

The Role of CCL20-CCR6 Axis in Ovarian Cancer Metastasis

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Background: Chemokine networks play a key and complex role in tumor progression. CCL20 and its unique receptor CCR6 have been reported to mediate malignant biological activities in various cancers, but their role in ovarian cancer metastasis remains unclear.

Purpose: Our study aims to explore the effect of CCL20-CCR6 axis on ovarian cancer metastasis and its potential mechanism.

Methods: The transwell assay was used to detect the cell migration and invasion after CCL20 treatment. The CCK-8 assay was used to detect the cell viability after CCL20 treatment and CCR6 depletion. The mRNA and protein expression were assayed through qRT-PCR and Western blotting. The siRNAs and CRISPR-Cas9 system were adopted to suppress CCR6 expression. Intraperitoneal xenograft mouse model was constructed to test the pro-metastasis effect of CCL20-CCR6 axis in vivo. The differentially expressed genes induced by CCL20 were identified through RNA-sequencing, and immunohistochemistry staining was used to detect their protein expression in tumor tissues.

Results: Our results revealed that CCL20 treatment selectively promoted the migration and invasion of CCR6^{high} ovarian cancer cells, but had no effect on CCR6^{low} cells. Blockade of CCR6 expression effectively reversed the cell migration and invasion induced by CCL20 stimulation. Animal experiment proved that CCL20-CCR6 axis mediated ovarian cancer metastasis in vivo. The differentially expressed genes after CCL20 stimulation were associated with metastasis, and CCL20 induced an increased expression of CDH2 and VCAN and decreased CDH1 expression in cancer cells. Moreover, CCL20 stimulated the expression of N-cadherin and versican in tumor tissues and inhibited the expression of E-cadherin, while CCR6 knockout successfully blocked the expression changes.

Conclusion: Our findings revealed that CCL20-CCR6 axis promotes ovarian cancer metastasis both in vivo and in vitro, probably through increasing cancer cell adhesion and epithelial-mesenchymal transition. Blockade of CCL20-CCR6 axis might become a novel anti-tumor therapeutic target for ovarian cancer.

Keywords: CCL20, CCR6, ovarian cancer, metastasis, transcriptional regulation

Introduction

In 2020, the mortality rate of ovarian cancer ranks fifth among American women.¹ Due to the lack of effective screening and early clinical diagnosis methods, more than 70% of patients are diagnosed as advanced stage and often accompanied by extensive cancer metastasis.^{2,3} The current standard treatment for ovarian cancer is cytoreductive surgery followed by platinum and taxane chemotherapy.⁴ Although the initial response rate of chemotherapy is up to 80%, more than 70% of patients will still experience cancer recurrence.⁵ In the past decade, targeted drugs have been widely tested in clinical trials for the treatment of recurrent ovarian cancer,

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and some targeted drugs have been approved by the United States Food and Drug Administration (FDA) and/or European Medicine Agency for clinical use.⁶ Such targeted drugs include angiogenesis inhibitors (bevacizumab), poly (ADP-ribose) polymerase (PARP) inhibitors (olaparib, rucaparib and niraparib), neurotrophic tropomyosin receptor kinase (NTRK) inhibitors (entrectinib and larotrectinib) and immune checkpoint inhibitors (pembrolizumab). Several targeted drugs have improved the prognosis of patients to a certain extent and showed less side effects than chemotherapy,^{7,8} but there are some limitations in application. For example, olaparib is only recommended for the treatment of ovarian cancer patients with a BRCA1/2 mutation and entrectinib only for patients with NTRK fusion-positive solid tumors,^{9–11} and the efficacy of pembrolizumab as a single-agent therapy is very limited in treatment of ovarian cancer.¹² Therefore, it is urgent to further investigate the mechanisms underlying the metastasis and aggressiveness of ovarian cancer and develop novel antitumor strategies for ovarian cancer.

Among various factors that affect tumor progression, one of the important factors is the chemokine superfamily. Members of this family include a large number of low-molecular-weight (8–14 kDa) chemotactic cytokines, which exert various biological functions by activating seven transmembrane domain G protein-coupled receptors on target cells.^{13–15} To date, approximately 50 chemokine ligands and 20 receptors have been identified. In malignant tumors, chemokine receptors are not only expressed in immune cells in the tumor microenvironment, but also in tumor cells and endothelial cells.^{16,17} Therefore, in addition to recruit anti- or pro-tumor immune cells to the tumor site, chemokines can also directly act on tumor cells and endothelial cells to regulate tumor cell growth, angiogenesis, invasiveness and metastasis.^{18–20} Different cytokines can play different roles in different types of cancers. For instance, CCL5 derived from ovarian cancer stem cells could recruit Treg cells to the tumor site and thereby facilitating tumor metastasis,^{20,21} whereas CCL14 could promote hepatocellular carcinoma (HCC) cell apoptosis by inhibiting cell cycle progression via Wnt/ β -catenin signaling pathway, making it a favorable prognostic indicator in HCC patients.^{22,23} Despite increasing studies exploring the functions of chemokines in tumors, knowledge about the complex roles of chemokines in different types of tumors remains limited, and further studies should be focused on deciphering the roles and mechanisms of specific chemokines in specific types of tumors.

Recent studies have shown that the CCL20-CCR6 axis plays a key role in tumor progression. CCL20 is a 9 kDa chemokine and the only known ligand for CCR6 receptor.²⁴ Non-tumor cells in the tumor microenvironment, such as stromal cells in giant cell tumor of bone, astrocytes in glioblastoma, and tumor-associated macrophages in melanoma are important sources of CCL20, which can recruit suppressive immune cells to the tumor site to maintain an immunosuppressive microenvironment, or act on endothelial cells to promote angiogenesis.^{25–30} In addition, CCL20 can also directly mediate the migration and/or invasion process of various cancer cells, including lung adenocarcinoma, cholangiocarcinoma, colon cancer and HCC,^{31–34} but its role in ovarian cancer metastasis remains unclear. Our previous studies have demonstrated that classically activated macrophages after cisplatin stimulation promoted the migration of ovarian cancer cells through the CCL20-CCR6 axis.³⁵ In this study, we further verified the role of the CCL20-CCR6 axis in the metastasis of ovarian cancer and explored its related molecular mechanisms.

