ORIGINAL RESEARCH

Effect of variable circuit flow rate during the expiratory phase on CO₂ elimination

Peter A Keszler¹ Pankaj Nagaraj¹ Kabir Abubakar¹ Martin Keszler²

¹Georgetown University, Washington, DC, USA; Georgetown University Hospital, Washington, DC, USA; ²Brown University, Women and Infants Hospital of Rhode Island, Providence, RI, USA

Correspondence: Martin Keszler Women and Infants Hospital of Rhode Island, 101 Dudley Street, Providence, RI 02905, USA Tel +1 401 274 1122 Fax +1 401 453 7571 Email mkeszler@wihri.org **Background:** Some continuous flow infant ventilators allow independent setting of inspiratory and expiratory circuit flow rate. In the Dräger Babylog 8000+ ventilator, this is called "variable inspiratory, variable expiratory flow" (VIVE). Some clinicians believe that lower expiratory flow decreases expiratory resistance. The minimum expiratory flow rate needed to avoid re-breathing of carbon dioxide (CO_2) has never been established.

Objective: We sought to determine if re-breathing becomes evident at the lowest possible expiratory flow rate setting of 1 L/min.

Design/methods: We conducted a bench study using end-tidal CO₂ (ETCO₂) measurement and a 45 mL (90 mL for the "term" model) test lung pre-filled with 100% CO₂. We previously showed that the time needed for ETCO₂ to be eliminated from the lung is a highly reproducible indicator of efficiency of ventilation. Re-breathing would thus be identified by an increase in the time required for the CO₂ to be washed out from the test lung at stable settings of rate and tidal volume (V_T). Using a Babylog 8000+ ventilator in volume guarantee mode with VIVE and a standard ventilator circuit, we tested the effect of decreasing expiratory flow rate under conditions simulating three sizes of patients: extremely low birth weight infant, wt = 600 g (V_T = 3.5 mL, respiratory rate (RR) = 60 breaths min⁻¹, minute ventilation (MV) = 210 mL/min, expiratory flow rate = 3 L/min, 2 L/min, and 1 L/min), very low birth weight infant, wt = 1.5 kg (V_T = 7 mL, RR = 60 breaths min⁻¹, MV = 420 mL/min, expiratory flow rate = 4 L/min, 3 L/min, 2 L/min, and 1 L/min), and term infant, wt = 3.6 kg (V_T = 16 mL, RR = 60 breaths min⁻¹, MV = 960 mL/min, expiratory flow rate = 5 L/min, 4 L/min, 3 L/min, 2 L/min, and 1 L/min). Each measurement was repeated four times and the mean values were compared by analysis of variance for repeated measures.

Results: The CO_2 elimination times were always within 10% of each other with each repetition. There was no significant increase in the elimination time even at the lowest flows.

Conclusion: Clinically significant re-breathing does not occur even with expiratory flow rate at the lowest possible setting of 1 L/min. VIVE is safe to use, although its clinical utility has not been established.

Keywords: mechanical ventilation, newborn, bench study, re-breathing, expiratory flow rate, end-tidal CO₂

Background

The majority of ventilators currently in use in North America use demand flow during the inspiratory phase to meet inspiratory flow requirements and return circuit flow to a low baseline during the expiratory phase. The Dräger Babylog 8000+ (Dräger Medical, Lübeck, Germany) is a continuous flow infant ventilator that uses a continuous flow in the ventilator circuit throughout the respiratory cycle in the default mode, but

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provides a feature that allows the user to select an expiratory flow rate independent of inspiratory flow rate. This feature is called "variable inspiratory, variable expiratory flow rate" or VIVE.^{1,2} There are no published data to guide the use of VIVE. Many clinicians believe that lower expiratory flow rate decreases expiratory resistance and thus lowers the work of breathing, based on a paper suggesting that continuous flow ventilators may be associated with higher expiratory work of breathing.3 Although this may no longer be true with modern technology, VIVE remains a commonly used option on one of the most widely used neonatal ventilators and there are no data to guide the clinician in setting appropriate expiratory flow rate in VIVE. Whether or not it affects expiratory resistance, lower circuit flow during the expiratory phase would lower the consumption of medical gases and sterile water for humidification and may therefore be desirable. However, if the circuit flow rate during the expiratory phase were to fall below that, which is needed to sweep exhaled gas down the expiratory limb of the circuit, re-breathing of CO₂ would occur and impair CO₂ elimination.

