethylformamide (DMF). Cells were extracted with DMF overnight. The cells were removed by centrifugation ($1,000 \times g$ for 5 min), and the absorbance of the supernatant was measured using a spectrophotometer (V-550, JASCO, Tokyo, Japan). Total chlorophyll (a + b) concentration was calculated as $\text{Chls a + b (mg/mL in DMF)} = 17.67 A_{646.8} + 7.12 A_{663.8}$, where $A_{646.8}$ and $A_{663.8}$ are the absorbance at wavelengths of 646.8 nm and 663.8 nm, respectively.

All the experiments were performed in triplicate. Although the results of representative single experiments are shown in Figures 2, 3, and 4, they were confirmed to be highly reproducible (there was less than 2.5% deviation in the results).

Results

The spectrophotometer with special cuvette

An apparatus for evaluating the photocatalyst activity of suspended fine particles was developed. It comprised a spectrophotometer equipped with a special cuvette. Figure 1 shows a schematic diagram of the system. The characteristics of this apparatus are as follows; (a) UV light (365 nm) was

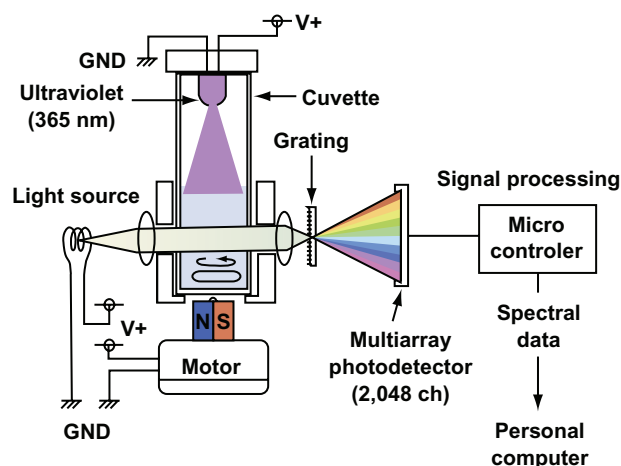


Figure 1 Schematic diagram of an apparatus for evaluating the photocatalytic activity of suspended fine particles in the liquid.

irradiated continuously at 0.7 mW/cm^2 in the surface of the sample by LED, which produces very low heat; (b) a magnetic stirrer is situated under the cuvette (diameter = 10 mm, length = 43 mm, working volume = 2.0 mL, the temperature was kept at 25°C) and the content can be mixed with a mini magnetic bar (diameter = 2 mm, length = 5 mm, the agitation speed = 600 rpm); (c) the absorbance and spectra (wavelength from 400 nm to 700 nm) can be monitored continuously by means of personal computer. This apparatus enables the samples in the cuvette to be mixed homogeneously, giving a stable absorbance over a long period of time. It is therefore possible to obtain stable absorbance readings for even heterogeneous fine particles with high specific gravity (it was confirmed in various TiO_2 solutions without MB under UV irradiation or no irradiation [data not shown]). The degradation of MB was used to evaluate photoactivity of TiO_2 . Changes in the spectra (400–700 nm) of photocatalytic decomposition of MB were monitored. It was observed that the absorbance decreased with

time at each wavelength (data not shown). A wavelength of 664 nm was selected as the optimal wavelength for monitoring MB decomposition (Otsuka-Yao-Matsuo et al used the same wavelength for the measurement of MB decomposition).¹⁹ The absorbance at 664 nm of the sample solution (MB + TiO_2) was not allowed to exceed to 1.9, which was the limit for possible quantitative evaluation.

Figure 2 shows changes in the absorbance at 664 nm during photocatalytic decomposition of MB (5 mg/L) using ANA (125 mg/L). Before UV irradiation, the A_{664} did not change. After UV irradiation, the A_{664} decreased sharply due to photocatalytic decomposition of MB and then leveled off after about 300 min. The above results showed that the process of photocatalytic decomposition of MB by TiO_2 could be monitored optically, making it possible to use changes in the A_{664} to measure the rate of reaction and to estimate the end of reaction.

Photocatalytic activities of TiO_2 fine particles depend to a large extent on their crystalline form. In the case of the photoinactive RU (125 mg/L) or AM (125 mg/L) suspended in MB (5 mg/L) solution at 25°C , the A_{664} did not change under UV irradiation (data not shown). Figure 3 shows the changes in A_{664} during photocatalytic reaction with ANA and AP25. The photocatalytic activity of ANA in the distilled water is slightly higher than that of AP25. In artificial seawater, there was no photocatalytic activity of AP25, while the activity of ANA decreased significantly (Figure 3).