Materials and Methods

Cell Culture

The human ovarian epithelial cell line IOSE and human ovarian cancer cell lines A2780, SKOV3, HEY and ES2 were purchased from the Cell Bank of the Chinese Academy of Sciences (Shanghai, China). All cells were cultured in DMEM/high glucose medium (HyClone, South Logan, UT, USA) containing 10% fetal bovine serum (FBS) (Gibco, Gaithersburg, MD, USA) and 1% penicillin-streptomycin (Sigma Aldrich, St Louis, MO, USA) in a humidified atmosphere with 5% CO₂ at 37°C.

Transwell Migration and Invasion Assays

For migration assay, ovarian cancer cells were harvested and resuspended in FBS-free medium with recombinant human CCL20 protein (rhCCL20, 20 ng/mL) (Peptrotech, Cranbury, NJ, USA). Cell suspensions (A2780: 1.5×10^5 cells/200 μ L, SKOV3, HEY and ES2: 1.5×10^4 cells/200 μ L) were then placed into the upper chamber of transwell plates (8 μ m; Corning, Corning, NY, USA). For invasion assays, 40 μ L matrigel matrix (pre-diluted 1:5 with serum-free medium) (BD Biosciences, Franklin Lakes, NJ, USA) was placed into the upper chamber of transwell plates, and cell suspensions (A2780: 3×10^5 cells/200 μ L, SKOV3, HEY and ES2: 3×10^4 cells/200 μ L) were added 2 hours

later. Subsequently, 700 μ L medium containing 20 ng/mL rhCCL20 and 10% FBS was added into the lower chamber. After incubation for 24 hours (migration) or 72 hours (invasion) at 37°C, cells in the upper chamber were carefully removed. Migrated or invaded cells were fixed with 4% paraformaldehyde, stained with 0.1% crystal violet (Sigma Aldrich), and quantified using an IX71 inverted microscope (Olympus Corp, Shinjuku-ku, Japan) at $\times 200$ magnification.

Cell Counting Kit-8 (CCK-8) Cell Viability Assay

Ovarian cancer cells (4000/well) were cultured in 96-well plates with increasing concentrations of rhCCL20. At indicated time-points, 10 μ L CCK-8 reagent (Dojindo, Kumamoto, Japan) was added for 2-hour incubation, and then the absorbance was measured using a microplate reader (Thermo Scientific, Waltham, MA, USA) at 450 nm. Concentration-response curves were created using GraphPad Prism 6.0 (La Jolla, CA, USA).

Quantitative Real-Time PCR (qRT-PCR)

Total RNA was extracted using TRIZOL reagent (Invitrogen, Carlsbad, CA, USA) and reversely transcribed into cDNA using a PrimeScript RT Reagent Kit (Takara, Otsu, Japan). qRT-PCR was performed using QuantStudio™ Test Development Software (Thermo Scientific) with SYBR Green qPCR Master Mix (EZBioscience, Roseville, MN, USA). The data were analyzed using the $2^{-\Delta\Delta CT}$ method. Glyceraldehyde 3-phosphate dehydrogenase was used for data normalization. The sequences of primers were as follows: CCR6, forward: GCT TGA GCT ACT CTT TGG TTT C, reverse: CAA GCA CCA CAG CTA TGA TTA C; VCAN, forward: ACT GAA ACT TCC TAC GTA TGC A, reverse: CTC ACA AAG TGC ACC AAC ATA A; CDH1, forward: AGT CAC TGA CAC CAA CGA TAA T, reverse: ATC GTT GTT CAC TGG ATT TGT G; CDH2, forward: CGA TAA GGA TCA ACC CCA TAC A, reverse: TTC AAA GTC GAT TGG TTT GAC C. All primers were purchased from Sangon Biotech (Shanghai, China).

Western Blotting

Total protein was extracted using RIPA lysis buffer (Beyotime, Shanghai, China) containing 1% phenylmethane-sulfonyl fluoride (Beyotime). Equal quantities of protein were electrophoresed in polyacrylamide gel (Beyotime), transferred

onto PVDF membranes (Millipore, Billerica, MA, USA), and blocked in 5% bovine serum albumin for 1 hour. Antibodies used in Western blotting were CCR6 (Abcam, Cambridge, UK) and β -actin (Sigma-Aldrich). Species-specific secondary anti-mouse and anti-rabbit antibodies (LI-COR, Lincoln, NE, USA) were used to visualize the blots under the Odyssey imaging system (LI-COR).

Cell Transfection

CCR6-specific siRNAs (siCCR6-1: 5'-GCUCCGAUCCA GAACACUATT-3', siCCR-2: 5'-GGGCAGAAGUUCA GAAACUTT-3') and negative control siRNA (siRNA-NC: 5'-UUCUCCGAACGUGUCACGUTT-3') were constructed by Genephama Biotechnologies (Shanghai, China). A2780 and SKOV3 cells were transiently transfected with siRNAs using Lipofectamine 2000 (Invitrogen) according to the manufacturer's protocols and were collected for further transwell migration and invasion experiments.

CRISPR-Cas9 System-Mediated Knockout of CCR6

The construction of transfer plasmids (lentiCRISPRv2) and packaging plasmids (delta8.9 and VSVG) was entrusted to Obio technology corporation (Shanghai, China). Briefly, a pair of annealed oligonucleotides was cloned into lentiCRISPRv2 plasmids to obtain lentiviral single-guide RNA (sgRNA) plasmids. The forward sequence of sgCCR6 was: ACC GTT ACT GTG CTC CTT GCA GG, and the reverse sequence was: AAA CCC TGC AAG GAG CAC AGT AA. The lentiviral sgRNA plasmids and packaging plasmids were co-transfected into 293T cells using lipofectamine 3000 (Invitrogen). After purification and ultracentrifugal concentration, the supernatant was collected for titer detection and ovarian cancer cell infection. To generate CCR6 knock-out cells, A2780 cells were cultured in supernatant containing lentiviral particles together with 5 μ g/mL polybrene (Sigma-Aldrich). 72 hours after infection, 5 μ g/mL puromycin was added for selection, and the surviving cells were collected for future monoclonal cell selection.