We hypothesized that there is a minimum expiratory flow rate that is needed to avoid re-breathing of exhaled gas and therefore avoid impairing CO_2 elimination and that this flow rate is related to the tidal volume (V_T) and minute ventilation (MV) of the infant.

Methods

We conducted a bench study using a Dräger Babylog 8000+ continuous flow infant ventilator (Dräger Medical, Lübeck, Germany) with a standard Fisher and Paykel ventilator circuit and humidifier (Fisher and Paykel, Auckland, New Zealand). We pre-filled a 45 mL silicone rubber bellows test lung (IngMar Medical, Pittsburgh, PA) with 100% CO₂ and used a CO₂SMO Plus respiratory monitor mainstream ETCO₂ system (Novametrix Medical Systems, Wallingford, CT) to detect the rate at which CO₂ was eliminated from the test lung (Figure 1). We previously showed that the time needed for ETCO, to fall below 8 mmHg (the threshold of reliable detection by this instrument) is a highly reproducible indicator of efficiency of ventilation in this model.⁴ The basic principle is that with the initiation of ventilation, CO₂ is detected by the ETCO, monitor at the airway opening, initially at high concentration when the test lung is filled with pure CO_2 . Subsequently, the ETCO₂ declines in an exponential fashion as the CO₂ concentration in the test lung decreases and therefore less CO₂ is cleared from the test lung with each breath (Figure 2). The time needed for the ETCO, to drop below the threshold of detection is proportional to effective alveolar minute ventilation.

Re-breathing would thus be identified by an increase in the time required for the CO₂ to be washed out from the test lung (the CO₂ elimination time) at any given MV determined by stable settings of respiratory rate (RR) and V_T. We tested the effect of decrements in expiratory flow rate under conditions simulating three sizes of patients: extremely low birth weight infant, wt = 600 g (V_T = 3.5 mL, RR = 60 breaths min⁻¹, MV = 210 mL/min, expiratory flow rate = 3 L/min, 2 L/min, and 1 L/min), very low birth weight infant, 1.5 kg (V_T = 7 mL, RR = 60 breaths min⁻¹, MV = 420 mL/min, expiratory flow rate = 4 L/min, 3 L/min, 2 L/min, and 1 L/min),



Figure I The experimental setup. Abbreviations: ET, endotracheal; ETT, endotracheal tube.



Figure 2 The smaller panel in the figure shows the ETCO₂ waveform generated with each breath. The larger panel is a compressed waveform of a single trial (note the difference in time scale). The peak of the curve is flattened, because the upper limit of detection of the capnograph is 150 mmHg, while the actual ETCO₂ value should be close to atmospheric pressure at the onset of ventilation when the lung is filled with pure CO₂. The exponential decline in the readings is evident. Initially, at high CO₂ concentrations, a larger amount of CO₂ is eliminated with each breath, but as the level of CO₂ drops, a progressively smaller quantity is removed, as indicated by progressively lower end-tidal CO₂ values. The elimination time is a reflection of the effective alveolar ventilation. **Abbreviation**: ETCO₂, end-tidal CO₂.

and term infant, wt = 3.6 kg ($V_T = 16 \text{ mL}$, RR = 60 breaths min⁻¹, MV = 960 mL/min, expiratory flow rate = 5 L/min, 4 L/min, 3 L/min, 2 L/min, and 1 L/min). For the term infant model, a double bellows lung (volume = 90 mL) was used because of the larger V_T needed.

