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CASE REPORT

Identification of a Novel SLC8A1-ALK Fusion and Non-Canonical Expression Significantly Responding to ALK-TKIs in Lung Adenocarcinoma: A Case Report

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rearrangements are sensitive to treatment with tyrosine kinase inhibitors (TKIs). However, different types of ALK fusion influence the clinical outcomes of this therapeutic approach. Approximately 10–40% of patients with ALK-fusion positive non-small cell lung cancer do not response to ALK-TKI therapy. Therefore, it is important to accurately identify the types of ALK rearrangement for appropriate selection of clinical treatment.
 Case Report: Using a DNA-targeted next-generation sequencing technique, we found a novel solute carrier family 8 member A1 (SLC8A1)-ALK fusion type in a patient with hung advancement for approximate selection of clinical treatment.

a nover solute carrier failing a memoer AT (SECSAT)-AEK fusion type in a patient with lung adenocarcinoma. Further reverse transcriptase-polymerase chain reaction and Sanger sequencing demonstrated the rearrangement as a B-cell CLL/lymphoma 11A (BCL11A)-ALK fusion at the transcriptional level. The patient showed a rapid and strong response to treatment with crizotinib, which lasted for 9 months. The patient also responded well to treatment with alectinib after developed resistance to crizotinib.

Background: Approximately 2-7% of patients with non-small cell lung cancer harbor

anaplastic lymphoma kinase (ALK) rearrangement events. Of note, typical ALK actionable

Conclusion: A strategy combining DNA-targeted next-generation sequencing with RNA reverse transcriptase-polymerase chain reaction and sequencing, besides fluorescence in situ hybridization and immunohistochemistry, may provide an effective and practical solution for correct identification of partner genes and fusion structures in the diagnosis of ALK rearrangements, particularly for non-canonical expression patterns of ALK fusion events. The combined approach may lead to more benefits for patients.

Keywords: lung adenocarcinoma, anaplastic lymphoma kinase rearrangement, tyrosine kinase inhibitor, treatment, case

Introduction

In non-small cell lung cancer (NSCLC), approximately 2–7% of patients harbor anaplastic lymphoma kinase (ALK) gene rearrangement events. These events often result in aberrant expression and oncogenic activation of ALK driven by the promotion of fusion-partner genes.^{1,2} With the development of targeted therapy using tyrosine kinase inhibitors (TKIs), ALK-TKIs (eg, alectinib and crizotinib) have markedly improved the clinical outcomes of patients with advanced ALK fusion-positive NSCLC.^{3–7} Thus far, >90 ALK rearrangement types have been found.⁸ In addition, approximately half of ALK fusions are non-canonical or complex, which have been

Received: 20 June 2021 Accepted: 9 September 2021 Published: 27 September 2021 © 2021 Zhu et al. This work is published and licensed by Dove Medical Press Limited. The full terms of this license are available at https://www.dovepress.com/terms.php you hereby accept the Terms. Non-commercial uses of the work are permitted without any further permission form Dove Medical Press Limited, provided the work is properly attributed. For permission for commercial use of this work, please apargraphs 4.2 and 5 of our Terms (http://www.dovepress.com/terms.php). associated with poor clinical response to ALK-TKI therapy, accounting for 10-40% of ALK rearrangement events.9 Fusions that upregulate the expression of the entire ALK domain, such as rearrangements with EMAP like 4 (EML4), nucleophosmin (NPM), tropomyosin 3 (TPM3), trafficking from ER to golgi regulator (TFG), 5-aminoimidazole-4-carboxamide ribonucleotide formyltransferase/IMP cyclohydrolase (ATIC), clathrin heavy chain (CLTC), etc, have been linked to good responses to ALK-TKI therapy.3-5,8,9 Thus, the clinical outcomes of therapy with ALK-TKIs vary depending on the type of fusion. Therefore, it is necessary to accurately detect ALK rearrangement events for proper selection of clinical treatment. In this article, we report a rare solute carrier family 8 member A1 (SLC8A1)-ALK fusion type identified using DNA targeted next-generation sequencing (NGS) and further examined the expression of the entire ALK domain on RNA levels in a patient with lung adenocarcinoma; we also report on the follow-up of treatment with ALK-TKIs.