In vivo Experiment

All animal experiments followed the guidelines of the Institutional Animal Care and Use Committee of Ren Ji Hospital, and all experimental procedures were approved by the Institutional Animal Care and Use Committee of Ren Ji Hospital. For in vivo metastasis mouse model, 2×10^6 of wild type (WT) A2780 cells or CCR6 knock-out (KO)

A2780 cells were injected into the abdominal cavity of 5-week-old female nude mice. One week after intraperitoneal inoculation of tumor cells, each group was further randomly divided into two groups and treated with PBS or 1.5 mg/kg rhCCL20 (dissolved in PBS) intraperitoneally, and twice a week afterwards. Thus experiment mice were divided into 4 groups (six mice per group) based on the types of inoculated cells and treatments: WT-PBS group (WT-A2780 cells inoculation and PBS treatment), WT-CCL20 group (WT-A2780 cells inoculation and rhCCL20 treatment), KO-PBS group (CCR6-KO-A2780 cells inoculation and PBS treatment) and KO-CCL20 group (CCR6-KO-A2780 cells inoculation and rhCCL20 treatment). Mice were euthanized 1 month after inoculation and the metastatic tumors were surgically excised and weighted.

Immunohistochemical (IHC) Staining and Hematoxylin and Eosin (H&E) Staining

Tumor tissues were fixed in 10% formalin for 24h, embedded in paraffin, and sectioned into 5 μ m thick sections. After deparaffinization and rehydration, the sections were stained with hematoxylin and eosin. For immunohistochemistry, after neutralizing the endogenous peroxidase with 1% H₂O₂ and a specific protein block, the slices were incubated with antibodies against N-cadherin (Servicebio, Wuhan, China), E-cadherin (Servicebio) or versican (Boster, Wuhan, China) at 4°C overnight followed by incubating with HRP-conjugated anti-rabbit or anti-mouse antibody for 45 minutes. The sections were visualized with diaminobenzidine and counterstained with hematoxylin. Images were scanned at 40 \times magnification using a P-MIDI (3D HISTECH, Budapest, Hungary).

RNA-Sequencing

The RNA-sequencing was entrusted to OE biotechnology corporation (Shanghai, China). Briefly, total RNA was extracted using a mirVana miRNA Isolation Kit (Invitrogen), and the integrity of RNA was assessed using the Agilent Bio analyzer 2100 (Agilent Technologies, Santa Clara, CA, USA). The RNA integrity number of all samples was ≥ 7 , suggesting good RNA quality. The cDNA libraries were constructed using the TruSeq Stranded mRNA LT Sample Prep Kit (Illumina, San Diego, CA, USA) in accordance with the manufacturer's instructions, and then were sequenced on an Illumina Hiseq X Ten platform. *P* value < 0.05 and fold change > 1.5 or < 0.67 was set as the threshold for

significantly differential expression. Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway enrichment analysis of differentially expressed genes was performed using an R program.

Statistical Analysis

All the experiments were performed in triplicate. Data were presented as mean \pm standard deviation (SD) and analyzed by SPSS 21.0 software. The differences between two groups were analyzed by two-tailed Student's *t*-test. One-way analysis of variance (ANOVA) was used for comparisons of more than two groups. *P* < 0.05 was considered statistically significant.

Results

Effects of CCL20 on Migration and Invasion of Ovarian Cancer Cell Lines

Our previous study revealed that cisplatin-stimulated classically activated macrophages promote ovarian cancer cell migration through CCL20-CCR6 axis.³⁵ To further study the role of CCL20-CCR6 axis in ovarian cancer metastasis and its underlying mechanisms, we first examined the effects of rhCCL20 protein on the migration and invasion capacity as well as proliferation of ovarian cancer cell lines. The results showed that rhCCL20 protein significantly promoted the migration and invasion of A2780 and SKOV3 cells, but had no such pro-effect on ES2 and HEY cells (Figure 1A and B). In addition, rhCCL20 protein had no significant effect on the proliferation of A2780, SKOV3, ES2 and HEY cells (Figure 1C).

CCR6 Expression in Ovarian Cancer Cell Lines

CCR6 is currently the only known receptor for CCL20 ligand. Our previous studies have shown that ovarian cancer patients with higher CCR6 expression had a worse clinical prognosis, and their gene expression profiles were highly enriched in gene sets associated with metastasis.³⁵ Considering that rhCCL20 protein had inconsistent effects on the promotion of migration and invasion in different ovarian cancer cells, we tested the expression of its receptor CCR6 in ovarian cancer cell lines. The result (Figure 2A) showed that compared to human ovarian epithelial cell line IOSE, CCR6 mRNA was generally highly expressed in ovarian cancer cell lines (SKOV3, A2780, ES2 and HEY cells), and mRNA expression of CCR6 in SKOV3 and