The ventilator circuit was connected to the ETCO₂ sensor which was attached to one end of the Babylog flow sensor connected to a 2.5 mm endotracheal tube cut to 10 cm length and inserted tightly into the open port of the test lung. A 3.0 mm tube was used for the simulated 1.5 kg infant and 3.5 mm tube for the simulated term infant. The test lung connection had a side port that allowed filling of the test lung with pure CO₂ prior to each test. Prior to each set of experiments, the flow sensor and ETCO, monitor were calibrated according to manufacturers' instructions and tightness of all connections was verified. The test lung was flushed with 100% CO₂ at 2 L/min for 2 minutes just prior to each trial to ensure that it was completely filled with pure CO₂, the CO₂ port was then clamped and the test lung was attached to the ventilator circuit taking care to minimize movement that might agitate the gas. The test lung was enclosed in an airtight plastic container to avoid inadvertent change in volume (ie, "spontaneous breathing"). The ETCO₂ monitor was switched on and ventilation started in volume guarantee mode at a specific expiratory flow rate setting on VIVE. The inspiratory flow rate remained at 6 L/min; inspiratory time was set at 0.35 seconds, pressure

limit at 30 cm H_2O , and positive end-expiratory pressure at 5 cm H_2O .

Each set of experiments consisted of a series of trials simulating the three clinical situations with the appropriate V_{T} using the different flow rates described above in random order. Each series was performed four times. We made sure that the target $V_{_{\mathrm{T}}}$ was delivered and increased the pressure limit as needed if necessary. The ETCO₂ measurement was stopped when the ETCO₂ reading fell below 5 mmHg or when the capnograph stopped displaying values (typically below 6 mmHg). The elimination time was determined from the digital data exported from the capnograph as the time from onset of ventilation to the point when the ETCO, value dropped below 8 mmHg for three consecutive breaths. This was an arbitrary cutoff point made necessary by the exponential decline nature of the CO₂ elimination curve and the fact that the instrument begins to lose accuracy at very low CO₂ levels and stops reporting values below approximately 6 mmHg. All measurements were carried out by a single observer (PAK) to eliminate a preventable source of variability.

Statistical analysis

The digital data were exported from the capnograph software to an Excel spreadsheet, graphed, and visually inspected for consistency and to ensure that there were no evident distortions to the expected exponential decline. The start of

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ventilation and the end point of three consecutive end-tidal values below 8 mmHg were identified. The time difference between the two points was recorded and entered into a spreadsheet for subsequent analysis. Analysis of variance for repeated measures was used to evaluate the effect of different expiratory flows, with P values <0.05 considered significant.

Results

The measurements were highly reproducible and the elimination times were always within 10% of each other at each setting. There were no significant differences in elimination time even at the lowest setting of 1 L/min in each size simulated patient (Table 1). Larger V_T resulted in a shorter elimination time as would be anticipated, but not to as great an extent as would be predicted from the conventional assumptions about instrumental dead space; doubling of V_T from 3.5 mL to 7 mL reduced the elimination time by a lesser amount than the predicted increase in alveolar ventilation, which is consistent with our previous observations.⁴

Discussion

To our knowledge, this is the first study to investigate the impact of expiratory flow rate on CO₂ elimination in the bench or clinical setting at values relevant to neonatal patients. We found no differences in elimination time between different expiratory flow rates, including the lowest setting of 1 L/min, indicating that there was no re-breathing even in the extreme situation of the simulated term infant with total MV near 1 L/min. This finding is consistent with a lack of clinical reports of the adverse effect of VIVE and with a theoretical model prediction. The 1 L/min expiratory flow should be just sufficient to provide fresh gas to meet the simulated term infant's needs, as it provides 16.7 mL of fresh gas/second, slightly exceeding the 16 mL V_{T} used in this study at a rate of 1 breath/second. We were unable to obtain acceptable recording when we attempted to use V_T of 18 mL and 20 mL, with the ETCO, monitor consistently recording error messages despite careful checking of all connections and repeated re-calibration. This was likely because the $ETCO_2$ monitor algorithm rejected the abnormal CO_2 pattern resulting from the effects of the predicted re-breathing of exhaled gas that should occur when the V_T exceeded available fresh gas supply.