Evaluation of the photocatalytic activities of various fine particles based on reaction rate constant

The degradation of MB by photocatalysis is a first order reaction and can be expressed by Equation 1:

$$\ln (At - As)/(A_0 - As) = -kt \quad (1)$$

where t = time (min), k = a constant (min^{-1}), A_0 = the initial absorbance, At = the absorbance at time t , and As = absorbance at the end of the process. The value of $\ln (At - As)/(A_0 - As)$ was plotted against time during the degradation of MB by photocatalysis (Figure 4). The major part of the reaction (about 70%) can be described by simple first order kinetics. The rate constant (k) was calculated from the initial straight line. Table 1 shows a comparison of the k values for various photocatalysts. The k values varied depending on the suspending liquid and on the type of photocatalysts. The photocatalytic activity of ANA in artificial seawater decreased

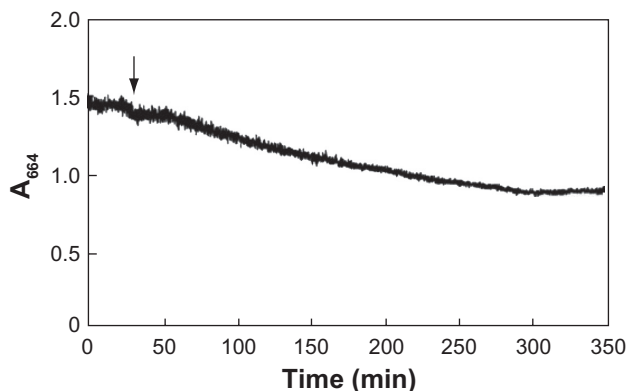


Figure 2 Changes in absorbance at 664 nm during photocatalytic reaction. ANA (TiO_2 anatase form) 125 mg/L, methylene blue 5 mg/L in the distilled water at 25°C . The arrow indicates the start of UV (365 nm) irradiation.

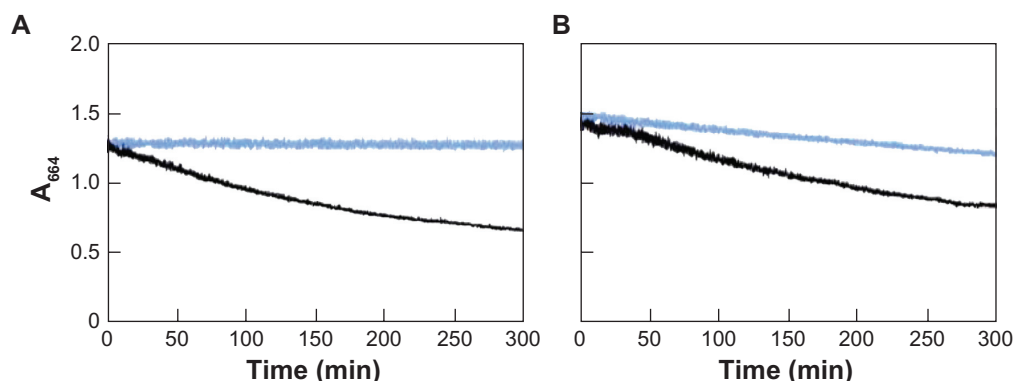


Figure 3 Changes in absorbance at 664 nm during photocatalytic reaction. **A)** ANA 125 mg/L, MB 5 mg/L at 25°C, **B)** AP25 125 mg/L, MB 5 mg/L at 25°C. Black lines are in the distilled water, Blue lines: in artificial sea water.

to one third of that in ultrapure water. The k value of STS-01 is 3.5 times larger than that of ANA or AP25. By using the values of k , it was possible to quantitatively evaluate and compare the photocatalytic activities of various fine particles.

Effect of photocatalysis using various TiO_2 fine particles on the physiological activities of *E. gracilis*

The effect of photocatalysis using various TiO_2 fine particles on the physiological activities of *E. gracilis* was investigated (Figure 5). The k values of ANA (Figure 5C), AP25 (Figure 5D), and RU (Figure 5E) in the *E. gracilis* cell culture medium were determined on the basis of MB degradation by

using the apparatus described earlier. The k values of ANA, AP25, and RU in the medium were $7.1 \pm 0.09 \times 10^{-3} [\text{min}^{-1}]$, $5.1 \pm 0.07 \times 10^{-3} [\text{min}^{-1}]$, and $0 [\text{min}^{-1}]$, respectively. For the distilled water, the k value for ANA was $7.65 \pm 0.15 \times 10^{-3} [\text{min}^{-1}]$, for AP25, $8.05 \pm 0.12 \times 10^{-3} [\text{min}^{-1}]$, while for RU, $0 [\text{min}^{-1}]$ as shown in Table 1. The k values decreased due to the salts in the medium.