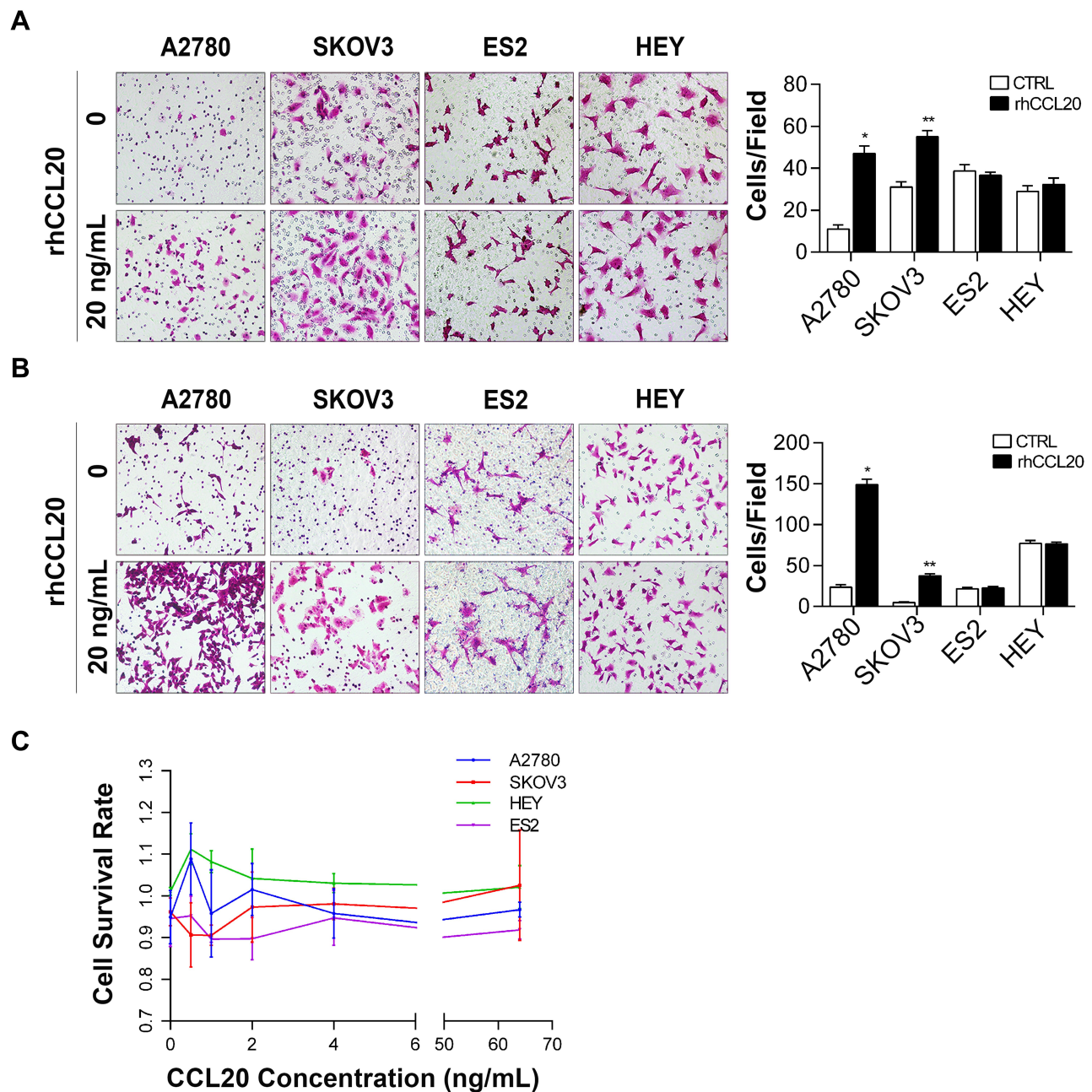


Figure 1 CCL20 treatment selectively promotes ovarian cancer cell migration and invasion.

Notes: (A) Migration of ovarian cancer cells treated with rhCCL20 (20 ng/mL). (B) Invasion of ovarian cancer cells treated with rhCCL20 (20 ng/mL). (C) Proliferation of ovarian cancer cells treated with increasing concentrations of rhCCL20 (0.5, 1, 2, 4, 8, 16, 32, 64 ng/mL). *Compared to A2780 CTRL group, $P < 0.05$; **Compared to SKOV3 CTRL group, $P < 0.05$.

Abbreviations: rhCCL20, recombinant human CCL20 protein; CTRL, control.

A2780 cells was significantly higher than that in ES2 and HEY cells. We further verified the protein expression of CCR6 in IOSE and ovarian cancer cells by Western blotting, and the result was consistent with CCR6 expression at the mRNA level (Figure 2B).

Taken all together, we can conclude that rhCCL20 protein selectively promotes the migration and invasion of CCR6^{high} ovarian cancer cells (A2780 and SKOV3 cells), but has no such effect on CCR6^{low} ovarian cancer cells (ES2 and HEY cells).

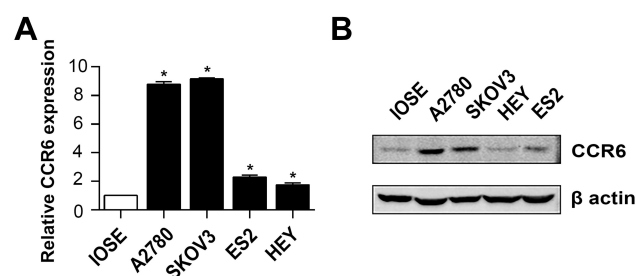


Figure 2 CCR6 expression in ovarian cancer cell lines.

Notes: (A) mRNA expression of CCR6 in human ovarian epithelial cell line IOSE and ovarian cancer cell lines. (B) Protein expression of CCR6 in IOSE and ovarian cancer cells. *Compared to IOSE, $P < 0.05$

Blockade of CCR6 Reverses CCL20-Mediated Migration and Invasion of Ovarian Cancer Cells

To further confirm the role of CCR6 activation in ovarian cancer cell migration and invasion, two CCR6-specific siRNAs (siCCR6-1 and siCCR6-2) were transfected into A2780 and SKOV3 cells to inhibit CCR6 expression. Our previous study proved that these two siRNAs were efficient to reduce CCR6 expression by at least 50% and did not affect the viability of ovarian cancer cells.³⁵ Here the result showed that rhCCL20 protein significantly promoted migration and invasion of ovarian cancer cells transfected with negative siRNA (siRNA-NC), whereas knockdown of CCR6 by siCCR6 significantly decreased ovarian cancer cell migration (Figure 3A) and invasion (Figure 3B) induced by rhCCL20 protein, indicating that CCR6 activation by CCL20 mediated the migration and invasion of ovarian cancer cells.

In addition, we used the CRISPR-Cas9 system to construct sgCCR6 lentivirus and transfected it into A2780 cells to construct CCR6 knock-out (CCR6-KO) A2780 cells. After lentivirus transfection and puromycin selection, we collected 7 monoclonal cell lines. Western blotting was used to verify the knockout effect. The result showed that the expression of CCR6 in No.6 monoclonal cells was significantly suppressed (Figure 4A); therefore, we chose No.6 monoclonal cells as CCR6-KO cells for subsequent experiments. Consistent with the above results, knockout of CCR6 by sgCCR6 lentivirus did not affect viability of A2780 cells (Figure 4B), but did inhibit ovarian cancer cell migration and invasion promoted by rhCCL20 protein (Figure 4C), further confirming the conclusion that CCL20-CCR6 axis mediates the migration and invasion of ovarian cancer cells.