The V_{T} we used for the simulated 1.5 kg and term patients were approximately 4.5 mL/kg, based on previously published values for $V_{\rm \scriptscriptstyle T}$ used with volume-targeted ventilation measured at the airway opening, which have been shown to be associated with normocapnia. $^{\rm 5-8}$ We used a larger $V_{\rm _T}$ of 5.8 mL/kg for the smallest infant, because, as we previously demonstrated, the tiniest infant requires proportionately larger V_{T} to compensate for the fixed instrumental dead space, which becomes proportionately larger in the smallest infant.9 The elimination times we found reflected the size of the V_T in a manner consistent with our previous work in which we demonstrated that effective alveolar ventilation occurs with $V_{\rm T}$ at or below the theoretical dead space.⁴ For this study, we reconfigured the setup slightly, which resulted in a slightly smaller instrumental dead space than in our previous study, accounting for the modestly shorter CO₂ elimination time recorded in this series of experiments, compared to the original study with V_{T} of 3.5 mL.⁴ For these experiments, we deliberately chose a $\boldsymbol{V}_{_{T}}$ that would lead to a relatively long CO, elimination time, so that we could detect any effect of re-breathing more easily. The remarkable reproducibility of the CO₂ elimination times within each series validates the accuracy of this technique in assessing the effective alveolar ventilation.

Our study has some important limitations. We used a simple lung model which is inherently different from a biological lung. The lung model is a passive device, not engaged in CO_2 production or gas exchange, or the variable breathing patterns that are seen in preterm infants. Although this may be seen as a limitation, it actually represents an important advantage by eliminating these potential confounders that could obscure the effect of expiratory flow rate on CO_2 removal. We were not able to examine the largest possible V_{T} , which would likely have resulted

Table I CO₂ elimination time for the three different "size" patients

	Expiratory flow rate				
	I L/min	2 L/min	3 L/min	4 L/min	5 L/min
$V_{T} = 3.5 \text{ mL}$	163.8±11.3	160.0 ± 5.0	161.0 ± 6.3		
$V_{T} = 7.0 \text{ mL}$	60.5 ± 4.2	$\textbf{61.8} \pm \textbf{4.8}$	61.8 ± 1.5	63.0 ± 3.2	
$V_{T} = 16 \text{ mL}^{*}$	$\textbf{48.5} \pm \textbf{5.0}$	47.5 ± 3.7	49.3 ± 3.3	47.5 ± 3.0	48.3 ± 3.3

Notes: Data are presented as mean \pm SD. *Double bellows lung with 90 mL volume. Abbreviation: $V_{_{TT}}$ tidal volume.

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in re-breathing, as the presumed re-breathing rendered the $ETCO_2$ instrument incapable of generating a valid readout based on its signal rejection algorithm. While this conclusion remains speculative, the fact that the failed recordings reproducibly occurred exactly at the point at which re-breathing would be predicted to occur serves as indirect evidence to support the original hypothesis.

Conclusion

Clinically significant re-breathing will not occur even with expiratory flow rate at the lowest possible setting of 1 L/min in all but the largest term infant. VIVE is safe to use even at very low expiratory flows, although its clinical utility has not been established.

Disclosure

M Keszler has been a consultant to Dräger Medical, the manufacturer of the device used in these studies. He does not have any equity interest in the company and no one related to the company has had any input into the design, execution, or reporting of the study. No financial or equipment support was provided by Dräger Medical for this study.

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