E. gracilis was cultivated for 72 hours in a medium containing RU (Figure 5E). The cell growth, chlorophyll content, and alpha-tocopherol content were the same as those obtained in a medium without RU (Figure 5B). On the other hand, when ANA (Figure 5C) or AP25 (Figure 5D) was added to the medium, the cell growth was suppressed, chlorophyll content was the same, while alpha-tocopherol contents of *E. gracilis* were 1.3 and 1.2 times higher than that of the control, respectively.

Discussion

A spectrophotometer with special cuvette described here makes it possible to monitor continuously and the photocatalytic decomposition of MB by suspended fine particles using small amount of sample solution (2.0 mL)

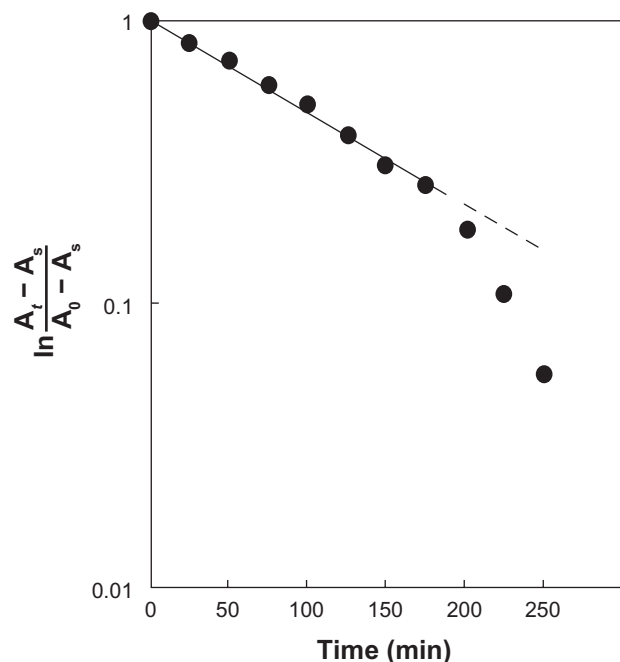


Figure 4 Logarithmic plot of $\ln(A_t - A_s)/(A_0 - A_s)$ against time during photocatalytic reaction. Condition: ANA 125 mg/L, MB 5 mg/L in the distilled water at 25°C (the data of Figure 2 was used).

Table 1 k values for various photocatalysts

Photocatalysts	Suspending liquid	$k \times 10^{-3} (\text{min}^{-1})$
ANA	distilled water	7.65 ± 0.15
AP25	distilled water	8.05 ± 0.12
ANA	artificial seawater	2.47 ± 0.07
AP25	artificial seawater	0
STS-01	distilled water	28.7 ± 0.9
STS-02	distilled water	3.16 ± 0.10
RU	distilled water	0
AM	distilled water	0

Notes: ANA: 100% anatase crystalline form (Wako); AP25: 20% rutile 80% anatase crystalline form (Japan Aerosil); STS-01: photocatalytic sol (Ishihara Sangyo); STS-02: photocatalytic sol (Ishihara Sangyo); RU: 100% rutile crystalline form (Wako); AM: amorphous (Wako). Numbers represent mean value \pm SD.

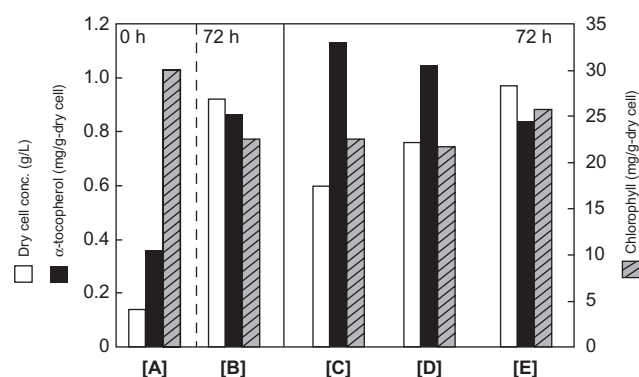


Figure 5 Effect of various photocatalysts on the physiological activities of *E. gracilis* at 72 h of cultivation. **A)** Inoculum at 0 h, **B)** no addition, **C)** 0.01% (w/v) ANA, **D)** 0.01% (w/v) AP25, **E)** 0.01% (w/v) RU. The data are expressed as mean \pm SD ($n = 3$).

without sampling procedure. It was also possible to use changes in the absorbance to estimate the end of the reaction. In the case of sample containing heterogeneous fine particles in terms of size and specific gravity, it is difficult to evaluate the photocatalytic activity accurately (the results lack reproducibly). The method described here solves the problems, and it is excellent for measuring the photocatalytic activities of various fine particles accurately, reproducibly, and conveniently.