CCL20-CCR6 Axis Mediates Ovarian Cancer Metastasis in vivo

To further investigate the pro-metastasis effect of CCL20-CCR6 axis on ovarian cancer in vivo, we randomly divided nude mice into 4 groups based on the inoculated cells and treatments: WT-PBS group (WT-A2780 cells inoculation + PBS treatment), WT-CCL20 group (WT-A2780 cells inoculation + rhCCL20 treatment), KO-PBS group (CCR6-KO-A2780 cells inoculation + PBS treatment) and KO-CCL20 group (CCR6-KO-A2780 cells inoculation + rhCCL20 treatment). Mice were euthanized 1 month after inoculation and the tumors were surgically excised ($n = 6/\text{group}$). There were no significant differences in body weight among the four groups throughout the experiment (Figure 5A). Strikingly, the mice in WT-CCL20 group exhibited obvious tumor metastasis compared with that in WT-PBS group, whereas no significant differences were observed between KO-PBS and KO-CCL20 groups (Figure 5B). Next, nude mice were dissected out of abdominal metastatic tumors (Figure 5C), and the number (Figure 5D) and weight (Figure 5E) of the tumors were counted and compared. Results showed that rhCCL20 protein could significantly promote the number and weight of WT-A2780 tumors in the abdominal cavity of nude mice, but it had no such effect on CCR6-KO-A2780 tumors. Collectively, these data suggest that CCL20-CCR6 axis facilitates ovarian cancer metastasis in vivo.

Differentially Expressed Genes Elicited by CCL20 in Ovarian Cancer Cells

To further study the mechanism of CCL20-mediated migration and invasion of ovarian cancer cells, we used RNA sequencing to examine the effect of rhCCL20 treatment on gene expression profiles in ovarian cancer cells. The data have been deposited in NCBI's gene expression omnibus (GEO) and is accessible through GEO series accession number GSE161799 (<https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE161799>). The differentially expressed genes are shown in Figure 6A and Table S1. KEGG analysis of these differential genes revealed a significant enrichment in cell adhesion molecules (CAMs) (Figure 6B). VCAN and CDH2, whose protein products play a crucial role in the regulation of cell motility, cell-cell adhesion and epithelial-mesenchymal transition (EMT), were greatly up-regulated after rhCCL20 treatment (Figure 6C and D). In addition, rhCCL20 exhibited a profound reduction in the transcription of CDH1,

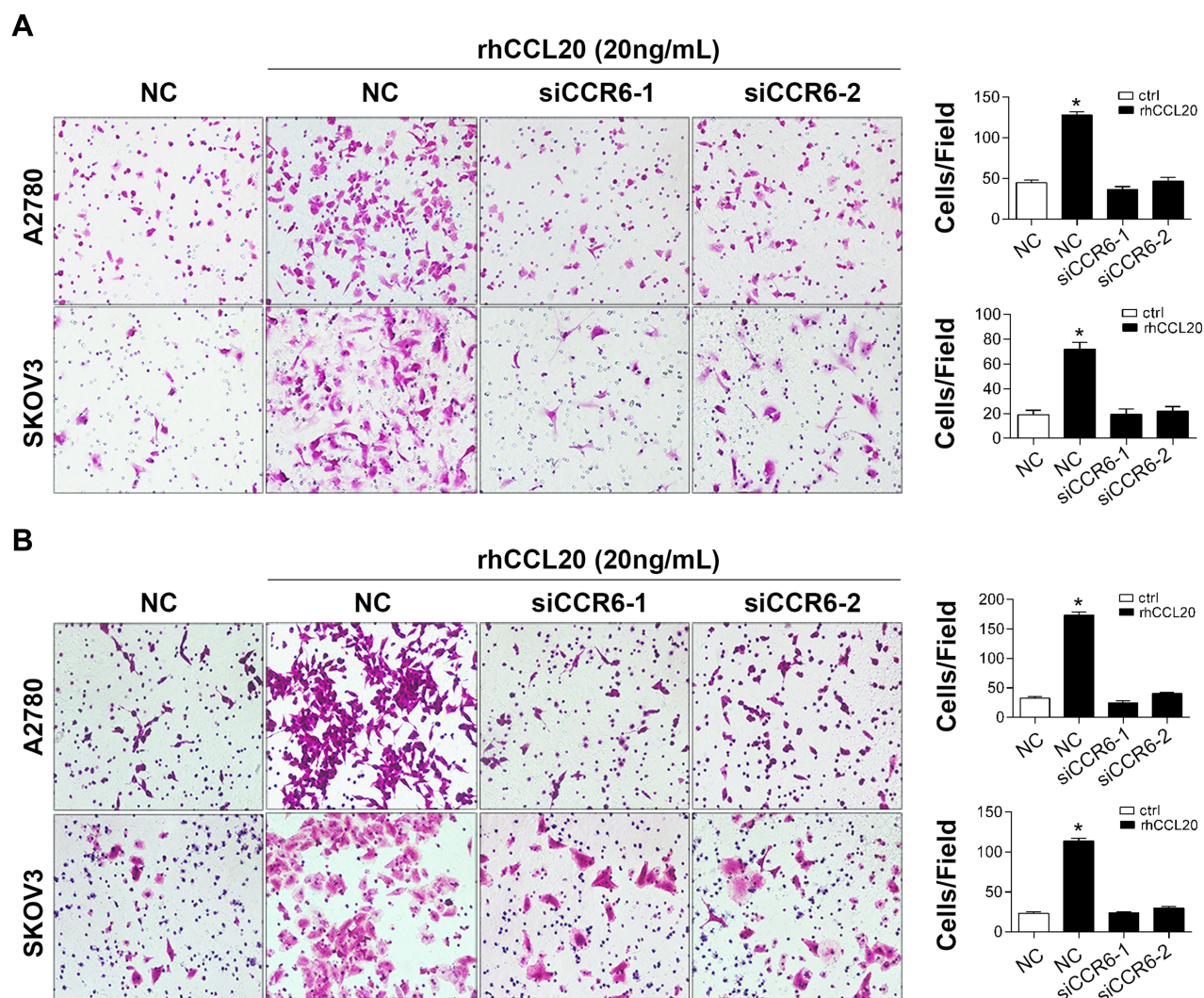


Figure 3 Knockdown of CCR6 inhibits CCL20-mediated migration and invasion of ovarian cancer cells.