Case Presentation

A 62-year-old Chinese man without smoking history was admitted to Sir Run Run Shaw Hospital in September 2019 because of cough with phlegm lasting for 2 months and presence of blood in the phlegm. On physical examination, there was no dysmorphic features. Chest enhanced computed tomography (CT) scan showed a mass at the right lower lobe, multiple nodular thickening with right horizontal fissures, and local atelectasis in the middle and lower lobes of the right lung, accompanied by bilateral pleural effusion and inflammatory lesions (more severe on the right side) and plump mediastinal lymph-nodes. Further examination did not detect metastatic lesions. Subsequently, CT-guided biopsy of the lung tumor tissue was performed. Pathological examination showed NSCLC, and the clinical analysis indicated stage IVA (cT3N2M1a). The immunohistochemical analysis revealed positive expression for napsin A, Ki-67, thyroid transcription factor 1 (TTF-1), and negative expression for P40 (Figure 1), indicating an adenocarcinoma origin of the lung tumor.

To determine the possible presence of somatic mutations suitable for targeted therapy, DNA-level NGS was also performed on the biopsy specimen based on a 10-gene panel (including ALK, BRAF, EGFR, ERBB2, KRAS, MET, PIK3CA, RET, ROS1 and TP53, Berry Oncology Corporation, Beijing, China). A novel solute carrier family 8 member A1 (SLC8A1)-ALK fusion type was identified with a mutant allele frequency of 5.98%. The sequencing results showed that the SLC8A1 exon 2 rearranged with ALK exon 19 (Figure 2A, B and D). The breakpoints and corresponding fusion sequences were further examined using reverse transcriptase-polymerase chain reaction (RT-PCR), followed by Sanger sequencing to confirm its expression pattern on RNA levels. Of note, the results of targeted RT-PCR sequencing revealed a different ALK rearrangement partner gene: a B-cell CLL/lymphoma 11A (BCL11A) exon 3 rearranged with ALK exon 20 (Figure 2A, C and E). The transcript fusion type retained the N-terminal part of BCL11A and the entire kinase



Figure I Pathological examination of the patient tumor tissue. Hematoxylin and eosin (HE) staining of the biopsy specimen showed a poorly differentiated tumor tissue. IHC experiments showed positive expressions of Napsin A, TTF-1, Ki-67 and negative expression of P40 (x200), revealing an adenocarcinoma origin of the lung cancer.



Figure 2 Diagrams showed breaking point of SLC8A1-ALK fusion and BCL11A-ALK fusion. (A) Positions of genes ALK, SLC8A1 and BCL11A in the chromosome 2 (Chr2). (B) The breaking point of partial SLC8A1 exon2 (blue color) fused to the last 54 bp of ALK exon 19 (orange color) detected on DNA levels. (C) The breaking point of partial BCL11A exon3 (purple color) fused to the first 1 bp of ALK exon 20 detected. (D and E) Diagrams showed the sequence around the breaking point of SLC8A1-ALK fusion and BCL11A-ALK fusion, respectively.

domain of ALK, which may lead to transformation of constitutive kinase activity and carcinogenesis.

Fluorescence in situ hybridization was performed to confirm the ALK rearrangement events on a chromosomal level in the biopsy specimen obtained from the patient (Figure 3A), with typical positive ALKrearrangement signals in comparison with the positive reference EML4-ALK fusion image (Figure 3B). Apart from the aforementioned ALK fusion, there was no epidermal growth factor receptor (*EGFR*), *TP53* and *KRAS* mutations or *ROS1* rearrangement detected in this patient.

The patient was treated with crizotinib (dose: 250 mg, twice daily) from October 16, 2019. Chest CT scan images demonstrated that the mass, plump mediastinal lymph



Figure 3 FISH images. (A) FISH analysis showed fused red-green signals (negative signal), split red-green signals (positive signal) and single red signals (positive signal) in the patient's biopsy specimen. (B) FISH analysis of typical EML4-ALK fusion gene expression, as the reference. The red arrows indicate cells with positive signals of ALK-fusion events and the green arrows indicate negative signals of non-ALK fusion. Both images were magnified 100 time (x100) under microscope.

nodes and pleural effusion had obviously resolved at 1 month and reached stable disease status at 3 months after the treatment (Figure 4). However, a chest CT scan (July 4, 2020) revealed an increased solid composition of the mass, multiple thickened nodules with right horizontal fissures,

and plum per mediastinal lymph-nodes appeared in comparison with previous CT scan images (January 11, 2020) (Figure 4). We considered that the patient developed progressive disease during the treatment with crizotinib, and the progression-free survival was 9 months.



Figure 4 Thoracic computed tomography (CT) images showed tumor responses to ALK-TKIs treatment before I month of therapy (baseline) and $1\sim3$ months after therapy. The CT scans demonstrated that a mass in the right lower lobe, plump mediastinal lymph nodes and pleural effusion of the right chest was obviously resolved I month after the crizotinib treatment. In 9 months, CT showed solid composition of mass increased (red arrow) and the tumor responded to alectinib treatment again in I3 and I7 months.