The changes in the absorbance varied depending on the type of photocatalysts and the suspending liquid. AP25 lost its photocatalytic activity, while that of ANA decreased significantly in artificial seawater (Figure 3). With this apparatus, it was possible to highlight the difference in the effects of seawater on ANA and AP25. It is clear that if ANA and AP25 are released into the sea, the environmental impact of ANA is more than that of AP25. It is known that the presence of inorganic anions such as chloride in water reduces the photocatalytic activity. This can be explained by the competition with dissolved organic carbons for adsorption or reaction.²⁰ It was reported that metal ions, (sodium and magnesium) resulted in enhanced photocatalytic activity at concentrations similar to those in seawater.²¹ The rate of photooxidative dye decolorization in seawater was slower than that in the NaCl solution even although sodium cation and chlorine anion are the major ions in seawater.²¹ The reasons for the differences observed between ANA and AP25 in artificial seawater is currently being investigated. By using the values of k (reaction rate constant), it was possible to quantitatively evaluate and compare the photocatalytic activities of various fine particles under various conditions (Table 1). In the case of ANA, the photocatalytic activity in artificial seawater decreased to one

third of that in distilled water. The photocatalytic activities of TiO_2 fine particles differed greatly depending on such factors as the crystalline form, composition, particle size, and surface area.^{8,13}

When ANA and AP25 were added to the medium under illumination by black light, the growth of *E. gracilis* was suppressed but their alpha-tocopherol contents increased (Figure 5). The effects of ANA on *E. gracilis* were more pronounced than those of AP25. This may be due to the fact that the k value of ANA in the *E. gracilis* cell culture medium was higher than that of AP25. The modified Hutner medium contains many kinds of salts and this resulted in decrease in the k values of ANA and AP25. This is consistent with the results obtained when they were suspended in artificial seawater (Figure 3). It is well known that various kinds of ROS are produced on TiO_2 surface exposed to light rays of wavelengths below 410 nm. Among these, hydroxyl radical is dominant and is known to be responsible for the oxidative stress on living cells.¹⁰ The ROS produced by photocatalytic reaction enhanced the alpha-tocopherol production by *E. gracilis*. This prevents oxidative decomposition of chlorophyll, lipid membranes, and proteins by ROS.²² through scavenging ROS released during oxidative stress.²³ It was reported that the antioxidant alpha-tocopherol synthesis was activated to prevent damage by ROS.²⁴ It has also been reported that antioxidant carotenoid biosynthesis is regulated by ROS in some microalgae.²⁵ The increase in alpha-tocopherol content of *E. gracilis* exposed to TiO_2 suggests that the TiO_2 fine particle promoted production of alpha-tocopherol and/or selected *E. gracilis* cells with high content of antioxidant. Thus, TiO_2 may be used to enhance production of useful antioxidants.

Quantitative evaluation of photocatalytic activities of fine particles using the apparatus developed in this study is very important, considering the ecotoxicity and environmental impacts of these particles. By knowing the potential risks of these photocatalytic particles, it is possible to make a risk-benefit analysis.

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Disclosure

The authors report no conflicts of interest in relation to this paper.