Notes: (A) Migration of ovarian cancer cells transfected with siRNAs and treated with rhCCL20. (B) Invasion of ovarian cancer cells transfected with siRNAs and treated with rhCCL20. *Compared to NC group, $P < 0.05$

Abbreviations: NC, negative control; rhCCL20, recombinant human CCL20 protein.

whose loss is considered the key process of EMT (Figure 6C and D). Moreover, IHC analysis of tumor tissues from in vivo experiment revealed that compared with WT-PBS group, N-cadherin and versican, the products of CDH2 and VCAN, were obviously increased in the tumors of WT-CCL20 group, and the product of CDH1 E-cadherin decreased dramatically. However, no significant differences were observed in the expression of N-cadherin, E-cadherin and versican between KO-PBS group and KO-CCL20 group (Figure 6E). Taken together, these data indicate that CCL20 promotes ovarian cancer cell migration and invasion through promoting the motility and epithelial-mesenchymal transition in ovarian cancer cells.

Discussion

Emerging evidence suggests that chemokines and their receptors are associated with the pathogenesis, progression and metastasis of various types of tumors, including ovarian cancer.³⁶ Up to now, several achievements have been made on cancer targeted therapy targeting the chemokine system. For example, mogamulizumab, an anti-CCR4 antibody, is used to treat patients with relapsed and refractory adult T cell leukemia-lymphoma.³⁷ Plerixafor, a CXCR4 antagonist, has been approved for the mobilization of hematopoietic stem cells for transplantation in non-Hodgkin's lymphoma or multiple myeloma.³⁸ Therefore, in-depth study about the roles and mechanisms of

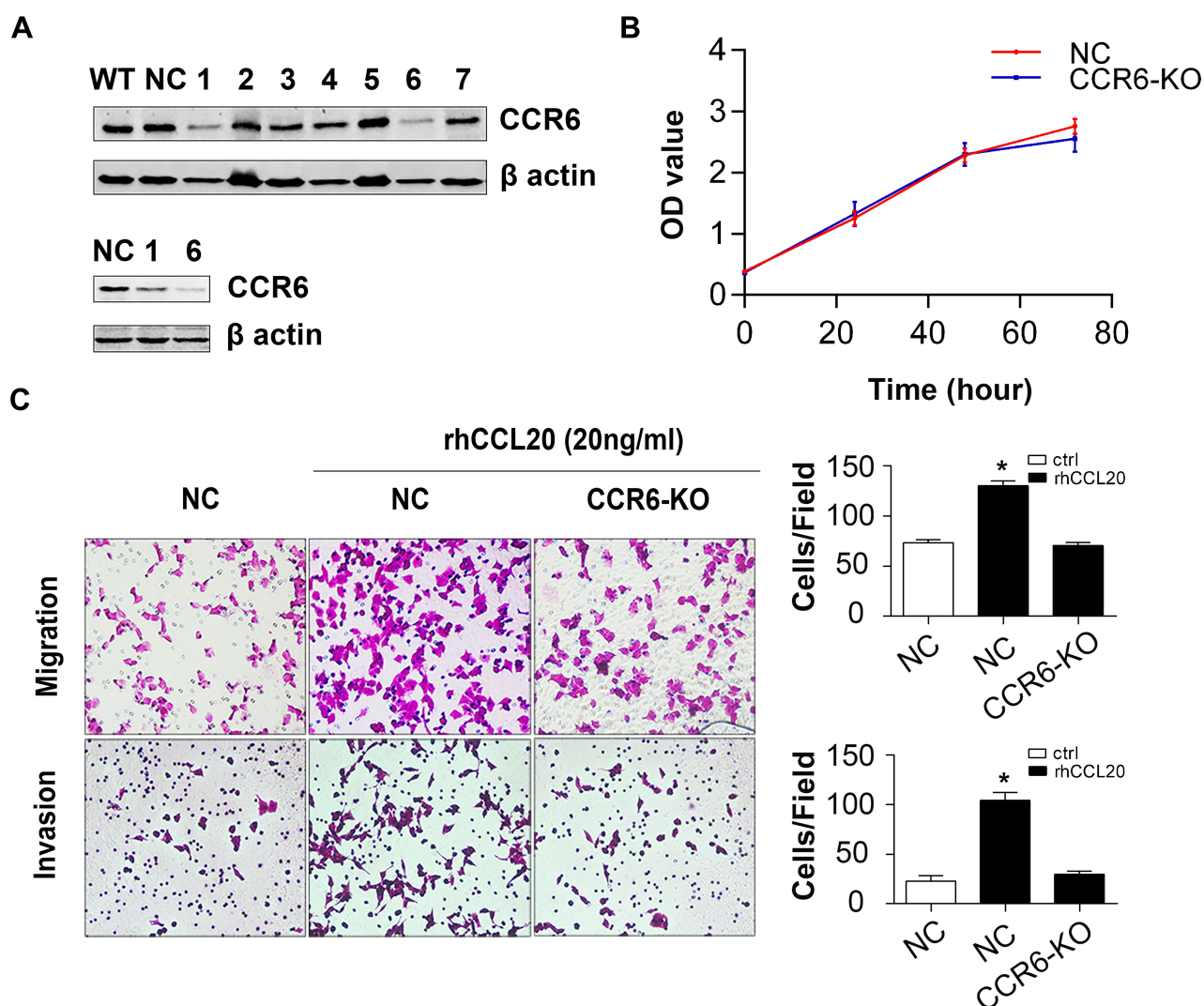


Figure 4 Knockout of CCR6 inhibits CCL20-mediated migration and invasion of ovarian cancer cells.

Notes: (A) Protein expression of CCR6 in A2780 monoclonal cells transfected with sgCCR6 lentivirus. (B) The effect of CCR6 knockout on proliferation of A2780 cells. (C) The effect of CCR6 knockout on CCL20-induced cell migration and invasion. *Compared to NC group, $P < 0.05$

Abbreviations: KO, knock-out; NC, negative control; rhCCL20, recombinant human CCL20 protein.

chemokine networks can help develop novel targeted drugs for cancer treatment. In this study, we studied the effect of CCL20-CCR6 axis on metastasis of ovarian cancer and explored its potential mechanisms.