Abbreviations: PR, partial response; SD, stable disease; PD, progressive disease; bid, twice a day.

Thenceforward, alectinib was administered (dose: 600 mg, twice daily) from July 4, 2020. A chest CT scan performed after 4 and 8 months showed partial response. The patient remains under surveillance up to now.

Discussion

To the best of our knowledge, this is the first reported case of the SLC8A1 exon 2-ALK exon 19 fusion type. Fluorescence in situ hybridization analysis also confirmed the positive ALK-rearrangement signals on DNA levels. However, the response of SLC8A1-ALK fusion to treatment with ALK-TKIs remains unknown. The response of TKI treatments for ALK-rearrangement patients largely depends on whether the ALK domain is expressed entirely and at high levels driven by the rearranged-partner genes, which are influenced by complex rearrangement structures and RNA splicing mechanisms, and sometimes disaccord on DNA levels and RNA levels.9 Therefore, RT-PCR and Sanger sequencing were performed to validate the transcription pattern of the fusion gene and confirm the sequences around the breaking point. Notably, the RT-PCR sequencing demonstrated a different type of ALK fusion transcript (ie BCL11A exon 3-ALK exon 20 fusion) in this patient (Figure 3). Fortunately, this ALK-fusion expression pattern is similar to that of BCL11A exon 4-ALK exon 20, which was recently reported to respond strongly to the treatment with crizotinib in NSCLC.¹⁰ This finding suggested that ALK-TKI treatment may theoretically benefit the patient of this case. Thus, we administered him crizotinib, and the response was prompt and markedly in the first 9 months (Figure 4). Following the development of resistance to crizotinib, treatment with alectinib was initiated. Thus far, this therapy remains effective, suggesting that alectinib may be an alternative option in patients with this ALK-fusion type after development of resistance to crizotinib.

The exact mechanism through which the ALK fusion participates in carcinogenesis remains to be determined. Nevertheless, it is accepted that the fusion gene can upregulate the expression of ALK and increase its activity, thereby activating ALK self-phosphorylation and further triggering downstream oncogenic signaling pathways.^{1,2} As reported by an increasing number of studies, the aberrant kinase activity varies depending on the structures of different ALK rearrangements.⁹ Different fusion structures influence the integrity of the kinase domain, intracellular localization and stability of the fusion protein.^{11–13} Moreover, they may lead to fusion with different partner

genes at the transcriptional levels and DNA levels mainly due to complex RNA splicing mechanisms during expression, as shown in this case. These influencing factors resulted in variable clinical responses to treatment with ALK-TKIs ranging from sensitivity to resistance.^{9,14–16} Notably, some coexistent gene mutations, such as *TP53* mutations, also impair the efficacy of ALK-TKI treatment.⁹

Conclusion

Using a targeted DNA sequencing panel, we report a NSCLC patient harboring a novel SLC8A1-ALK fusion. Further RT-PCR and Sanger sequencing demonstrated this ALK-fusion type expressed as another similarly established BCL11A-ALK fusion pattern at the transcriptional levels. The results assisted in the drug selection of ALK-TKI treatment, which eventually benefited the patient. Therefore, a strategy combining DNA-based NGS with RNA-based RT-PCR sequencing should be considered in clinical practice for the accurate diagnosis of ALK rearrangements.

Abbreviations

NSCLC, non-small cell lung cancer; ALK, anaplastic lymphoma kinase; TKIs, tyrosine kinase inhibitors; NPM, nucleophosmin; TPM3, tropomyosin 3; TFG, trafficking from ER to golgi regulator; ATIC, 5- aminoimidazole-4-carboxamide ribonucleotide formyltransferase/IMP cyclohydrolase; CLTC, clathrin heavy chain; SLC8A1, solute carrier family 8 member A1; NGS, targeted nextgeneration sequencing; CT, computed tomography; RT-PCR, reverse transcriptase-polymerase chain reaction; TTF-1, thyroid transcription factor 1; BCL11A, B-cell CLL/lymphoma 11A; EGFR, epidermal growth factor receptor.

Data Sharing Statement

The data and materials in this case are available from the corresponding author on reasonable request.

Ethics

Written informed consent was obtained from the patient for publication of this case report and any accompanying images. This study protocol was reviewed and the need for approval was waived by Ethics Committee of Sir Run Run Shaw Hospital.

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Disclosure

Yin Wang, Xiaoxing Su, Yan Lei and Shuangxiu Wu are employees of Berry Oncology Corporation. The authors report no other conflicts of interest in this work.

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