References

- Nagasawa H, Ichimura A, Isoda S. Carboxylate-passivated silver nanoparticles and their application to sintered interconnection: a replacement for high temperature lead-rich solders. *J Electron Mater*. 2010;39:1233–1240.
- Liu SQ, Tang ZY. Nanoparticle assemblies for biological and chemical sensing. *J Mater Chem*. 2010;20:24–35.
- Narayanan R. Recent advances in noble metal nanocatalysts for Suzuki and Heck cross-coupling reactions. *Molecules*. 2010;15:2124–2138.
- Ogura H, Maruyama M, Matsubayashi R, et al. Nanoscale particle therapies for wounds and ulcers. *Nanomedicine*. 2010;5:641–656.
- Millis A, Lepre A, Elliott N, Bhopal S, Parkin I, O'Neill S. Characterisation of the photocatalyst Pilkington Active™: a reference film photocatalyst? *J Photochem Photobiol A Chem*. 2004;160:213–224.
- Esterkin CR, Negro AC, Alfano OM, Cassano AE. Air pollution remediation in a fixed bed photocatalytic reactor coated with TiO₂. *AIChE J*. 2005;51:2298–2310.
- Choi H, Stathatos E, Dionysiou DD. Sol-gel preparation of mesoporous photocatalytic TiO₂ films and TiO₂/Al₂O₃ composite membranes for environmental applications. *Appl Catal B Environ*. 2006;63:60–67.
- Adams LK, Lyon DY, Alvarez PJJ. Comparative eco-toxicity of nanoscale TiO₂, SiO₂, and ZnO water suspension. *Water Res*. 2006;40:3527–3532.
- Kaida T, Kobayashi K, Adachi M, Suzuki F. Optical characteristics of titanium oxide interference film and the film laminated with oxides and their applications for cosmetics. *J Cosmet Sci*. 2004;55:219–220.
- Cai R, Kubota Y, Shuin T, et al. Induction of cytotoxicity by photoexcited TiO₂ particles. *Cancer Res*. 1992;52:2345–2348.
- Otaki M, Hirata T, Ohgaki S. Aqueous microorganisms inactivation by photocatalytic reaction. *Water Sci Technol*. 2000;43:115–118.
- Sunada K, Watanabe T, Hashimoto K. Studies on photokilling of bacteria TiO₂ thin film. *J Photochem Photobiol A Chem*. 2003;156:227–233.
- Hund-Rinke K, Simon M. Ecotoxic effect of photocatalytic active nanoparticles (TiO₂) on algae and daphnids. *Environ Sci Pollut Res*. 2006;13:225–232.
- Oberdorster G, Oberdorster E, Oberdorster J. Nanotoxicology: an emerging discipline evolving from studies of ultrafine particles. *Environ Health Perspect*. 2005;113:823–839.
- Xu NP, Shi ZF, Fan YQ, Dong JH, Shi J, Hu MZC. Effects of particle size of TiO₂ on photocatalytic degradation of methylene blue in aqueous suspensions. *Ind Eng Chem Res*. 1999;38:373–379.
- Ogbonna JC, Tanaka H. Cyclic autotrophic/heterotrophic cultivation of photosynthetic cells: a method of achieving continuous cell growth under light/dark cycles. *Bioresour Technol*. 1998;65:65–72.
- Shigeoka S, Onishi T, Nakano Y, Kitaoka S. The contents and subcellular distribution of tocopherols in *Euglena gracilis*. *Agric Biol Chem*. 1986;50:1063–1065.
- Porra RJ, Thompson WA, Kriedemann PE. Determination of accurate extinction coefficients and simultaneous equations for assaying chlorophylls a and b extracted with four different solvents: verification of the concentration of chlorophyll standards by atomic absorption spectroscopy. *Biochim Biophys Acta*. 1989;975:384–394.
- Otsuka-Yao-Matsuo S, Omata T, Yoshimura M. Photocatalytic behavior of cerium titanates, CeTiO₄ and CeTi₂O₆ and their composite powders with SrTiO₃. *J Alloys Compd*. 2004;376:262–267.
- Chen HY, Zahraa O, Bouchy M. Inhibition of the adsorption and photocatalytic degradation of an organic contaminant in an aqueous suspension of TiO₂ by inorganic ions. *J Photochem Photobiol A Chem*. 1997;108:37–44.
- Makita M, Harata A. Photocatalytic decolorization of rhodamine B dye as a model of dissolved organic compounds: Influence of dissolved inorganic chloride salts in seawater of the sea of Japan. *Chem Eng Process*. 2008;47:859–863.
- Tani Y, Tsumura H. Screening for tocopherol-producing microorganisms and alpha-tocopherol production by *Euglena gracilis* Z. *Agric Biol Chem*. 1989;53:305–312.
- Munne-Bosch S, Alegre L. The function of tocopherols and tocotrienols in plants. *CRC Crit Rev Plant Sci*. 2002;21:31–57.
- Fujita T, Ogbonna JC, Tanaka H, Aoyagi H. Effects of reactive oxygen species on alpha-tocopherol production in mitochondria and chloroplasts of *Euglena gracilis*. *J Appl Phycol*. 2009;21:185–191.
- Kobayashi M, Kakizono T, Nagai S. Enhanced carotenoid biosynthesis by oxidative stress in acetate induced cyst cells of a green unicellular alga, *Haematococcus pluvialis*. *Appl Microbiol Biotech*. 1993;59:867–873.

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