Our previous study revealed that classically activated macrophages after cisplatin stimulation promoted ovarian cancer cell migration, and CCL20 played a vital role in the process.³⁵ In this study, we firstly treated ovarian cancer cells with rhCCL20 protein and found that rhCCL20 selectively promoted the migration and invasion of A2780 and SKOV3 cells, but did not affect ES2 and HEY cells. Since CCR6 is currently the only receptor for CCL20, and our previous study revealed a potential correlation between CCR6 expression and ovarian cancer metastasis, therefore

we tested CCR6 expression in ovarian cancer cell lines. The result showed that CCR6 was generally highly expressed in ovarian cancer cell lines compared to normal ovarian epithelial cells IOSE, and its expression in A2780 and SKOV3 cells was significantly higher than that in ES2 and HEY cells. Furthermore, blockade of CCR6 by CCR6-specific siRNAs and sgCCR6 lentivirus effectively reversed cell migration and invasion induced by CCL20 stimulation in A2780 and SKOV3 cells. Moreover, rhCCL20 strongly promoted the tumorigenesis of WT-A2780 metastatic tumors in the abdominal cavity of nude mice, but had no effect on CCR6-KO-A2780 metastatic tumors. Taken all above, we can conclude that CCL20-CCR6 axis mediates ovarian cancer metastasis both in vivo and in vitro.

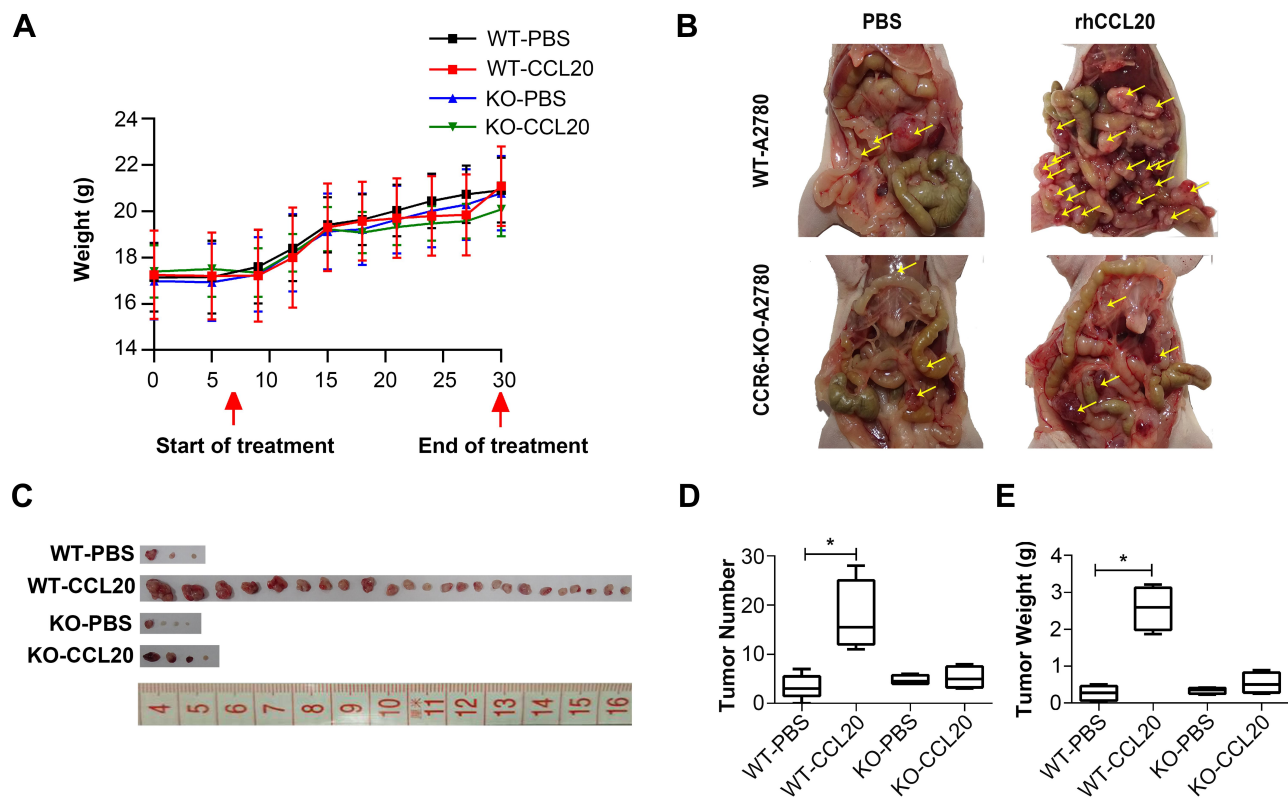


Figure 5 CCL20-CCR6 axis promotes ovarian cancer metastasis in vivo.

Notes: (A) Body weight of nude mice throughout the experiment. (B) The abdominal metastasis of ovarian cancer in nude mice. (C) Tumors dissected out from the abdominal cavity of nude mice. (D) The number of tumors dissected out from the abdominal cavity of nude mice. (E) The weight of tumors dissected out from the abdominal cavity of nude mice. *Compared to WT-PBS group, $P < 0.05$; Yellow arrows, the abdominal metastatic ovarian tumors.

Abbreviations: WT, wild type; KO, knock-out; rhCCL20, recombinant human CCL20 protein.

Recent studies have shown that the CCL20-CCR6 axis plays an important role in cancer progression. The expression of CCR6 is significantly up-regulated in various tumor tissues or tumor cells, including laryngeal cancer, thyroid cancer, colon cancer, gastric cancer and liver cancer.^{33,34,39-41} In colon cancer, CCR6 expression is closely related to lymph node status and distant metastasis: CCR6 expression is higher in lymph node positive tumor tissues and the highest in metastatic cases.³³ In gastric cancer, high expression of CCR6 is significantly correlated with cancer relapse and poor overall survival.⁴¹ CCL20, the ligand of CCR6, can directly regulate the malignant biological activities of tumor cells. Studies have shown that the CCL20-CCR6 axis promotes cancer cell proliferation, invasion and/or migration, probably by promoting AKT,^{42,43} MAPK,⁴³ PI3K or NF κ B signaling pathways,^{39,42,43} or by promoting the expression of EMT-related proteins or matrix metalloproteinases (MMPs).^{33,34,40} In our study, the RNA-sequencing analysis demonstrated that rhCCL20 treatment did not cause a widespread change of gene transcriptome in ovarian

cancer cells, and the differentially expressed genes were enrichment in cell adhesion molecules. qRT-PCR confirmed that the crucial genes that mediate cell motility, cell-cell adhesion and epithelial-mesenchymal transition, including VCAN (versican) and CDH2 (N-cadherin), were greatly up-regulated after rhCCL20 treatment. In addition, the expression of CDH1 (E-cadherin), whose loss is considered the key process of epithelial-mesenchymal transition, was strikingly suppressed after rhCCL20 treatment. Furthermore, immunohistochemical analysis of tumor tissues from in vivo experiment verified that rhCCL20 significantly promoted the expression of N-cadherin and versican in metastatic tumors, and inhibited the expression of E-cadherin, while knockout of CCR6 successfully blocked the expression changes induced by rhCCL20. Collectively, the result suggests that CCL20 could promote the expression of EMT markers in ovarian cancer cells.

It is well known that EMT is a crucial mechanism of tumor metastasis. EMT is a reversible dynamic process that allows epithelial cells temporarily shedding their epithelial

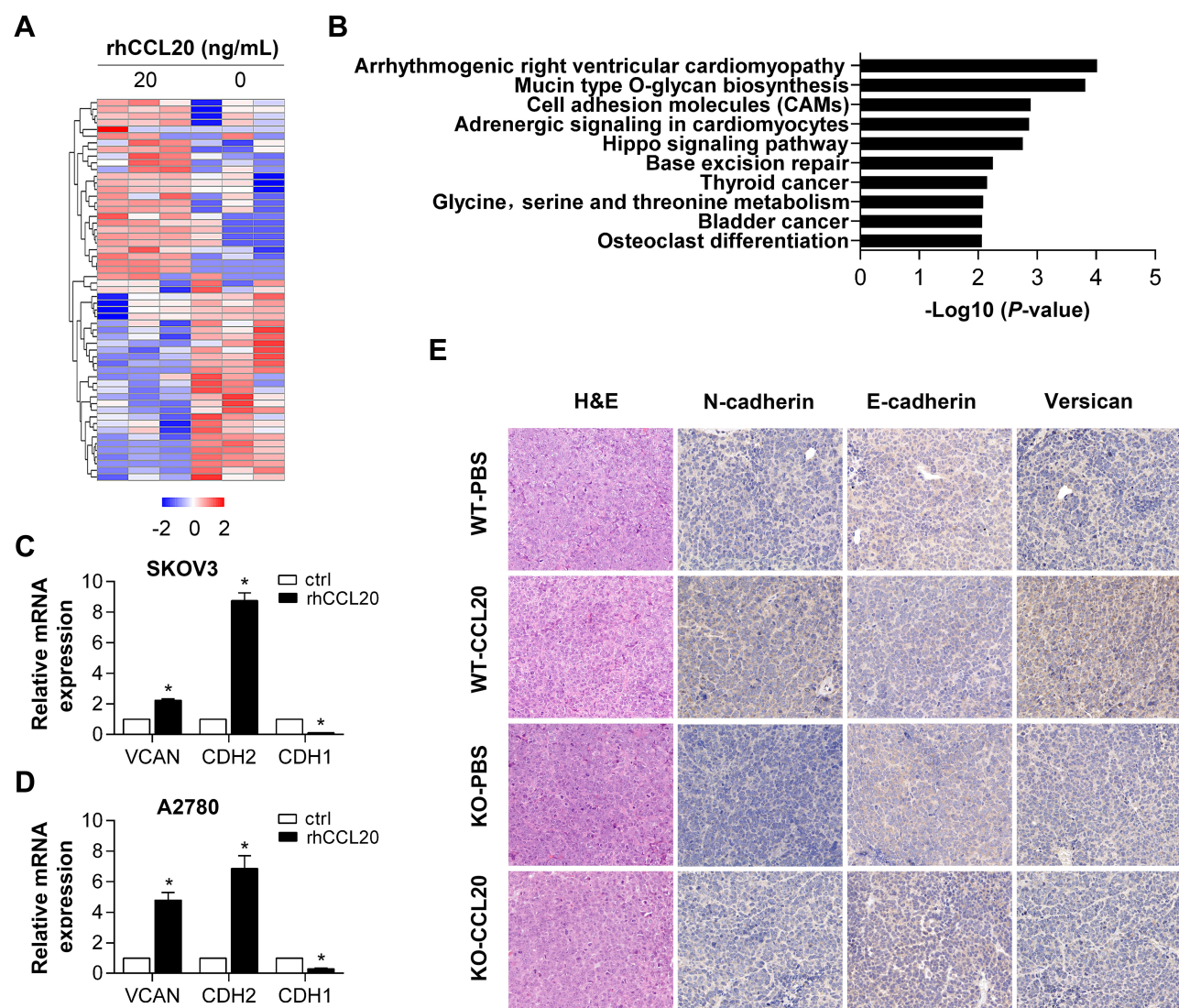


Figure 6 Differentially expressed genes after rhCCL20 treatment in ovarian cancer cells.

Notes: (A) Heatmap of differentially expressed genes after rhCCL20 treatment in SKOV3 cells for 24 hours. (B) KEGG analysis of differentially expressed genes after rhCCL20 treatment. (C, D) qRT-PCR verification of the expression of VCAN, CDH1 and CDH2 in SKOV3 and A2780 cells after rhCCL20 treatment. (E) H&E staining and IHC staining of N-cadherin, E-cadherin and versican on tumor tissues from in vivo experiment. *Compared to CTRL group, $P < 0.05$

Abbreviations: KEGG, Kyoto Encyclopedia of Genes and Genomes; CTRL, control; H&E, hematoxylin and eosin; IHC, immunohistochemical; rhCCL20, recombinant human CCL20 protein.

cell phenotype and acquiring the mesenchymal cell phenotype, which is characterized by upregulation of mesenchymal markers and molecules that induce EMT, and downregulation of epithelial markers.⁴⁴ The expression changes of EMT-related genes could lead to enhanced migration and invasion capacity, increased resistance to apoptosis, and a remarkable increased production of extracellular matrix components.⁴⁵ In ovarian cancer, EMT contributes to chemotherapy resistance and stemness, making it a potential target to reverse therapy resistance. For example, blockade of type II TGF- β receptor could reverse EMT and thereby improving the response to carboplatin in metastatic ovarian cancer models.⁴⁶

In conclusion, the above results suggest that CCL20-CCR6 axis promotes the metastasis of ovarian cancer, probably through increasing cancer cell mobility and epithelial–mesenchymal transition. Combined with our previous research, CCL20-CCR6 blockade may provide a novel therapeutic strategy for ovarian cancer treatment.

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Disclosure

The authors report no conflicts of interest in this work.

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